

Poly(ADP-ribose) Engages the TDP-43 Nuclear-Localization Sequence to Regulate Granulo-Filamentous Aggregation

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S Supporting Information

ABSTRACT: TAR DNA-binding protein of 43 kDa (TDP-43) forms granulo-filamentous aggregates in affected brain regions of >95% of patients with ALS and \sim 50% of patients with frontotemporal degeneration (FTD). Furthermore, in disease, TDP-43 becomes Nterminally truncated resulting in protein deposits that are mainly composed of the C-terminal prion-like domain (PrLD). The PrLD is inherently aggregation-prone and is hypothesized to drive protein aggregation of TDP-43 in disease. Here, we establish that the N-terminal region of the protein is critical for rapid TDP-43 granulofilamentous aggregation. We show that the biopolymer poly(ADP-ribose), or PAR, inhibits granulo-filamentous aggregation of TDP-43 by engaging PAR-binding motifs (PBMs) embedded in the TDP-43 nuclear-localization sequence. We demonstrate that progressive N-terminal truncation of TDP-43 can decelerate aggregation kinetics and promote formation of thread-like filaments. Thus, the N-terminal region and the PBMs of TDP-43 promote rapid granulo-filamentous aggregation and antagonize formation of thread-like fibrils. These findings illustrate the complexity of TDP-43 aggregation trajectories.

myotrophic lateral sclerosis (ALS) and frontotemporal Alobar degeneration with ubiquitin-positive inclusions (FTLD-U) are two fatal neurodegenerative disorders characterized by the presence of insoluble aggregates of TAR DNAbinding protein of 43 kDa (TDP-43) in affected brain regions.^{1,2} To date, most of the disease-causing mutations in TDP-43 occur in the C-terminal prion-like domain (PrLD).³ PrLDs are intrinsically disordered regions that can switch from unfolded states to self-templating fibril forms such as the amyloid-like cross- β fibrils.³⁻⁷ The majority of TDP-43 aggregates in ALS/FTLD-U patients have the appearance of granular filaments, but a subset have amyloid-like qualities.^{8–11} Full-length TDP-43 forms granulo-filamentous aggregates in vitro that can transition into thread-like fibrils.4,12,13 This transition is promoted by certain disease-linked mutations in the PrLD, including Q331K.¹² An emerging hypothesis is that the PrLD of TDP-43 may drive the protein aggregation observed in disease.¹²

PrLDs have also been implicated in liquid-liquid phase separation (LLPS), a process by which proteins condense into reversible liquid droplets.¹⁴⁻¹⁶ Of interest are the ALS-linked proteins hnRNPA1, FUS, and TDP-43 which undergo LLPS in vitro.4,16-20 We uncovered that the biopolymer poly(ADPribose) (PAR) potently promotes TDP-43 LLPS in vitro²⁰ and that PAR is elevated in ALS motor neuron nuclei.²¹ PAR is generated by poly(ADP-ribose) polymerases (PARPs),²² and inhibitors of various PARPs (PARP-1, PARP-2, PARP-5a, and PARP-5b) mitigate cytoplasmic aggregation of TDP-43 and TDP-43-associated toxicity to primary neurons and in Drosophila.^{20,21} These findings raised the possibility that PAR may directly regulate TDP-43 aggregation.

To determine if PAR could impact TDP-43 aggregation, we purified full-length human TDP-43 with a His₆-SUMO solubility tag²³ (Figures S1A and S2A). At physiological concentrations of TDP-43 protein,²⁴ cleavage of the His₆-SUMO tag with ubiquitin-like specific protease (Ulp1) induced TDP-43 aggregation over a 200 min period (Figure 1A). The addition of PAR to His₆-SUMO-TDP-43-WT



Figure 1. PAR inhibits TDP-43 aggregation. (A) Ulp1-cleavage of His₆-SUMO-TDP-43-WT increased optical density (OD). Coincubation with 6 µM PAR reduced the optical density of TDP-43-WT. (B) Mono(ADP-ribose) (mADPr, 6 μ M) had no effect on the optical density of TDP-43-WT.

significantly reduced TDP-43-WT aggregation (Figure 1A, Figure S2B,C), while mono(ADP-ribose) had no effect (Figure 1B). Our previous studies established that LLPS of TDP-43 can occur in the presence of a crowding reagent and is promoted by PAR.²⁰ We examined TDP-43-WT by differential interference contrast (DIC) microscopy; before cleavage with and without PAR, the protein remained diffuse and did not form any visible micron-sized aggregates (Figure S3A). However, 30 min after Ulp-1 cleavage, we observed the

Received: August 30, 2018 Revised: December 10, 2018 Published: December 12, 2018 formation of spherical droplets that appeared to coalesce into solid structures after a further 30 min (Figure S3B). Our present data indicate that under conditions that lack a crowding reagent, PAR reduces filamentous aggregation of TDP-43.

The nuclear-localization sequence (NLS) of TDP-43 is a region of intrinsic disorder²⁵ (Figure S1B) and is critical for physically binding to PAR and as well as LLPS of TDP-43 *in vitro*.²⁰ In contrast to cleaved His₆-SUMO-TDP-43-WT, cleaved His₆-SUMO-TDP-43- Δ PAR-binding motif (PBM) (Figure S4A) exhibited decelerated aggregation kinetics (Figure 2A) and took over 18 h to aggregate (Figure 2B).



Figure 2. PAR-binding motifs enable rapid TDP-43 aggregation. (A) Compared to TDP-43-WT, the TDP-43- Δ PAR-binding motif (PBM) did not aggregate in the same time frame. (B) TDP-43- Δ PBM aggregated over 54 h. PAR (6 μ M) had no effect on the optical density of TDP-43- Δ PBM.

The addition of PAR had no effect on the aggregation of TDP-43- Δ PBM (Figure 2B and Figure S4B). Examination of TDP-43- Δ PBM before cleavage revealed no preformed micron-sized aggregates (Figure S3A). Thus, the N-terminal region of TDP-43, and specifically the PBMs, enables rapid aggregation of TDP-43, and PAR engages PBMs within the NLS to reduce TDP-43 aggregation.

Transmission electron microscopy (TEM) revealed that cleavage of the His₆-SUMO tag from both TDP-43-WT and TDP-43- Δ PBM led to the formation of granulo-filamentous aggregates (Figure 3A), consistent with previous TEM studies and of TDP-43 aggregates in human tissue.^{8,10,12} PAR did not drastically alter the structure of the TDP-43-WT or TDP-43- Δ PBM aggregates (Figure 3A). However, PAR significantly reduced the overall size of the TDP-43-WT aggregates, while having no effect on the size of the TDP-43- Δ PBM aggregates (Figure 3B). Indeed, PAR promoted retention of TDP-43-WT in the supernatant fraction after low-speed centrifugation (Figure 3C and Figure S5). Thus, we propose that PAR reduces granulo-filamentous aggregation of TDP-43 via an interaction with PBMs embedded within the NLS.

In ALS and FTLD-U, splicing defects and proteolytic cleavage can elicit formation of TDP-43 C-terminal fragments that contain the PrLD.^{26–28} As the C-terminal fragments of TDP-43 either contain a partial PAR-binding region (TDP-43-C35) or lack the PAR-binding region (TDP-43-C25) (Figure S1A), we examined the aggregation kinetics of these two C-terminal fragments. Strikingly, the ability of TDP-43-C35 and TDP-43-C25 to form turbid aggregates was, like TDP-43- Δ PBM, reduced compared to TDP-43-WT (Figure 4A). Examination by TEM revealed that TDP-43-C35 formed granulo-filamentous aggregates and thread-like fibrils (Figure 4B). The TDP-43-C25 aggregates were unreactive to the



Figure 3. PAR reduces TDP-43 aggregation. (A) Ulp1 cleavage of HIS₆-SUMO-TDP-43-WT and HIS₆-SUMO-TDP-43-ΔPBM led to granulo-filamentous aggregation (hatched boxes). PAR (6 μM) reduced aggregate size of TDP-43-WT and had no effect on TDP-43-ΔPBM (hatched boxes). (B) Quantification of aggregate size. Mean (±SD), one-way ANOVA (P < 0.0001), and Kruskal–Wallis test. (C) PAR (6 μM) reduced the amount of TDP-43-WT in the pellet fraction at 400g (Figure S5). Mean (±SD), two-way ANOVA, and Tukey's test.



Figure 4. C-terminal fragments of TDP-43 have altered aggregation properties. (A) The increase in optical density of TDP-43-C35 and TDP-43-C25 was reduced compared to TDP-43-WT. (B) TDP-43-WT, TDP-43-C35, and TDP-43-C25 formed granulo-filamentous protein aggregates (black arrows). TDP-43-C25 also formed thread-like aggregates (white arrows).

amyloid diagnostic dye Thioflavin T (Figure S6). Combined, these data reveal that the N-terminal portion of TDP-43 contributes to granulo-filamentous aggregation and antagonizes the transition into thread-like oligomers.

Here, we show that N-terminal portions of TDP-43 contribute to granulo-filamentous aggregation. Our data indicate that PAR interacts with PBMs embedded within the NLS of TDP-43 to reduce granulo-filamentous aggregation. Defining the mechanism by which PAR binding reduces TDP-43 aggregation will require further study. Regions within the N-terminal domain of TDP-43 regulate self-oligomerization.^{25,29-32}Thus, PAR-binding to the NLS adjacent to the N-terminal domain may physically block interactions that contribute toward aggregation. In disease, TDP-43 aggregates appear to be predominantly granulo-filamentous. Thus, agents that antagonize contributions from the N-terminal region of TDP-43 could have therapeutic utility. However, as oligome-rization is essential for TDP-43 function,^{25,29-32} agents that prevent this functional oligomerization could be detrimental. Understanding under what circumstances functional versus toxic TDP-43 assemblies form,³³ how they differ, and how they

are resolved will help develop therapeutic strategies to selectively target toxic assemblies.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.bio-chem.8b00910.

TDP-43 protein domains, protein purification analysis, turbidity assay, analysis of the sedimentation assay, and detailed materials and methods (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Lee, E. B., Lee, V. M., and Trojanowski, J. Q. (2012) Gains or losses: molecular mechanisms of TDP43-mediated neurodegeneration. *Nat. Rev. Neurosci.* 13, 38–50.

(2) Guo, L., and Shorter, J. (2017) Biology and Pathobiology of TDP-43 and Emergent Therapeutic Strategies. *Cold Spring Harbor Perspect. Med.* 7, a024554.

(3) Harrison, A. F., and Shorter, J. (2017) RNA-binding proteins with prion-like domains in health and disease. *Biochem. J.* 474, 1417–1438.

(4) Guo, L., Kim, H. J., Wang, H., Monaghan, J., Freyermuth, F., Sung, J. C., O'Donovan, K., Fare, C. M., Diaz, Z., Singh, N., Zhang, Z. C., Coughlin, M., Sweeny, E. A., DeSantis, M. E., Jackrel, M. E., Rodell, C. B., Burdick, J. A., King, O. D., Gitler, A. D., Lagier-Tourenne, C., Pandey, U. B., Chook, Y. M., Taylor, J. P., and Shorter, J. (2018) Nuclear-Import Receptors Reverse Aberrant Phase Transitions of RNA-Binding Proteins with Prion-like Domains. *Cell* 173, 677–692.

(5) Kato, M., Han, T. W., Xie, S., Shi, K., Du, X., Wu, L. C., Mirzaei, H., Goldsmith, E. J., Longgood, J., Pei, J., Grishin, N. V., Frantz, D. E., Schneider, J. W., Chen, S., Li, L., Sawaya, M. R., Eisenberg, D., Tycko, R., and McKnight, S. L. (2012) Cell-free formation of RNA granules: low complexity sequence domains form dynamic fibers within hydrogels. *Cell 149*, 753-767.

(6) Murray, D. T., Kato, M., Lin, Y., Thurber, K. R., Hung, I., McKnight, S. L., and Tycko, R. (2017) Structure of FUS Protein Fibrils and Its Relevance to Self-Assembly and Phase Separation of Low-Complexity Domains. *Cell* 171, 615–627.

(7) Alberti, S., Halfmann, R., King, O., Kapila, A., and Lindquist, S. (2009) A systematic survey identifies prions and illuminates sequence features of prionogenic proteins. *Cell* 137, 146–158.

(8) Robinson, J. L., Geser, F., Stieber, A., Umoh, M., Kwong, L. K., Van Deerlin, V. M., Lee, V. M., and Trojanowski, J. Q. (2013) TDP-43 skeins show properties of amyloid in a subset of ALS cases. *Acta Neuropathol.* 125, 121–131.

(9) Bigio, E. H., Wu, J. Y., Deng, H. X., Bit-Ivan, E. N., Mao, Q., Ganti, R., Peterson, M., Siddique, N., Geula, C., Siddique, T., and Mesulam, M. (2013) Inclusions in frontotemporal lobar degeneration with TDP-43 proteinopathy (FTLD-TDP) and amyotrophic lateral sclerosis (ALS), but not FTLD with FUS proteinopathy (FTLD-FUS), have properties of amyloid. *Acta Neuropathol.* 125, 463–465.

(10) Thorpe, J. R., Tang, H., Atherton, J., and Cairns, N. J. (2008) Fine structural analysis of the neuronal inclusions of frontotemporal lobar degeneration with TDP-43 proteinopathy. *J. Neural Transm* (*Vienna*) 115, 1661–1671.

(11) Fang, Y. S., Tsai, K. J., Chang, Y. J., Kao, P., Woods, R., Kuo, P. H., Wu, C. C., Liao, J. Y., Chou, S. C., Lin, V., Jin, L. W., Yuan, H. S., Cheng, I. H., Tu, P. H., and Chen, Y. R. (2014) Full-length TDP-43 forms toxic amyloid oligomers that are present in frontotemporal lobar dementia-TDP patients. *Nat. Commun.* 5, 4824.

(12) Johnson, B. S., Snead, D., Lee, J. J., McCaffery, J. M., Shorter, J., and Gitler, A. D. (2009) TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and increase toxicity. *J. Biol. Chem.* 284, 20329–20339.

(13) Jackrel, M. E., DeSantis, M. E., Martinez, B. A., Castellano, L. M., Stewart, R. M., Caldwell, K. A., Caldwell, G. A., and Shorter, J. (2014) Potentiated Hsp104 variants antagonize diverse proteotoxic misfolding events. *Cell* 156, 170–182.

(14) Hyman, A. A., Weber, C. A., and Julicher, F. (2014) Liquidliquid phase separation in biology. *Annu. Rev. Cell Dev. Biol.* 30, 39– 58.

(15) Brangwynne, C. P. (2013) Phase transitions and size scaling of membrane-less organelles. J. Cell Biol. 203, 875–881.

(16) Gomes, E., and Shorter, J. (2018) The molecular language of membraneless organelles. J. Biol. Chem., 1.

(17) Shin, Y., and Brangwynne, C. P. (2017) Liquid phase condensation in cell physiology and disease. *Science* 357, eaaf4382.

(18) Wang, A., Conicella, A. E., Schmidt, H. B., Martin, E. W., Rhoads, S. N., Reeb, A. N., Nourse, A., Ramirez Montero, D., Ryan, V. H., Rohatgi, R., Shewmaker, F., Naik, M. T., Mittag, T., Ayala, Y. M., and Fawzi, N. L. (2018) A single N-terminal phosphomimic disrupts TDP-43 polymerization, phase separation, and RNA splicing. *EMBO* J. 37, No. e97452.

(19) Conicella, A. E., Zerze, G. H., Mittal, J., and Fawzi, N. L. (2016) ALS Mutations Disrupt Phase Separation Mediated by alpha-Helical Structure in the TDP-43 Low-Complexity C-Terminal Domain. *Structure* 24, 1537–1549.

(20) McGurk, L., Gomes, E., Guo, L., Mojsilovic-Petrovic, J., Tran, V., Kalb, R. G., Shorter, J., and Bonini, N. M. (2018) Poly(ADP-Ribose) Prevents Pathological Phase Separation of TDP-43 by Promoting Liquid Demixing and Stress Granule Localization. *Mol. Cell* 71, 703–717.

(21) McGurk, L., Mojsilovic-Petrovic, J., Van Deerlin, V. M., Shorter, J., Kalb, R. G., Lee, V. M., Trojanowski, J. Q., Lee, E. B., and Bonini, N. M. (2018) Nuclear poly(ADP-ribose) activity is a therapeutic target in amyotrophic lateral sclerosis. *Acta neuropathologica communications 6*, 84–95.

(22) Gibson, B. A., and Kraus, W. L. (2012) New insights into the molecular and cellular functions of poly(ADP-ribose) and PARPs. *Nat. Rev. Mol. Cell Biol.* 13, 411–424.

(23) Molliex, A., Temirov, J., Lee, J., Coughlin, M., Kanagaraj, A. P., Kim, H. J., Mittag, T., and Taylor, J. P. (2015) Phase separation by low complexity domains promotes stress granule assembly and drives pathological fibrillization. *Cell 163*, 123–133.

(24) Ling, S. C., Albuquerque, C. P., Han, J. S., Lagier-Tourenne, C., Tokunaga, S., Zhou, H., and Cleveland, D. W. (2010) ALS-associated mutations in TDP-43 increase its stability and promote TDP-43 complexes with FUS/TLS. *Proc. Natl. Acad. Sci. U. S. A.* 107, 13318– 13323.

(25) Chang, C. K., Wu, T. H., Wu, C. Y., Chiang, M. H., Toh, E. K., Hsu, Y. C., Lin, K. F., Liao, Y. H., Huang, T. H., and Huang, J. J. (2012) The N-terminus of TDP-43 promotes its oligomerization and enhances DNA binding affinity. *Biochem. Biophys. Res. Commun.* 425, 219–224.

(26) Tsuji, H., Arai, T., Kametani, F., Nonaka, T., Yamashita, M., Suzukake, M., Hosokawa, M., Yoshida, M., Hatsuta, H., Takao, M., Saito, Y., Murayama, S., Akiyama, H., Hasegawa, M., Mann, D. M., and Tamaoka, A. (2012) Molecular analysis and biochemical classification of TDP-43 proteinopathy. *Brain 135*, 3380–3391.

(27) Xiao, S., Sanelli, T., Chiang, H., Sun, Y., Chakrabartty, A., Keith, J., Rogaeva, E., Zinman, L., and Robertson, J. (2015) Low molecular weight species of TDP-43 generated by abnormal splicing form inclusions in amyotrophic lateral sclerosis and result in motor neuron death. *Acta Neuropathol.* 130, 49–61.

(28) Kametani, F., Obi, T., Shishido, T., Akatsu, H., Murayama, S., Saito, Y., Yoshida, M., and Hasegawa, M. (2016) Mass spectrometric analysis of accumulated TDP-43 in amyotrophic lateral sclerosis brains. *Sci. Rep. 6*, 23281.

(29) Afroz, T., Hock, E. M., Ernst, P., Foglieni, C., Jambeau, M., Gilhespy, L. A. B., Laferriere, F., Maniecka, Z., Pluckthun, A., Mittl, P., Paganetti, P., Allain, F. H. T., and Polymenidou, M. (2017) Functional and dynamic polymerization of the ALS-linked protein TDP-43 antagonizes its pathologic aggregation. *Nat. Commun.* 8, 45.

(30) Jiang, L. L., Xue, W., Hong, J. Y., Zhang, J. T., Li, M. J., Yu, S. N., He, J. H., and Hu, H. Y. (2017) The N-terminal dimerization is required for TDP-43 splicing activity. *Sci. Rep.* 7, 6196.

(31) Romano, V., Quadri, Z., Baralle, F. E., and Buratti, E. (2015) The structural integrity of TDP-43 N-terminus is required for efficient aggregate entrapment and consequent loss of protein function. *Prion* 9, 1–9.

(32) Zhang, Y. J., Caulfield, T., Xu, Y. F., Gendron, T. F., Hubbard, J., Stetler, C., Sasaguri, H., Whitelaw, E. C., Cai, S., Lee, W. C., and Petrucelli, L. (2013) The dual functions of the extreme N-terminus of TDP-43 in regulating its biological activity and inclusion formation. *Hum. Mol. Genet.* 22, 3112–3122.

(33) Vogler, T. O., Wheeler, J. R., Nguyen, E. D., Hughes, M. P., Britson, K. A., Lester, E., Rao, B., Betta, N. D., Whitney, O. N., Ewachiw, T. E., Gomes, E., Shorter, J., Lloyd, T. E., Eisenberg, D. S., Taylor, J. P., Johnson, A. M., Olwin, B. B., and Parker, R. (2018) TDP-43 and RNA form amyloid-like myo-granules in regenerating muscle. *Nature 563*, 508.