Radioactive Marker (for RT-PCR)

1. Mix
   - 3 µl ddH_2O
   - 1 µl DNA (*Msp* I digest) 1 µg/µl
   - 1 µl NEB Buffer 2
   - 2 µl dATP, dGTP, dTTP (2.5 mM)
   - 1 µl Klenow (3' fill, 1/2 u/µl)
   - 2 µl α ^{32}P dCTP

2. Incubate 10 minutes at 37°C

3. Add 1 µl glycogen, 189 µl ddH_2O, 200 µl phenol/chloroform. Spin 2 minutes, max speed, 4°C.

4. Transfer top phase to fresh tube, add 600 µl 100% EtOH, 100 µl 5M NH_4OAc. Spin 10 minutes, max speed, 4°C. Look for pellet, and carefully pipette off ethanol and discard.

5. Wash pellet in 200 µl 70% EtOH. Spin 2 minutes, max speed, 4°C. Carefully pipette off ethanol and allow pellet to air dry.

6. Resuspend pellet in 50 µl ddH_2O, and add 150 µl of formamide buffer.

7. For use during the first week, make a 1:10 dilution, or to desired intensity.