OPINION

Rethinking ovarian cancer: recommendations for improving outcomes

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Abstract | There have been major advances in our understanding of the cellular and molecular biology of the human malignancies that are collectively referred to as ovarian cancer. At a recent Helene Harris Memorial Trust meeting, an international group of researchers considered actions that should be taken to improve the outcome for women with ovarian cancer. Nine major recommendations are outlined in this Opinion article.

Patient prognosis has considerably improved for many solid cancer types. However, the survival rate of women with epithelial ovarian cancer has changed little since platinumbased treatment was introduced more than 30 years ago¹⁻³ (FIG. 1). Invasive epithelial ovarian cancer is widely seen and treated as a single disease entity, with little stratification of the different histological or molecular subtypes. At a recent Helene Harris Memorial Trust (HHMT) meeting of leading ovarian cancer researchers and clinician scientists, which was sponsored and organized by Ovarian Cancer Action (see Further information) (BOX 1), we asked the question: what actions can we take to improve the outcome for women with ovarian cancer? A consensus regarding the major barriers to success and the current most pressing questions led delegates to propose nine priorities for action, which we describe in this Opinion article.

Ovarian cancer is many diseases

It is essential that researchers, pathologists, epidemiologists and clinicians understand that ovarian cancer is a general term for a series of molecularly and aetiologically distinct diseases that simply share an anatomical location. Recent pathological and genomic findings indicate that many ovarian cancers are derived from non-ovarian tissues and that the different ovarian cancer histotypes share few molecular similarities⁴ (FIG. 2). For example, the distal fallopian tube has been identified as a source of high-grade serous ovarian cancers⁵⁻⁷. The relative importance of the fallopian tube compared with the ovarian surface epithelium in the genesis of high-grade serous ovarian cancers is still being debated, however, this finding has important implications for screening, prevention and understanding the molecular biology of the disease. Ovarian clear cell and endometrioid cancers have a strong epidemiological link with endometriosis. High-frequency somatic mutations of the PI3K catalytic subunit PIK3CA and AT-rich interactive domain-containing protein 1A (ARID1A) in adjacent endometriotic lesions link endometriosis, clear cell cancers and endometrioid cancers⁸⁻¹⁰. Most invasive mucinous ovarian cancers are actually metastases to the ovary from other solid cancer types, including gastrointestinal tumours¹¹⁻¹³. Improved histotype

classification using immunological markers and genomic studies has shown that many tumours that have previously been designated high-grade endometrioid cancers should be reclassified as serous cancers^{14–16}. Low-grade invasive tumours are still regarded as ovarianderived, but the initiating cells are unknown and it is possible that their site of origin will be re-evaluated in the future. Nevertheless, it is clear that serous borderline cancers are not precursor lesions for the majority of highgrade serous ovarian cancer, as they have a distinct range of mutational events¹⁷.

Further subclassification of histotypes is based on signalling pathway activation¹⁸, genomic events or gene expression profiling¹⁹. Some ovarian cancers have more in common with certain types of renal cancer or breast cancer than they have with other ovarian histologies. For example, high-grade serous ovarian cancers share genomic and transcriptional features with basal-like breast cancers^{17,20}. Ovarian clear cell cancers have similar expression phenotypes to renal clear cell and uterine clear cell cancers²¹. and women with ovarian clear cell cancer may benefit from the use of drugs, such as sunitinib, that are active in patients with renal cancer ²². Taking a rigorous view, the ovarian histotypes should be regarded as distinct diseases, as their cell of origin, epidemiology and driver mutations are quite different.

The term ovarian cancer is therefore misleading. It is not a single disease, and a considerable proportion of tumours do not arise from ovarian tissue. The unifying clinical feature of all ovarian cancers is frequent loco-regional dissemination to the ovary and related pelvic organs. We considered whether the term ovarian cancer should be replaced with the terms pelvic or peritoneal cancer but we recognized the confusion that might ensue for patients and physicians, as well as in the scientific literature, especially during a transition period. Before the term ovarian cancer is abandoned, the disparate origins of this disease need to be more widely understood by patients, physicians and scientists.

Improved clinical trial design

A shift in emphasis is needed from Phase III clinical trials to earlier phase trial designs that include molecular studies. Given the



Figure 1 | **Survival in ovarian cancer**. Data from the United States (part **a**), Australia (part **b**) and Canada (part **c**) show that there has been little change in 1-year, 3-year and 5-year survival rates post-diagnosis of patients with ovarian cancer over the past 20 years. Data in part **a** are from the <u>Surveillance, Epidemiology and End Results</u> (SEER) database (see Further information) obtained between 1980 and 2004. Data in part **b** are from The Cancer Council of Victoria, Victoria, Australia, obtained between 1990 and 2004. Data in part **c** are from The Cheryl Brown Outcomes Unit, British Columbia, Canada, obtained between 1980 and 2004.

differences in ovarian cancer subtypes, clinical trials should no longer aggregate the various ovarian histologies. Trial design should also consider molecular parallels between ovarian cancer and other solid cancers. Clinical reporting and scientific publications should use reproducible diagnostic methods and standardized terms for these disease entities²³. The current single approach to treatment should rapidly evolve to evidence-based stratification that is based on molecular drivers and histotype-specific treatments.

The ovarian cancer field lags behind other cancer research fields in incorporating targeted therapies into standard treatment (FIG. 3). With the possible exception of poly(ADP-ribose) polymerase (PARP) inhibitors and angiogenesis inhibitors (discussed below), single-agent, molecularly targeted therapies have only yielded small increments in progression-free survival in ovarian cancers. Therefore, a major shift in the way clinical trials are designed is urgently required. The ceiling in efficacy that has been witnessed with all-inclusive Phase III trials means that the priority should now be on subtype-specific (randomized) Phase II clinical trials that are based on sound scientific rationale. In addition, 'window trials' (REF. 24) with clear readouts of pathway inhibition that correlate with clinical outcome will expedite the clinical investigation of novel therapies.

Improved blood and imaging biomarkers that accurately measure response and residual disease are also needed: the current criteria are an impediment to the better evaluation of response. Contrast-enhanced computed tomography (CT) imaging is the standard non-surgical method for staging and assessing response; however, it is difficult to detect small peritoneal deposits. For example, when peritoneal tumour deposits are <1cm in size, the sensitivity of CT is only 7-28%, and this is further dependent on the anatomical location of the disease²⁵. The measurement of circulating levels of the ovarian tumour antigen CA125 (also known as mucin 16) is routinely used to monitor disease recurrence; however, markers that are molecularly based and also sensitive for low-volume residual disease are needed. Preliminary results from the detection of the cellular by-products of ovarian cancer cells in the blood are promising but need wider evaluation²⁶.

Prevention and early detection

The identification of patients who are at an increased genetic risk of ovarian cancer currently offers the most effective measure for prevention and early detection of this disease. High-grade serous ovarian cancer is diagnosed at an advanced stage in approximately 70% of patients, and these women have a substantially worse outcome than the outcome of those with early stage disease. It is important to recognize that the poor prognosis in advanced stage disease is a function of at least two factors - the extent of disease, and thus the ability to surgically remove the tumour, and the differences in the biology of the tumours that remain confined to the pelvic space compared with those that widely disseminate27. Early detection tests should focus on identifying the precursors of advanced stage high-grade serous ovarian tumours. Tests should not be evaluated on patients with low-grade stage I/II serous tumours that have activating RAS pathway mutations, as these are not typically precursors of high-grade serous tumours.

Traditional concepts of metastasis are difficult to apply to ovarian cancer. There is no anatomical barrier to seeding throughout the peritoneal cavity for a high-grade serous ovarian cancer that arises on the surface of the ovary or fallopian tube, or a clear cell ovarian cancer that arises in an endometriotic deposit. Emerging insights into disease progression of high-grade serous ovarian cancers now suggest that the early detection of low-volume advanced stage, rather than early stage, ovarian cancer may be a more realistic goal of screening studies²⁸, although the clinical value of this strategy remains to be determined. More data about the natural history of precursor lesions in the fallopian tube epithelium and their rate of transformation to invasive carcinoma are needed to plan future screening methods and technologies²⁹. Recent data from the Prostate, Lung, Colorectal and Ovarian (PLCO) trial indicate that current serological diagnostics and imaging tests are insufficiently sensitive to alter outcome in screened populations³⁰, although results from the large UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS) trial remain to be reported. On the basis of current data, widespread screening is not yet justified and its value in the management of high-risk women is unknown³¹. Screening for endometrioid and clear cell ovarian cancer will be particularly challenging, as endometriosis is a common, often subclinical, disease and risk factors for transformation have not yet been identified.

The identification of women who carry germline mutations in BRCA1, BRCA2, RAD51C and genes that encode other DNA repair proteins provides the most effective preventative strategy for high-grade serous ovarian cancers, as the removal of the ovaries and fallopian tubes can reduce the risk of disease in carriers by 80%^{32,33}. Given the newly appreciated importance of the fallopian tube in the genesis of high-grade serous ovarian cancer, it is recommended that the complete removal of the fallopian tube should become standard of care in any woman undergoing hysterectomy and/or removal of the ovaries (oophorectomy). Oophorectomy in premenopausal women induces early menopause. As a consequence, and with the changed view of the role of the fallopian tube in ovarian cancer, some clinicians have recommended that only the fallopian tubes should be removed (salpingectomy) in women with germline BRCA1 or BRCA2 mutations, or in women with a strong family history of breast and/or ovarian cancer³⁴. However, until comprehensive comparative data are available, the group felt it was premature to recommend that only the fallopian tubes are removed in high-risk women.

Although there is no evidence that current screening strategies reduce mortality in highrisk women, future screening approaches are likely to require the identification of those women who are at the greatest genetic risk in order to reduce the number of falsepositive results in a screened population³⁰. Advances in high-throughput sequencing and mutation scanning offer the potential to greatly reduce the cost of mutation testing, and to therefore extend testing to more women, especially women with high-grade serous ovarian cancers. Recent genome-wide association studies, which were enabled by large cancer cohorts assembled by research consortia (see below), have identified a number of new ovarian cancer risk loci35,36. The identification of novel genetic risk loci is improved by histotype stratification^{35,36}, and therefore genetic and epidemiological studies of tumours should embrace the appropriate classification of ovarian cancer.

Identification of new targets

Novel approaches are needed to identify therapeutic targets, especially in high-grade serous ovarian cancer. Over the past decade, increasingly comprehensive genomic analyses of ovarian cancer have failed to reveal new oncogenic drivers in the more common histotypes. Large-scale gene expression¹⁹ and DNA copy number analyses^{37,38} and mutational screens39 have not identified new high-frequency 'drugable' targets in highgrade serous ovarian cancers, perhaps with the possible exception of cyclin E1 (CCNE1). CCNE1 amplification is associated with poor outcome⁴⁰ and oncogene addiction *in vitro*⁴¹. Further studies are needed to validate the prognostic significance of other amplified genes in high-grade serous ovarian cancers and to develop therapeutic approaches that target them. Mutations in the RAS pathway are found in more than 70% of low-grade serous ovarian cancers18, and specific clinical trials for this unique serous neoplasm

Box 1 | The meeting

For 25 years the Helene Harris Memorial Trust (HHMT) has recognized the importance of communication between scientists and clinicians for improving the early detection of ovarian cancers and the treatment of patients with advanced disease, bringing together international experts on a biannual basis (see the <u>Helene Harris Memorial Trust International Forum on Ovarian Cancer</u> website; see Further information). In January 2011 researchers met in Florida, USA, over 4 days at the HHMT 12th International Forum on Ovarian Cancer (sponsered by Ovarian Cancer Action) to consider the latest findings in basic, translational and clinical research in ovarian cancer. With both a sense of optimism that was associated with recent advances and a sense of frustration with the limited improvements in outcomes in the past, we considered new ways forwards on the basis of recent findings. The delegates attending the meeting are listed in <u>Supplementary</u> information S2 (table) (see Further information). The resulting Opinion article reflects the consensus of the meeting and the listed authors have all contributed to this manuscript.

PERSPECTIVES

One of the most challenging aspects of high-grade serous ovarian cancer is the recognition that widespread gene expression and DNA copy number changes provide extensive opportunities for adaptation and the development of resistance¹⁷. Ongoing genomic instability may drive intratumoural genetic heterogeneity and may increase the probability of treatmentresistant clone selection^{51,52}. However, genetic alterations in tumour cells are not the only contributor to therapy resistance. Cell non-autonomous effects are mediated by the tumour microenvironment, which is recruited by the tumour cells (see below), providing a cytokine milieu on which the tumour thrives, becomes resistant to current therapies and becomes more invasive. Single-agent treatment is unlikely to be effective when cancerous cells are capable of using other non-targeted pathways. Therefore, it is crucial that we develop combination therapeutic approaches that anticipate tumour adaptive responses to individual targeted agents. The exploration of novel combinations of targeted therapeutics will only be possible if we can overcome the current reluctance of the pharmaceutical industry to be involved in clinical trials that combine drugs developed by different companies. Combination studies must be underpinned by the preclinical investigation of dosing schedules in appropriate model systems.

The tumour microenvironment

Targeting the tumour microenvironment provides an important adjunct to molecular therapeutics and chemotherapy that are directed towards the tumour cell. The inherent instability of the genome of high-grade serous ovarian cancer has shifted attention to targeting the tumour microenvironment, which comprises a large proportion of the cell mass of many ovarian cancers. To date, encouraging results have been obtained with trials in ovarian cancer that target the angiogenic factor vascular endothelial growth factor (VEGF) using the therapeutic antibody bevacizumab. The addition of bevacizumab to conventional chemotherapy improves progression-free survival in relapsed, platinum-resistant ovarian cancer⁵³. In addition, large randomized Phase III trials that investigated the role of bevacizumab maintenance therapy in the first-line setting (Gynecologic Oncology Group (GOG-0218) (ClinicalTrials. gov identifier: NCT00262847) and ICON7 (NCT00483782)) and in platinum-sensitive relapsed disease (OCEANS) (NCT00434642) have been completed. The results of all three

are urgently needed. Activating mutations

in PIK3CA8, and amplification of recep-

tor tyrosine kinases including MET²² and

ERBB2 (REF. 42), provide novel therapeutic

approaches in clear cell ovarian cancers.

Although high-frequency, novel somatic

mutations have been found in granulosa43

and clear cell¹⁰ ovarian cancer, these uncom-

mon subtypes only account for a small pro-

Genomic studies have reinforced the cen-

portion of the overall disease mortality.

trality of TP53 (which encodes the tumour

suppressor p53) mutations²⁰; BRCA pathway

disruption44; and homologous recombination

repair (HRR) deficiency in high-grade serous

ovarian cancers. The notion that many high-

grade serous ovarian cancers are BRCA-like

through alterations in other proteins in the

HRR pathway45 has resulted in the explora-

tion of PARP inhibitors46. Preliminary find-

ings suggest efficacy of PARP inhibitors in

a number of settings, including the use of

olaparib as a maintenance treatment after

chemotherapy in women with platinum-

sensitive recurrent high-grade serous ovar-

ian cancers47. About 50% of all high-grade

serous ovarian cancers show disruption of

the BRCA pathway either by germline or

somatic mutation, or by epigenetic silenc-

ing of pathway members³⁹. Consistent with

mutational studies³⁹, functional assays show

ovarian cancers have defective formation of

repair-defective tumours correspond to those

PARP inhibitors, this may provide an impor-

that roughly 50% of all high-grade serous

HRR foci following DNA damage⁴⁸. If the

patients with relatively good responses to

tant predictive test of treatment response.

These data suggest that exploiting mutator

cancer can offer a valuable approach to novel

therapies^{49,50}. Synthetic lethality as a strategy

high-grade serous ovarian cancer should be

further explored with RNA interference and

small-molecule screens.

for targeting DNA repair and p53 pathways in

phenotypes in high-grade serous ovarian



Figure 2 | **The origins of ovarian cancer**. Ovarian cancer is a collective term for invasive cancers that are derived from different tissues. Most invasive mucinous ovarian cancers are metastases to the ovary, often from the gastrointestinal tract, including the colon, appendix or stomach. Endometrioid and clear cell ovarian cancers are derived from endometriosis, which in turn is associated with retrograde menstruation from the endometrium. High-grade serous ovarian cancers are derived from the surface of the ovary and/or the distal fallopian tube; the relative contribution that the two sites make to these tumours remains unclear. Benign and low-malignant potential (borderline) tumours are not shown. Such tumours are thought to be of ovarian origin, however, the originating cells are not defined and their derivation may be revised in the future. Histological images courtesy of R. Drapkin, Dana-Farber Cancer Institute, USA, and C. Crum, Brigham and Women's Hospital, USA.

trials, presented in abstract form only, suggest improvements in progression-free survival when bevacizumab is given as concurrent or maintenance therapy. However, despite the presumed stability of the tumour endothelium, resistance to anti-VEGF agents has rapidly emerged. Understanding resistance pathways and developing predictors of patient response are crucial for better exploiting anti-angiogenic therapies^{54,55}.

A spontaneous antitumour immune response in the form of tumour-reactive T cells and/or antibodies has been demonstrated in some patients with ovarian cancer^{56,57}. The increased infiltration of lymphocytes in tumour islets predicts significantly longer survival in ovarian cancer⁵⁸. Conversely, the detection of high numbers of T regulatory cells, which mediate immune suppression, predicts poor patient survival^{59,60}. The presence of additional immunosuppressive cell subtypes, such as B7-H4-expressing tumour macrophages⁶¹, has also been correlated with poor outcome.

Complex networks of inflammatory cytokines and chemokines62 regulate communication between malignant cells and supporting stroma in ovarian cancer. These cytokines and their intracellular signalling pathways can make malignant cells resistant to apoptosis, can facilitate the evasion of tumour immunity and can promote angiogenesis. Tumour cells typically trigger inflammatory cytokine networks as a means of escaping immune recognition in spite of surrounding inflammation63,64. In experimental animal models, targeting these key inflammatory cytokines and chemokines has been shown to abrogate these processes that are key to the progression of ovarian cancer^{62,65}.

The association of intratumoural T cells with increased survival, and T regulatory cells with worse survival, indicates that

ovarian cancers could respond to immune therapy. Pilot studies indicate that therapies that capitalize on pre-existing antitumour immune responses can be successful in ovarian cancer. For example, objective responses and/or prolonged survival have been seen with immune checkpoint blockade using an antibody against cytotoxic T lymphocyte protein 4 (CTLA4)66,67, and Phase II studies are currently underway. The understanding that some conventional chemotherapy drugs also have immunomodulatory activity offers new opportunities for designing combinatorial approaches for women with ovarian cancer whose tumours exhibit pre-existing antitumour immunity in terms of high levels of intraepithelial tumour-infiltrating lymphocytes. In addition, trials targeting inflammatory cytokines^{68,69} are currently in early stages, but they can offer important biological lessons for clinicians.

There needs to be rapid development of further Phase II/III studies that focus on agents targeting key pathways in the tumour microenvironment that not only assess their efficacy alongside established cytotoxic regimes, but that also aim to establish the optimal use of these agents for maintenance therapy.

Tumour adaptation and resistance

Improved understanding of clonal diversity in tumours prior to treatment and mechanisms of tumour adaptation and the acquisition of resistance following treatment are essential if more durable responses to therapy are to be achieved. Many women respond well to first-line treatment, but frequently relapse with chemotherapyresistant disease. Evolutionary models of clonal selection may explain drug resistance in cancer. In acute lymphocytic leukaemia and chronic myeloid leukaemia, point mutations that confer resistance to imatinib exist at low prevalence before treatment and can become highly enriched during relapse^{70,71}. Autopsy studies on advanced pancreatic cancer have shown profound genetic heterogeneity, with 52% of mutations being present in subclonal populations^{72,73}. Whether this paradigm can explain resistance in ovarian cancer, and in particular whether heterogeneity is associated with primary platinum resistance, needs further investigation. With advances in next-generation sequencing we now have the tools to investigate clonal diversity84,85 and to start inferring the evolutionary changes that may contribute to resistance.

Despite the plethora of research that has been dedicated to the mechanisms of platinum resistance, this is yet to translate to clinical practice. Undoubtedly, deficiencies in our knowledge of the fundamental changes that occur within the tumour — both the epithelial fraction and the microenvironment - throughout treatment have impeded progress in this area. More recently, studies with paired tumour samples that were collected before treatment and following disease relapse have provided some of the first insights into clonal variation and mechanisms of resistance in vivo^{52,74}, including reversion of germline mutations in BRCA1 or BRCA2 (REF. 86). Serially obtained biopsy samples of disease sites should be a central component in clinical trial design.

Importance of international consortia

International consortia involving large biological and clinical data sets must be encouraged; however, these require high fidelity and transparency in analytical approaches. Ovarian cancer is a relatively uncommon disease and, together with its histological diversity, it is thus difficult to collect substantial numbers of samples of specific subtypes. With the increased recognition of the importance of stratification by subtype, global collaborations and consortia such as the Ovarian Cancer Association Consortium (OCAC), The Cancer Genome Atlas (TCGA), The Australian Ovarian Cancer Study (AOCS), Ovarian Cancer Therapy - Innovative Models Prolong Survival (OCTIPS) and Ovarian Tumour Tissue Analysis Consortium (OTTA) are crucial to furthering our understanding of the molecular biology of ovarian tumours and genetic risk factors. Such collaborative endeavours require highly ordered, standardized and quality-controlled strategies for the collection and management of biological specimens. The combination of highthroughput technologies with large sample sets allows detailed biological and translational studies. However, the challenges of managing large molecular data sets, each identifying a multitude of putative abnormalities - many of which may prove to be artefacts — are substantial. Moreover, the opportunity to introduce human operator error remains unacceptably high. Methodologies used in the analysis of such data sets should be wholly transparent, reproducible and biologically validated in the laboratory before any such findings can be put into practice. Errors made in the development of the OvaCheck early



Figure 3 | **Evolution of chemotherapy for ovarian cancer over the past 50 years.** It has proved difficult to progress beyond platinum-based therapy, which was introduced in the late 1970s and which remains the standard of care. Cisplatin, and subsequently carboplatin, which has a lower toxicity profile, have been combined with other agents, including taxanes. Most recently, liposomal doxorubicin has become commonly used with carboplatin, especially in a relapse setting. It is notable that the combination of carboplatin and liposomal doxorubicin involves similar drugs to those that were used in the mid-1970s, albeit with reduced side effects. It is likely that ovarian cancer treatment will considerably evolve in the coming years with the introduction of molecularly targeted agents, such as poly(ADP-ribose) polymerase (PARP) inhibitors, histotype-specific treatments and dose-dense regimes, including the use of weekly taxane administration.

detection test underscore the importance of rigorous biostatistical support, especially when dealing with complex molecular data sets⁷⁵.

Better experimental models

Improved experimental models of ovarian cancer are required that more closely resemble the human disease. New model systems are needed that reflect the various originating cells, as well as the underlying genomic events driving each disease. Although mouse models of ovarian cancer have been developed⁷⁶⁻⁷⁹, these have not for the most part recapitulated human disease. In particular, a model that recapitulates the most important single histotype, high-grade serous ovarian cancer, has remained elusive. Mouse transgenic and knockout studies are likely to substantially benefit from an improved understanding of the key driver mutations, such as the loss of ARID1A in clear cell ovarian cancer and the importance of secretory cells of the fallopian tube in high-grade serous ovarian cancer. Recent reports of fallopian tube-based model systems will help to define the physiology and susceptibility of this epithelium to transformation7,80.

Now that it is appreciated that clear cell, mucinous, low-grade endometrioid and high-grade serous ovarian cancers are as molecularly distinct as, for example, breast and renal cancers, researchers must pay strict attention to the human cell line models that they use to explore different aspects of the disease. Comparative molecular studies that fail to recognize the histopathological origins of the ovarian cancer cell lines used are no longer appropriate. Currently, the origins of many of the ovarian cancer cell lines are poorly defined — the field would benefit enormously from the creation of new cell lines that reflect the different histotypes and molecular subtypes of ovarian cancer.

It is also timely to consider *in vitro* models that are more appropriate than the two-dimensional growth of cells on plastic. Three-dimensional cell culture systems that mimic the peritoneal microenvironment have provided important insights into ovarian cancer biology^{81,82}.

Quality of life and symptom benefit

It is essential that measures of quality of life and symptom benefit are included with response and survival rates as primary end points in clinical trials that investigate palliative treatments. The primary end point of most clinical trials has traditionally been objective response and survival. Most patients relapse after first-line treatment; response rates and time to progression typically diminish with each recurrence. Although there are numerous studies of palliative chemotherapy regimes in platinum-resistant ovarian cancer, there is scant evidence to confirm that these

treatments are truly palliative and improve symptom control⁸³. Attempts to address this important question with the currently available quality-of-life questionnaires have not been successful, and toxicity data completed by physicians may give a false impression of the patient's experience. Robust instruments are needed that measure the effect of palliative chemotherapy on symptom control given that this should be the major aim of treatment. Treatment should be stratified in accordance to prognosis, with more emphasis placed on minimizing side effects and avoiding inappropriate therapy in patients with a low likelihood of benefit. Prognostic indices are required to better categorize patients with recurrent ovarian cancer into poor, intermediate and good subsets rather than the current approach of grouping patients together.

Conclusion

The past 5 years have seen an explosion in our understanding of the heterogeneity of ovarian cancer. This comes with an emphatic commitment to changing the way that clinical trials in ovarian cancer should be designed. Treatment of ovarian cancer will benefit from the careful alignment of target, drug, patient and trial design. The greatest challenge will be to craft combinations of therapy that result in years of improved survival. There is an outstanding level of cooperation and willingness to share ideas among those in the ovarian cancer research field, as exemplified by the HHMT meeting. A high level of cooperation bodes well for the women whose lives we hope to improve.

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- McGuire, W. P. Maintenance therapy for ovarian cancer: of Helsinki and Hippocrates. J. Clin. Oncol. 27, 4633–4634 (2009).
- Omura, G. *et al.* A randomized trial of cyclophosphamide and doxorubicin with or without cisplatin in advanced ovarian carcinoma. A Gynecologic Oncology Group Study. *Cancer* 57, 1725–1730 (1986).

- Coleman, M. P. *et al.* Cancer survival in Australia, Canada, Denmark, Norway, Sweden, and the UK, 1995–2007 (the International Cancer Benchmarking Partnership): an analysis of population-based cancer registry data. *Lancet* **377**, 127–138 (2011).
- Kurman, R. J. & Shih le, M. The origin and pathogenesis of epithelial ovarian cancer: a proposed unifying theory. *Am. J. Surg. Pathol.* 34, 433–443 (2010).
- Piek, J. M. *et al.* Dysplastic changes in prophylactically removed Fallopian tubes of women predisposed to developing ovarian cancer. *J. Pathol.* **195**, 451–456 (2001).
- Lee, Y. *et al.* A candidate precursor to serous carcinoma that originates in the distal fallopian tube. *J. Pathol.* 211, 26–35 (2007).
- Levanon, K. *et al.* Primary *ex vivo* cultures of human fallopian tube epithelium as a model for serous ovarian carcinogenesis. *Oncogene* 29, 1103–1113 (2010).
- Kuo, K. T. *et al.* Frequent activating mutations of PIK3CA in ovarian clear cell carcinoma. *Am. J. Pathol.* **174**, 1597–601 (2009).
- Jones, S. *et al.* Frequent mutations of chromatin remodeling gene ARID1A in ovarian clear cell carcinoma. *Science* **330**, 228–231 (2010).
 Wiegand, K. C. *et al.* ARID1A mutations in
- Wiegand, K. C. *et al.* ARID1A mutations in endometriosis-associated ovarian carcinomas. *N. Engl. J. Med.* 363, 1532–1543 (2010).
- Lee, K. R. & Young, R. H. The distinction between primary and metastatic mucinous carcinomas of the ovary: gross and histologic findings in 50 cases. *Am. J. Surg. Pathol.* 27, 281–292 (2003).
- Zaino, R. J. *et al.* Advanced stage mucinous adenocarcinoma of the ovary is both rare and highly lethal: a Gynecologic Oncology Group study. *Cancer* 117, 554–562 (2011).
- Kelemen, L. E. & Kobel, M. Mucinous carcinomas of the ovary and colorectum: different organ, same dilemma. *Lancet Oncol.* 25 May 2011 (doi:10.1016/ S1470-2045(11)70058-4).
- Kobel, M. *et al.* Ovarian carcinoma subtypes are different diseases: implications for biomarker studies. *PLoS Med.* 5, e232 (2008).
- Kobel, M. *et al.* A limited panel of immunomarkers can reliably distinguish between clear cell and high-grade serous carcinoma of the ovary. *Am. J. Surg. Pathol.* 33, 14–21 (2009).
- Madore, J. *et al.* Characterization of the molecular differences between ovarian endometrioid carcinoma and ovarian serous carcinoma. *J. Pathol.* 220, 392–400 (2010).
- Bowtell, D. D. The genesis and evolution of high-grade serous ovarian cancer. *Nature Rev. Cancer* 10, 803–808 (2010).
- Ho, C. L., Kurman, R. J., Dehari, R., Wang, T. L. & Shih Ie, M. Mutations of BRAF and KRAS precede the development of ovarian serous borderline tumors. *Cancer Res.* 64, 6915–6918 (2004).
- Tothill, R. W. *et al.* Novel molecular subtypes of serous and endometroid ovarian cancer linked to clinical outcome. *Clin. Cancer Res.* 14, 5198–5208 (2008).
- Ahmed, A. A. *et al.* Driver mutations in TP53 are ubiquitous in high grade serous carcinoma of the ovary. *J. Pathol.* **221**, 49–56 (2010).
- Zorn, K. K. *et al.* Gene expression profiles of serous, endometrioid, and clear cell subtypes of ovarian and endometrial cancer. *Clin. Cancer Res.* **11**, 6422–6430 (2005).
- Anglesio, M. S. *et al.* IL6-STAT3-HIF signaling and therapeutic response to the angiogenesis inhibitor sunitinib in ovarian clear cell cancer. *Clin. Cancer Res.* 17, 2538–2548 (2011).
- Kobel, M. *et al.* Diagnosis of ovarian carcinoma cell type is highly reproducible: a transcanadian study. *Am. J. Surg. Pathol.* 34, 984–993 (2010)
- Am. J. Surg. Pathol. 34, 984–993 (2010).
 24. Glimelius, B. & Lahn, M. Window-of-opportunity trials to evaluate clinical activity of new molecular entities in oncology. Ann. Oncol. 22, 1717–1725 (2011).
- Kyriazi, S., Kaye, S. B. & deSouza, N. M. Imaging ovarian cancer and peritoneal metastases-current and emerging techniques. *Nature Rev. Clin. Oncol.* 7, 381–393 (2010).
- He, W. *et al.* Quantitation of circulating tumor cells in blood samples from ovarian and prostate cancer patients using tumor-specific fluorescent ligands. *Int. J. Cancer* **123**, 1968–1973 (2008).
- Shih le, M. & Kurman, R. J. Ovarian tumorigenesis: a proposed model based on morphological and molecular genetic analysis. *Am. J. Pathol.* 164, 1511–1518 (2004).

- Menon, U. *et al.* Sensitivity and specificity of multimodal and ultrasound screening for ovarian cancer, and stage distribution of detected cancers: results of the prevalence screen of the UK Collaborative Trial of Ovarian Cancer Screening (UKCTOCS). *Lancet Oncol.* **10**, 327–340 (2009).
- Brown, P. O. & Palmer, C. The preclinical natural history of serous ovarian cancer: defining the target for early detection. *PLoS Med.* 6, e1000114 (2009).
- Buys, S. S. *et al.* Effect of screening on ovarian cancer mortality: the Prostate, Lung, Colorectal and Ovarian (PLCO) Cancer Screening Randomized Controlled Trial. JAMA 305, 2295–2303 (2011).
- Hogg, R. & Friedlander, M. Biology of epithelial ovarian cancer: implications for screening women at high genetic risk. *J. Clin. Oncol.* 22, 1315–1327 (2004).
- Domchek, S. M. *et al.* Association of risk-reducing surgery in BRCA1 or BRCA2 mutation carriers with cancer risk and mortality. *JAMA* **304**, 967–975 (2010).
- Rebbeck, T. R., Kauff, N. D. & Domchek, S. M. Metaanalysis of risk reduction estimates associated with risk-reducing salpingo-oophorectomy in BRCA1 or BRCA2 mutation carriers. *J. Natl Cancer Inst.* **101**, 80–87 (2009).
- Dietl, J. & Wischhusen, J. The forgotten fallopian tube. *Nature Rev. Cancer* 11, 227; author reply 227 (2011).
- Bolton, K. L. *et al.* Common variants at 19p13 are associated with susceptibility to ovarian cancer. *Nature Genet.* 42, 880–884 (2010).
- Song, H. *et al.* A genome-wide association study identifies a new ovarian cancer susceptibility locus on 9p22.2. *Nature Genet.* 41, 996–1000 (2009).
- Gorringe, K. L. *et al.* Copy number analysis identifies novel interactions between genomic loci in ovarian cancer. *PLoS ONE* 5, e11408 (2010).
- Gorringe, K. L. *et al.* High-resolution single nucleotide polymorphism array analysis of epithelial ovarian cancer reveals numerous microdeletions and amplifications. *Clin. Cancer Res.* **13**, 4731–4739 (2007).
- The Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature* 474, 609–615 (2011).
- Farley, J. *et al.* Cyclin E expression is a significant predictor of survival in advanced, suboptimally debulked ovarian epithelial cancers: a Gynecologic Oncology Group study. *Cancer Res.* 63, 1235–1241 (2003).
- Etemadmoghadam, D. et al. Amplicon-dependent CCNE1 expression is critical for clonogenic survival after cisplatin treatment and is correlated with 20q11 gain in ovarian cancer. PLoS ONE 5, e15498 (2010).
- Tan, D. S. *et al.* Genomic analysis reveals the molecular heterogeneity of ovarian clear cell carcinomas. *Clin. Cancer Res.* **17**, 1521–1534 (2011).
- Shah, S. P. *et al.* Mutation of FOXL2 in granulosa-cell tumors of the ovary. *N. Engl. J. Med.* **360**, 2719–2729 (2009).
- Press, J. Z. *et al.* Ovarian carcinomas with genetic and epigenetic BRCA1 loss have distinct molecular abnormalities. *BMC Cancer* 8, 17 (2008).
- Turner, N., Tutt, A. & Ashworth, A. Hallmarks of BRCAness' in sporadic cancers. *Nature Rev. Cancer* 4, 814–819 (2004).
- Banerjee, S., Kaye, S. B. & Ashworth, A. Making the best of PARP inhibitors in ovarian cancer. *Nature Rev. Clin. Oncol.* 7, 508–519 (2010).
- Ledermann, J. A. *et al.* Phase II randomized placebocontrolled study of olaparib (AZD2281) in patients with platinum-sensitive relapsed serous ovarian cancer (PSR SOC). *J. Clin. Oncol. Abstr.* 29, 5003 (2011).
- Mukhopadhyay, A. *et al.* Development of a functional assay for homologous recombination status in primary cultures of epithelial ovarian tumor and correlation with sensitivity to poly(ADP-ribose) polymerase inhibitors. *Clin. Cancer Res.* 16, 2344–2351 (2010).
- Martin, S. A. *et al.* Methotrexate induces oxidative DNA damage and is selectively lethal to tumour cells with defects in the DNA mismatch repair gene MSH2. *EMBO Mol. Med.* 1, 323–337 (2009).
- 50. Martin, S. A. *et al.* DNA polymerases as potential therapeutic targets for cancers deficient in the DNA

mismatch repair proteins MSH2 or MLH1. *Cancer Cell* **17**, 235–248 (2010).

- Cooke, S. L. *et al.* Intra-tumour genetic heterogeneity and poor chemoradiotherapy response in cervical cancer. *Br. J. Cancer* **104**, 361–368 (2011).
- Cooke, S. L. et al. Genomic analysis of genetic heterogeneity and evolution in high-grade serous ovarian carcinoma. Oncogene 29, 4905–4913 (2010).
- O'Malley, D. M. *et al.* Addition of bevacizumab to weekly pacitaxel significantly improves progressionfree survival in heavily pretreated recurrent epithelial ovarian cancer. *Gynecol. Oncol.* **121**, 269–272 (2011).
- Ebos, J. M. & Kerbel, R. S. Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. *Nature Rev. Clin. Oncol.* 8, 210–221 (2011).
- Jubb, A. M. & Harris, A. L. Biomarkers to predict the clinical efficacy of bevacizumab in cancer. *Lancet Oncol.* 11, 1172–1183 (2010).
- Goodell, V. et al. Antibody immunity to the p53 oncogenic protein is a prognostic indicator in ovarian cancer. J. Clin. Oncol. 24, 762–768 (2006).
- Schlienger, K. *et al.* TRANCE- and CD40 ligandmatured dendritic cells reveal MHC class I-restricted T cells specific for autologous tumor in late-stage ovarian cancer patients. *Clin. Cancer Res.* 9, 1517–1527 (2003).
- Zhang, L. *et al*. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N. Engl. J. Med.* 348, 203–213 (2003).
- Curiel, T. J. *et al.* Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nature Med.* **10**, 942–949 (2004).
- Sato, E. et al. Intraepithelial CD8⁺ tumor-infiltrating lymphocytes and a high CD8⁺/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer. Proc. Natl Acad. Sci. USA 102, 18538–18543 (2005).
- Kryczek, I. *et al.* Relationship between B7-H4, regulatory T cells, and patient outcome in human ovarian carcinoma. *Cancer Res.* 67, 8900–8905 (2007).
- Kulbe, H. *et al.* The inflammatory cytokine TNF-a generates an autocrine tumour-promoting network in epithelial ovarian cancer cells. *Cancer Res.* 67, 585–592 (2007).
- Mantovani, A., Allavena, P., Sica, A. & Balkwill, F. Cancer-related inflammation. *Nature* 454, 436–444 (2008).
- Facciabene, X. *et al.* Tumour hypoxia promotes tolerance and angiogenesis via CCL28 and Treg cells. *Nature* 475, 226–230 (2011).
- Charles, K. A. *et al.* The tumor-promoting actions of TNF-a involve TNFR1 and IL-17 in ovarian cancer in mice and humans. *J. Clin. Invest.* **119**, 3011–3023 (2009).
- Hodi, F. S. *et al.* Biologic activity of cytotoxic T lymphocyte-associated antigen 4 antibody blockade in previously vaccinated metastatic melanoma and ovarian carcinoma patients. *Proc. Natl Acad. Sci. USA* **100**, 4712–4714 (2003).
- Hodi, F. S. *et al.* Immunologic and clinical effects of antibody blockade of cytotoxic T lymphocyteassociated antigen 4 in previously vaccinated cancer patients. *Proc. Natl Acad. Sci. USA* **105**, 3005–3010 (2008).
- Coward, J. *et al.* Interleukin-6 as a therapeutic target in human ovarian cancer. *Clin. Cancer Res.* 27 Jul 2011 (doi:10.1158/1078-0432. CCR-11-0945).
- Balkwill, F. & Mantovani, A. Cancer and inflammation: implications for pharmacology and therapeutics. *Clin. Pharmacol. Ther.* 87, 401–406 (2010).
- Choi, S. *et al.* Relapse in children with acute lymphoblastic leukemia involving selection of a preexisting drug-resistant subclone. *Blood* 110, 632–639 (2007).
- Roche-Lestienne, C. *et al.* Several types of mutations of the Abl gene can be found in chronic myeloid leukemia patients resistant to STI571, and they can pre-exist to the onset of treatment. *Blood* **100**, 1014–1018 (2002).
- Yachida, S. *et al.* Distant metastasis occurs late during the genetic evolution of pancreatic cancer. *Nature* 467, 1114–1117 (2010).

- Campbell, P. J. *et al.* The patterns and dynamics of genomic instability in metastatic pancreatic cancer. *Nature* 467, 1109–1113 (2010).
- Stronach, E. A. *et al.* HDAC4-regulated STAT1 activation mediates platinum resistance in ovarian cancer. *Cancer Res.* 71, 4412–4422 (2011).
- Baggerly, K. A., Morris, J. S., Edmonson, S. R. & Coombes, K. R. Signal in noise: evaluating reported reproducibility of serum proteomic tests for ovarian cancer. *J. Natl Cancer Inst.* **97**, 307–309 (2005).
- Connolly, D. C. *et al.* Female mice chimeric for expression of the simian virus 40 TAg under control of the MISIIR promoter develop epithelial ovarian cancer. *Cancer Res.* 63, 1389–1397 (2003).
- Wu, R. *et al.* Mouse model of human ovarian endometrioid adenocarcinoma based on somatic defects in the Wnt/B-catenin and P13K/Pten signaling pathways. *Cancer Cell* **11**, 321–333 (2007).
- Xing, D. & Orsulic, S. A genetically defined mouse ovarian carcinoma model for the molecular characterization of pathway-targeted therapy and tumor resistance. *Proc. Natl Acad. Sci. USA* **102**, 6936–6941 (2005).
- Orsulic, S. *et al.* Induction of ovarian cancer by defined multiple genetic changes in a mouse model system. *Cancer Cell* 1, 53–62 (2002).
- Karst, A. M., Levanon, K. & Drapkin, R. Modeling high-grade serous ovarian carcinogenesis from the fallopian tube. *Proc. Natl Acad. Sci. USA* **108**, 7547–7552 (2011).
- Kenny, H. A., Kaur, S., Coussens, L. M. & Lengyel, E. The initial steps of ovarian cancer cell metastasis are mediated by MMP-2 cleavage of vitronectin and fibronectin. J. Clin. Invest. 118, 1367–1379 (2008).
- Iwanicki, M. P. *et al.* Ovarian cancer spheroids use myosin-generated force to clear the mesothelium. *Cancer Discov.* 1, 144–157 (2011).
- Al-Barrak, J. et al. Exploring palliative treatment outcomes in women with advanced or recurrent ovarian clear cell carcinoma. *Gynecol. Oncol.* 122, 107–110 (2011).
- Khalique, L. *et al.* The clonal evolution of metastases from primary serous epithelial ovarian cancers. *Int. J. Cancer* **124**, 1579–1586 (2009).
- Khalique, L. *et al.* Genetic intra-tumour heterogeneity in epithelial ovarian cancer and its implications for molecular diagnosis of tumours. *J. Pathol.* 211, 286–295 (2007).
- Norquist, B. et al. Secondary somatic mutations restoring BRCA1/2 predict chemotherapy resistance in hereditary ovarian carcinomas. J. Clin. Oncol. 29, 3008–3015 (2011).

Acknowledgements

The authors would like to thank the Ovarian Cancer Action, its Chair, A. Kaye, its staff and its many supporters, without whose drive and generosity the Helene Harris Memorial Trust meetings would not be possible. The authors also thank K. Swenerton, Cheryl Brown Outcomes Unit, British Columbia Cancer Agency, Canada, for providing some of the data in Figure 1 and for the concept of Figure 3. Further acknowledgements and sources of funding for some of the authors of this article can be found in the <u>Supplementary information S1</u> (text) (see Further information).

Competing interests statement

The authors declare competing financial interests. See $\underline{\text{Web}}$ version for details.

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FURTHER INFORMATION

David D. Bowtell's homepage: <u>http://www.petermac.org/</u> <u>Research/CancerGeneticsGenomics</u>

Frances R. Balkwill's homepage: http://www.bci.qmul.ac.uk/ research/centre-profiles/cancer-a-inflammation.html Helene Harris Memorial Trust International Forum on Ovarian Cancer: http://www.ovarian.org.uk/aboutus/ research/meetings.asp

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