Impaired periamygdaloid-cortex prodynorphin is characteristic of opiate addiction and depression

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Negative affect is critical for conferring vulnerability to opiate addiction as reflected by the high comorbidity of opiate abuse with major depressive disorder (MDD). Rodent models implicate amygdala prodynorphin (Pdyn) as a mediator of negative affect; however, evidence of PDYN involvement in human negative affect is limited. Here, we found reduced PDYN mRNA expression in the postmortem human amygdala nucleus of the periamygdaloid cortex (PAC) in both heroin abusers and MDD subjects. Similar to humans, rats that chronically self-administered heroin had reduced Pdyn mRNA expression in the PAC at a time point associated with a negative affective state. Using the in vivo functional imaging technology DREAMM (DREADD-assisted metabolic mapping, where DREADD indicates designer receptors exclusively activated by designer drugs), we found that selective inhibition of Pdyn-expressing neurons in the rat PAC increased metabolic activity in the extended amygdala, which is a key substrate of the extrahypothalamic brain stress system. In parallel, PAC-specific Pdyn inhibition provoked negative affect–related physiological and behavioral changes. Altogether, our translational study supports a functional role for impaired Pdyn in the PAC in opiate abuse through activation of the stress and negative affect neurocircuitry implicated in addiction vulnerability.

Introduction

Chronic negative emotional states are a compelling motivation for relapse to most drugs of abuse, especially opiates (1). The cluster of symptoms that make up these negative emotional states include loss of motivation for natural reward, dysphoria, anhedonia, and anxiety (1, 2). Persistent negative affect is also a cardinal feature of major depressive disorder (MDD), which is the most common comorbid psychiatric diagnosis with opiate addiction (3). The high comorbidity rate of these disorders has been hypothesized to be due to overlapping neurobiological abnormalities in neural circuits regulating emotion (4, 5).

In accordance with this hypothesis, human imaging studies of drug addiction and MDD patients converge on structural and functional disturbances in limbic neurocircuitry such as the amygdala (6–10). The amygdaloid complex comprises several heterogeneous nuclei that mediate emotional responses to fear, dysphoria, and activation of the brain’s stress system (11–13). Accordingly, neurotransmitters within the amygdala may underlie the neuropathology of MDD and addiction disorders and are target candidates for understanding the neurobiological basis of negative affect to opiate addiction.

Across species, the amygdala displays a dense expression of the endogenous opioid neuropeptide precursor prodynorphin (PDYN) (14), highly implicated in negative mood states (13, 15, 16). Preclinical animal studies support that PDYN disturbances are characteristic of negative affect states in drug self-administration, stress exposure, and depression-like behaviors (17–20). Specifically within the amygdala, PDYN expression is altered by acute and chronic administration of heroin (18–20). Although such animal studies point to amygdala PDYN as a relevant factor in negative affect– and opiate disorder–related phenotypes, very few studies have investigated PDYN mRNA expression in the human amygdala related to neuropsychiatric disorders. Therefore, our goal was to investigate whether amygdala PDYN expression was abnormal in opiate abuse subjects and was a shared neuropathological feature of MDD. The current results highlighted a common PDYN impairment within a virtually unexplored sub-region of the amygdala, the periamygdaloid cortex (PAC) nucleus. This unusual finding prompted us to utilize a rat behavioral model to interrogate the in vivo functional connectivity of PAC-PDYN neurons using a novel molecular imaging technique.

Results

Heroin abusers and MDD subjects share a reduction of PDYN in the PAC. We assessed molecular alterations of amygdala PDYN gene expression in human opiate abusers. PDYN mRNA expression was evaluated using in situ hybridization histochemistry (ISHH) on amygdala from 2 postmortem heroin abuse cohorts: (a) an ethnically mixed cohort from the US (ref. 21) and Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI70395S1; and (b) a white Hungarian population (ref. 21 and Supplemental Table 2). Congruent with previous

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OF PAC-PDYN mRNA levels in heroin cohorts, MDD subjects had a significant downregulation (Supplemental Table 3). Similar to those in both heroin cohort II (Supplemental Figure 1). Similarly a second population of heroin abusers in cohort II displayed a specific reduction of PDYN mRNA expression in the PAC (Figure 1B; \( n_{\text{Control}} = 18, n_{\text{Heroin}} = 28; P = 0.037 \); Supplemental Figure 2). Further, no significant correlation was found between heroin metabolites and PDYN-PAC expression in either heroin cohort, suggesting that the PDYN changes were likely due to chronic heroin abuse.

Next, we examined whether amygdala PDYN disturbances were evident in an MDD population of the same ethnic makeup as the heroin population (Figure 1A; \( n_{\text{Control}} = 7, n_{\text{Heroin}} = 8; P = 0.045 \)). No other amygdala nuclei assessed showed any difference (Supplemental Figure 1). Similarly a second population of heroin abusers in cohort II displayed a specific reduction of PDYN mRNA expression in the PAC (Figure 1B; \( n_{\text{Control}} = 18, n_{\text{Heroin}} = 28; P = 0.037 \); Supplemental Figure 2). Further, no significant correlation was found between heroin metabolites and PDYN-PAC expression in either heroin cohort, suggesting that the PDYN changes were likely due to chronic heroin abuse.

Reduced Pdyn mRNA in the PAC, and concomitant sensitization of neural stress marker induced in rats with chronic heroin self-administration. The shared alterations of PDYN mRNA levels in heroin abusers and MDD subjects brought attention to the PAC. However, information about this subnucleus was limited, thus, warranting studies in animal models to help address the limitations of human postmortem studies such as incomplete knowledge of drug use history and time of death following drug use. To this end, we used the well-validated model of heroin self-administration in rats (23). First, we found that there was detectable Pdyn mRNA expression in the analogous structure of the PAC in rats (Figure 2A). Next we used the operant paradigm of heroin self-administration in which animals robustly self-administered heroin (30 μg/kg/infusion on a designated ‘active’ lever), but not saline (Figure 2B; \( n = 6–7 \) per group ANOVA, \( P < 0.001 \)). After 10 days of chronic heroin self-administration, animals were euthanized 24 hours after last drug use, a time point characterized by negative affective state and stress activation associated with heroin withdrawal and drug seeking (24). Figure 2C shows that Pdyn mRNA expression was reduced in the PAC at 24 hours after the last heroin session (\( P = 0.031, n = 5–6 \)).

In order to verify that animals were in a heightened stress state 24 hours after last heroin intake, we examined the expression of the brain stress marker corticotropin-releasing factor (Crf) within the central amygdala (CeA), an extrahypothalamic brain stress site. Increased mRNA expression of CeA Crf, has been correlated with the aversive motivational properties of opiate withdrawal (25) and was indeed apparent at the 24-hour withdrawal period (\( P = 0.045 \); Figure 2D; \( n = 5–7 \)).

**DREAMM in vivo behavioral imaging reveals a direct link of inhibition of PAC-Pdyn neurons with hyperactivation of the extended amygdala.** Due to the unknown functional connectivity of PAC-Pdyn neurons in relation to the complex heterogeneous organization of the amygdala, we used a novel strategy to identify in vivo functional brain circuits associated with modulation of PAC-Pdyn neuronal activity (Supplemental Methods). This technique, entitled DREAMM, integrates viral-mediated designer receptors exclusively activated by designer drugs (DREADD) (26) and metabolic mapping using a direct marker of brain metabolism, \( [^{18}F] \) Fluorodeoxyglucose (FDG) and μPET imaging. (27).

Rats were unilaterally infused in the right PAC with a neuronal Pdyn promoter-specific DREADD viral construct that encodes expression of a G\(_i\)-coupled receptor (hM4Di) solely activated by the otherwise inert pharmacological substance, clozapine-n-oxide (CNO) (Supplemental Figure 4) (26, 28). CNO binding to the hM4Di-Pdyn DREADD receptor activates G\(_i\)-mediated signaling, leading to a transient inhibition of Pdyn neuronal firing (26, 28). FDG uptake occurred in behaving animals, and rats were scanned in 2 states: following vehicle or CNO. As compared with the vehicle state, CNO administration and subsequent reduction of PAC-Pdyn neuronal activity led to a strong increase (\( P < 0.05 \)) of FDG uptake in the ipsilateral (right) macrostructure known as the extended amygdala (ExA)
The ExA as a circuit associated with negative affect and stress that consists of the medial amygdala (MeA), CeA, bed nucleus stria terminais (BNST), and nucleus accumbens (NAc) shell (Figure 3B; Supplemental Figure 5) (29).

To validate the specificity of this global ExA activation, we quantified individual responses based on voxel intensity in the discrete substructures of the ExA, including the BNST, CeA, MeA, and NAc shell (Figure 4A). Administration of CNO versus vehicle significantly increased voxel intensity within the majority of the right ExA components with a trend in the CeA, ipsilateral to the site of injection (BNST, \( P = 0.002 \); right NAc shell, \( P = 0.031 \); MeA, \( P = 0.008 \); CeA, \( P = 0.1 \); Figure 4B). The percentage increases (3%–10%) in voxel intensity of FDG uptake are within the range associated with physiological and clinical changes in humans (30–32) and with lesions of the substantia nigra in rodents (33).

Inhibitory modulation of PAC-Pdyn neurons induces depression-related behavioral and physiological phenotypes. The ExA as a circuit and its individual components are strongly implicated in aversive state and serves as an interface between stress and addiction (1). As such, we next evaluated behavioral and physiological parameters of stress in PAC-hM4D-iPdyn animals. First, we found that CNO administration in PAC-hM4D-Pdyn rats led to significantly elevated corticosterone levels as compared with vehicle administration in the same animal (\( P = 0.014 \); Figure 5A) and this effect was not attributable to the order of CNO administration, as the animals were counterbalanced (\( P = 0.93 \)). No effect of CNO was observed in control PAC-GFP-Pdyn animals (Supplemental Figure 6). Hedonic state was examined using the sucrose preference task in animals expressing either PAC-hM4D-Pdyn or PAC-GFP-Pdyn (control). CNO administration led to a decrease in sucrose consumption in PAC-hM4D-Pdyn animals as compared with control GFP-Pdyn (\( P = 0.032 \); Figure 5B). Last, depressive-like phenotype was assessed using the forced swim test, a commonly used depression model of behavioral despair (34). Rats expressing either PAC-hM4D-Pdyn or PAC-GFP-Pdyn were exposed to a 15-minute swim pretest 24 hours before the test. On test day, animals were evaluated for 5 minutes and time spent immobile was scored. PAC-hM4D-Pdyn animals displayed significantly higher immobility as compared with control rats, which was indicative of increased depression-like behavior (\( n = 7 \) group, \( P = 0.003 \); Figure 5C). This impairment was not due to a gross motor deficit, as there was no significant difference between hM4D-Pdyn and PAC-GFP-Pdyn animals in overall locomotor behavior assessed in an open field (Supplemental Figure 7).

**Discussion**

Using a translational approach, we have shown that impaired PDYN mRNA expression in the human PAC is a shared characteristic of the diagnostically distinct but symptomatically related disorders of opiate addiction and MDD. The consistent disturbance of PAC-PDYN expression is intriguing but raises questions as to the potential functional relevance of this impairment, since the PAC, unlike other amygdala nuclei, has not been extensively studied. As such, we expanded insights about the PAC-Pdyn neurons by examining their in vivo functional connectivity. Our use of the molecular in vivo imaging strategy DREAMM in rats highlighted the selective recruitment of the ExA in relation to impaired PAC-
Pdyn neuronal activity, therefore linking the Pdyn in the PAC to a functional neural network that regulates negative affect responses relevant to opiate addiction vulnerability.

Our finding of an association between the Pdyn system and negative affect is in line with a large body of evidence. While much of the published data have been obtained from experimental animal studies (15, 16, 35), recent clinical investigations have also demonstrated that variants of the PDYN gene relate to transcriptional alterations of PDYN mRNA expression and are associated with depressive traits and negative craving in alcoholics (36, 37). The current study adds to the field by directly examining the human brain to provide neuropathophysiological insights that have not been previously reported. The shared reduction of PDYN mRNA expression in the amygdala PAC of human heroin and MDD subjects supports the hypothesis that common disturbances in brain areas linked to emotional regulation underlie the high comorbidity rates of drug addiction with MDD. Although human postmortem studies have many challenges, the PDYN mRNA impairments observed were replicated in multiple cohorts of opiate abusers and disturbances detected in the MDD subjects also matched those previously reported (38). In addition, unlike the PAC, where PDYN expression was similarly reduced in addiction and MDD subjects, PDYN mRNA is differentially affected in the striatum of presumably depressed suicide subjects (39) and heroin abusers (40).

The prevailing hypothesis in the field posits that increased dynorphin mediates negative affect. While the reduction of PDYN mRNA expression in the striatum that reduced PDYN mRNA expression can occur concomitant to elevated dynorphin levels in some heroin abusers (40). Future insights about the release of PDYN-derived peptides from PAC-Pdyn neurons will be very informative. It is also important to emphasize that the relationship between PDYN and different aspects of negative affect is complex and still requires further investigation. For example, although stress-induced aversion is established to be mediated by elevated dynorphin (41, 42), a number of investigations have documented that animals lacking the Pdyn gene have enhanced anxiety behavior (43, 44). Moreover, our study underscores the heterogeneity of PDYN expression even within discrete brain regions, suggesting different PDYN circuits could contribute to distinct behavioral components relevant to negative affect.

A notable finding in the present study was that MDD subjects, in addition to the PAC, also had reduced PDYN mRNA expression in the AB and AHA nuclei. These disturbances were not, however, evident in the lateral nucleus (Supplemental Figure 3), which also expresses PDYN, suggesting that the alterations in MDD individuals are not global but instead discretely localized. Further studies are needed to delineate the specific contribution of PDYN AB and AHA neurons to distinct components of MDD neuropathophysiology.

Direct molecular studies of the human brain are of marked importance, but some limitations of human postmortem investigations are the lack of complete knowledge about the subjects’ drug abuse and other confounding factors, including nicotine consumption, medical history, and time of last drug use prior to death, which are relevant for interpretation of the data. However, interestingly, a reduction of PAC-Pdyn mRNA expression was also found in rats that self-administered heroin at a time point associated with acute withdrawal related
to hyperactivation of brain stress systems (upregulation of *Crf* expression in the CeA). Indeed, extracelluar and mRNA *Crf* changes in the CeA are consistently associated with acute withdrawal from opiates (25, 35, 45). Our human data also underscore the importance of the PAC-PDYN alterations related to chronic neuroadaptations, given that acute toxicology of heroin metabolites did not correlate with amygdala PDYN expression. Altogether, these observations are in line with PAC-PDYN impairments underlying negative affect that may contribute to the motivational drive for drug-seeking and relapse behavior and that have been associated with a decrease in the function of neuronal reward circuits that contributes to addiction, which Koob et al. have characterized as the “dark side” of addiction (46).

Our use of the innovative molecular and in vivo imaging strategy of DREAMM in rats also clearly highlighted the striking selective association of PAC-Pdyn neurons with the ExA that serves as an interface between drug abuse and negative affect (13, 47). Moreover, the fact that impaired firing of *Pdyn* neurons directly contribute to the motivational drive for drug-seeking and relapse behavior and that have been associated with a decrease in the function of neuronal reward circuits that contributes to addiction, which Koob et al. have characterized as the “dark side” of addiction (46).

Recent imaging studies in humans and primates have expanded the concept of the ExA in higher order species and support a role for its dysfunction in responses to antidepressants (48, 49). As such, the activation of the ExA due to inhibition of *Pdyn* neurons in the PAC of behaving animals suggests that chronic PAC-Pdyn dysregulation may be relevant to relapse that is driven by enhanced negative affect/stress/anxiety that maintains the insidious cycle of drug abuse. Further studies are needed to determine whether PAC-Pdyn neurons are a feasible target for addiction-related behaviors, particularly during the withdrawal phase.

It is important to note that species differences exist in the expression pattern of *Pdyn* in the amygdala of humans and rodents. Whereas humans have pronounced *Pdyn* expression in cortical amygdala nuclei such as the PAC and in the AB (22), rodents have the strongest expression in the CeA, basolateral, and medial nuclei (50). Despite the species difference, the PAC circuitry variations examined in several primate and nonprimate models have been shown to be complementary (51, 52). As such, the similar changes in PAC-Pdyn observed in the rodent heroin self-administration model and in human heroin abusers would be predicted to affect similar types of neural circuitry.

Overall, the common disturbance of PDYN in the PAC in opiate addiction and MDD offers new pathophysiological insights into the amygdala relevant to stress and negative affect endophenotypes common in opiate addiction. Moreover, the in vivo imaging technique of DREAMM to manipulate neurochemically distinct cells in discrete brain areas with high molecular specificity in awake freely moving animals will be a valuable neurobiological tool in helping to enhance knowledge about discrete neuronal circuits relevant to neuropsychiatric disorders.

**Methods**

**Human postmortem subjects**

*Cohort I heroin abuse population.* The postmortem human brain specimens used for this population were previously described (21). Briefly, specimens from heroin users and nonabuse control subjects were collected within 24 hours after death. The brains were immediately frozen using dry-ice–cooled isopentane, subsequently cryosectioned, and thaw-mounted onto poly-L-lysine–treated slides. Cause and manner of death were determined after medico-legal examination by the medical examiner. Demographics of this cohort are described in Supplemental Table 1. Inclusion criteria were death associated with heroin intoxication, as verified by toxicology, physical signs of heroin use such as needle track marks, and a history of heroin abuse. Exclusion criteria for all subjects were postmortem interval (PMI) of greater than 24 hours, age less than 50 years, HIV-positive status, and history of alcoholism. Positive alcohol was evident in some of the controls and heroin subjects; however, ethanol concentrations in both groups were not significantly different.

**Cohort II nonabuse control population.** Inclusion criteria were death associated with nonabuse-related causes (e.g., suicide, accidents, trauma, natural death) and no history of illicit drug use. Exclusion criteria were PMI of greater than 24 hours, age less than 50 years, HIV-positive status, and history of alcoholism. Positive alcohol was evident in some of the control subjects; however, ethanol concentrations were not significantly different.
Cohort II heroin abuse and MDD populations. Postmortem human brain specimens from white Hungarian subjects were collected for 3 study populations: (a) heroin abusers, (b) those who committed suicide by hanging with a diagnosis of MDD, and (c) healthy control subjects. All specimens were without head trauma and retrieved at autopsy within 24 hours after death. The brains were collected and stored similarly to those from cohort I. The demographics and general characteristics for cohort II are described in Supplemental Tables 2 and 3. All cases were assessed for common drugs of abuse (including alcohol and marijuana) and also for therapeutic agents. Cigarette toxicity was not assessed, but cigarette use was common for subjects in all groups.

Subjects in the MDD group had a verified psychiatric history and diagnosis of depression and had no documented history or positive toxicity for illicit drug abuse. The same control group was used for both heroin and MDD cohort II subjects (but studied in separate experiments), since the brains of these individual cadavers were collected during the same time, were ethnographically similar, and adhered to the following criteria: negative toxicity of illicit drugs and antidepressants; no history of illicit drug abuse or mood and anxiety disorders. Positive alcohol was evident in very few controls in which ethanol concentrations were similar to those of the limited alcohol-positive subjects identified in the heroin group (none of the MDD subjects were alcohol positive). Exclusion criteria for all subjects were as follows: PMI greater than 24 hours, age greater than 50 years, HIV-positive status, and history of alcoholism.

ISHH

Briefly, as previously described, riboprobes complementary to the human PDYN gene were synthesized from a 1.2-kb human cDNA from the main exon of the PDYN gene (38). The Crf riboprobe used for the rat tissue was a 760-bp cDNA fragment of the gene (1196 bases; GenBank 81648). ISHH was performed on 20-μM coronal cryosections of human or rodent amygdala specimens as previously described (38). The high quality of the post-mortem human brain samples suitable for mRNA expression studies is evident in our ability to routinely detect mRNAs of various genes in these samples (21, 40, 53, 54).

Image analysis

Optical density values were measured using Scion Image (NIH) from digitalized images or FujiFilm Multigauge V3.0 from phosphofilm exposed slides. Measurements were converted to disintegrations per minute (DPM)/mg by reference to coexposed C14 standards. Measurements were taken within discrete amygdala subnuclei at similar levels according to previously published data on the human brain (38). Background noise was also measured and subtracted from the measurements taken. DPM/mg values from duplicate slides were averaged.

Animal studies

Adult (PND 55-69) male Long-Evans rats were obtained from Charles River Laboratories and housed on a reversed 12-hour dark/12-hour light cycle with food and water available ad libitum except as noted below.

Heroin self-administration

Rats were anesthetized with isoflurane (2.5%-4.0% in O2) and catheters (Brian Fromant) implanted into the right jugular vein. Following 1 week recovery, the heroin self-administration paradigm was initiated as previously described (55). Briefly, self-administration experiments were conducted during the dark phase of the light/dark cycle in standard operant chambers housed in sound attenuating boxes (MED Associates Inc.) with 2 retractable levers. Depression of the drug-paired lever (defined as the active lever) resulted in an intravenous heroin (NIDA Drug Supply) injection, whereas depression of the inactive lever had no programmed consequence. Rats were allowed to self-administer heroin (30 μg/kg/injection) under a fixed-ratio-1 (FR1) reinforcement schedule in daily 3-hour sessions for 10 days. Brains were collected at 24 hours following the final drug self-administration session.

Quantitative PCR gene expression

Fresh-frozen bilateral 12-gauge punches were taken from the rat posterior medial lateral cortical (PM/LCo) nuclei equivalent of the PAC; total RNA was isolated using the 5 Prime PerfectPure RNA Tissue Kit and reverse transcribed to cDNA (qScript Kit; VWR). Quantitative real-time PCR (qPCR) analysis was performed using TaqMan-based Ahyn primers and probes (Pdyn Rn00571351_m1; Applied Biosystems). Reactions were run in triplicate, and eukaryotic 18S rRNA (Applied Biosystems) multiplexed as an endogenous control. qPCR and subsequent analysis were performed with a Roche Light Cycler 480 sequence detection system. Quantification of Pdyn gene expression was normalized to eukaryotic 18S rRNA and analyzed using the ΔΔCT method (56).

Viral-mediated DREADD expression

All rats were stereotaxically injected with 2 μl (0.2 μl/min) of purified herpes simplex virus (HSV) vectors expressing a triple hemagglutinin (HA) epitope-tagged hM4Di gene (1567 Kb) under the control of the
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Pdyn promoter (PAC-hM4Di-Pdyn) or a control vector that expresses GFP, also under the control of the Pdyn promoter (PAC-GFP-Pdyn) (28). The vectors were injected unilaterally (right hemisphere) into the PAC (PM1bLCo) using the following coordinates (from bregma): AP, −4.3; ML, 3.7; DV, 9.6 mm.

DREAMM
The DREAMM technique was conducted as described in Supplemental Methods. Rats received an i.p. injection of either vehicle-saline or the DREADD-activating ligand CNO (7–10 days post vector injection) (1 mg/kg). Immediately after vehicle or CNO injection, rats were injected i.p. with FDG (~0.6 mCi). Order of CNO administration was counterbalanced in animals due to the within subjects design of the experiment. Thirty minutes later, rats were anesthetized with 1.5% isoflurane, placed in a prone position on the bed of an R4 microPET rodent scanner (Siemens Medical Solutions), and scanned for 20 minutes using a static acquisition protocol. Analysis of the scans, reconstruction of the images, and statistical parametric mapping (SPM) were conducted as described in Supplemental Methods. Briefly, anatomical regions were based on Paxinos coordinates, and sites of voxel analysis were validated in conjunction with a rat brain imaging atlas intrinsic to the analytical software.

Corticosterone assessment
Whole blood was obtained from a tail-vein catheter under isoflurane anesthesia in hM4Di-Pdyn rats (given either 1 mg/kg of CNO or vehicle i.p. thirty minutes prior to anesthesia). After centrifugation, the remaining supernatant (plasma) was stored at ~80°C. Levels of corticosterone were measured from the blood plasma using the IDS Corticosterone HS EIA kit (Immuno-diagnostics Systems) according to the manufacturer’s instructions.

Sucrose preference test
To measure anhedonic traits, the sucrose preference test was carried out in hM4Di-Pdyn or GFP-Pdyn rats. Rats were given the choice of drinking from 2 bottles, one of which contained a 1% sucrose solution and the other of standard water. Baseline sucrose preference of each animal was determined over a 24-hour period prior to the test. Subsequently, animals were food and water restricted for 15 hours prior to the test session. One hour prior to the test session, all animals (hM4Di- or GFP-Pdyn) received 1 mg/kg i.p. of CNO. Choice preference was observed over 24 hours.

Forced swim test
The procedure used was based on that initially described by Porsolt and modified as described (34). Behavioral studies were carried out from 11:00 to 17:00. Rats were placed individually in a Plexiglass cylinder 46 cm in height with a 21-cm internal diameter, which was filled with water (23°C–24°C) to a depth sufficient to keep adult rats from supporting themselves by placing their paws or tails on the base of the cylinder. Water was changed between swim sessions. There were 2 swim sessions: a pretest 15-minute session and, 24 hours after, a 5-minute swim test. The cumulative time spent in immobile posturing (minimal effort to keep head above water) during the 5-minute test was recorded.

Open field/locomotor activity
Prior to testing, rats were acclimated to the test room for 1 hour. General locomotor activity was examined in a MED Associates activity arena (16 in. x 16 in.) outfitted with a grid of 32 infrared beams. All movements were automatically tracked by beam breaks in 1-hour sessions run in dim lighting for each subject. Between animals, all cages were thoroughly wiped clean with ethanol followed by vinegar.

Statistics
For ISHH, parametric univariate analysis for normal distribution was used to determine group differences in DPM/mg levels. Independent variables (age, PMI, sex, toxicology [prescription and illicit], and brain pH) were included in the general linear model if results from univariate analysis (ANOVA) showed a P value of less than 0.05. For unequal variances (as determined by the 2-sided F test), Welch’s test was used to determine P value. We used 2-tailed, unpaired Student’s t tests (for comparison of 2 groups), 1-way ANOVAs followed by Tukey’s honest significant difference test or 2-tailed Student’s t tests when appropriate (for 3 groups), and 1-way repeated-measures ANOVAs followed by 1-way ANOVAs (to examine significant repeated-measure effects). All values represent mean ± SEM. P < 0.05 and statistical calculations were performed using JMP software (SAS) or GraphPad Prism 5.0.

Study approval
Postmortem human brains for cohort I were obtained as part of the routine autopsy process under protocols approved by Wayne State University’s Human Investigation Committee. Postmortem human brains in cohort II were obtained through the Department of Forensic Medicine at Semmelweis University and from the National Institute of Forensic Medicine (Karolinska Institutet). All of the specimens from cohort II were collected under guidelines approved by the Semmelweis University Research Ethical Committee and the Karolinska Institute Research Ethical Committee. All animal experiments were approved by the institutional animal care and use committees at the Icahn School of Medicine at Mount Sinai and Brookhaven National Laboratory and were conducted in accordance with NIH guidelines.

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