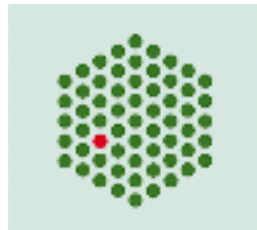


Digital Imaging

Data Handling

Basic Image Processing

Stefan Terjung
Advanced Light Microscopy Facility



EMBL

European Molecular Biology Laboratory

Overview

Part I:

- File formats (data storage)
- Programs for
image viewing / processing / representation

Part II:

- Basic Image Processing (using ImageJ)

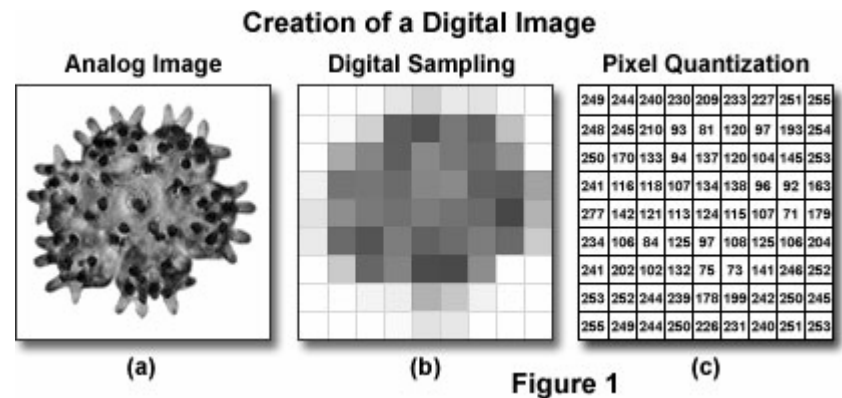
File Formats – data storage

- Lossless image formats
- Lossy compression formats
- Custom formats (microscope companies)
- Sequence vs. single image per file
- 8bit, 12bit, 16bit, 32bit, RGB

- Storage:
 - Always have at least 1 copy of the data
 - Very suitable: file servers (automatic backup)

Lossless Image Formats

- TIFF (with or without compression)
- BMP (windows uncompressed)
- GIF (graphics interchange format)
- PNG (portable network graphics)
- Raw data
- 'text image'



Microscopy Primer

<http://micro.magnet.fsu.edu/primer>

Image Format: TIFF

- Tag Image File Format
 - Image header with flexible set of ‘tags’ which can be used to store e.g. microscopic settings
- Flexible in color space and bit depth
 - Microscopy: grayscale 8bit, 16 bit (12bit data)
 - Color (e.g. Overlay): RGB (red green blue 8bit each)
 - Quantification: 32bit (floating point values)
- Always lossless: Uncompressed or compressed
- Multiple images possible in one file

Lossless image compression

- Redundant information can be compressed
- Identical information can be grouped
- Repeated patterns can be used to compress data size

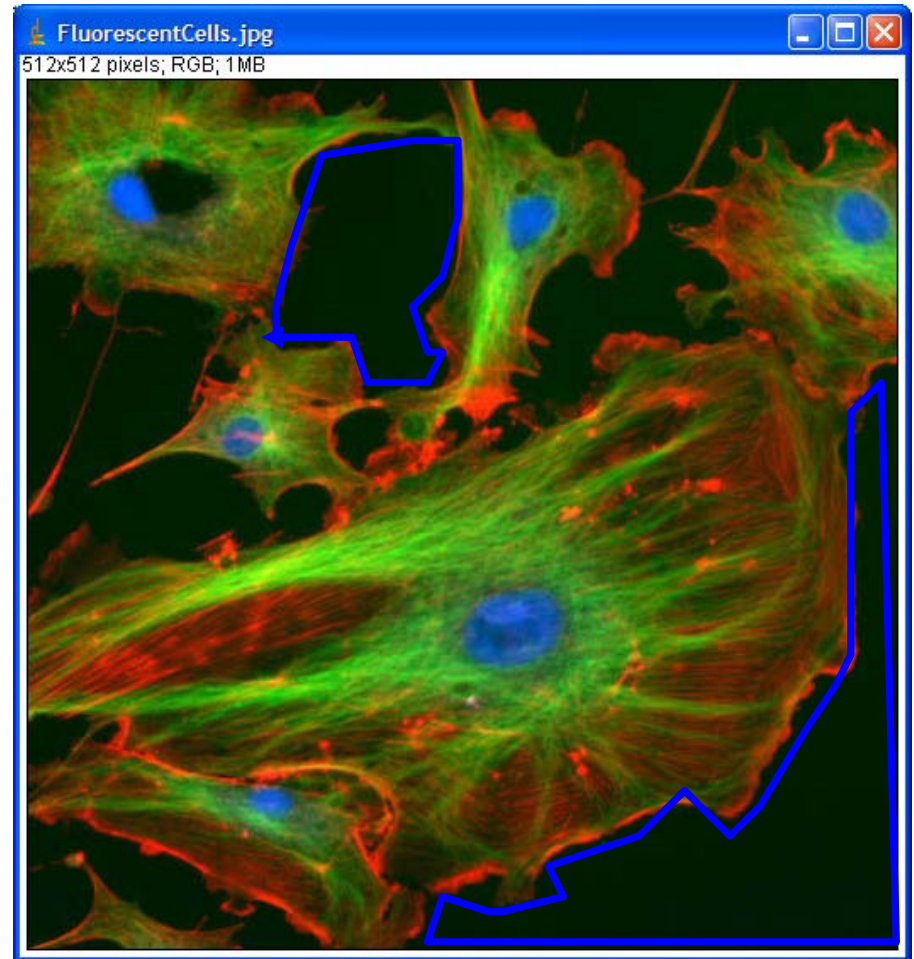
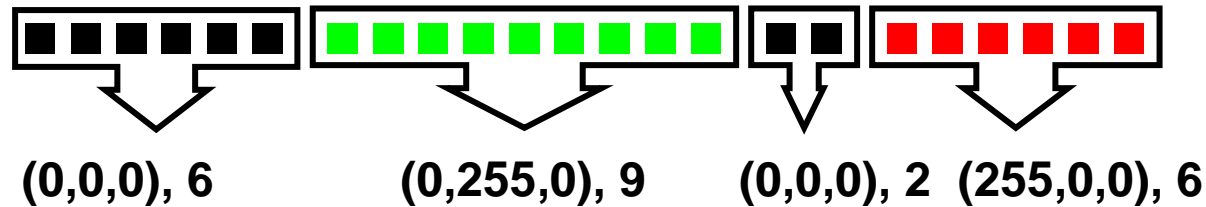


Image Compression: TIFF

- Run Length Coding (RLE): first number describes the color, the second the number of following pixels having the same color.



- LZW (Lempel-Ziv-Welch): Find repetitive patterns of values and give them a number which points to an entry of a „dictionary“ (LUT).

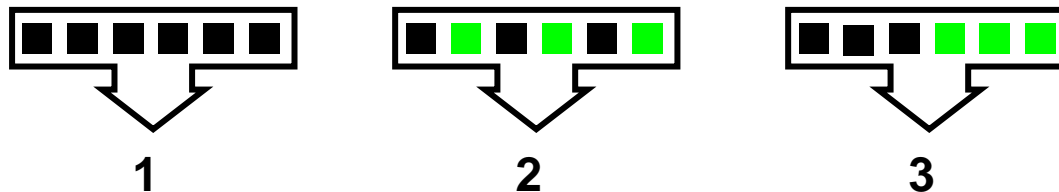
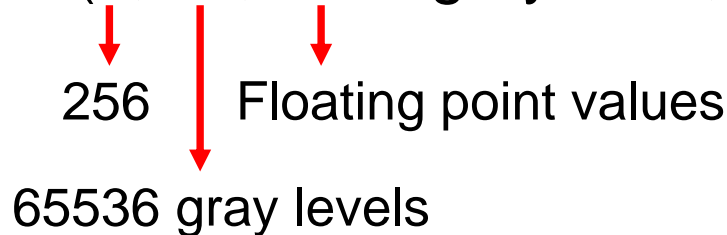


Image Compression: TIFF

Pros:

- Extra infos can be written in the ,tags‘ (e.g. microscope data, like objective lens, voxel size)
- Most programs can read it
- Lossless
- Flexible (8, 16, 32bit grayscale, 8:8:8bit RGB)



Cons:

- Big files
- Compressed files can't be loaded by ImageJ

Lossy Image Formats

- The lossy compression algorithm takes advantage of the limitations of the human visual senses and discards information that would not be sensed by the eye. (like mp3 in audio).
- Compression level is usually flexible, but the more compressed the more information is lost and artifacts become visible by eye



Original Lena Image
(12KB size)



Lena Image, Compressed
(85% less information,
1.8KB)



Lena Image, Highly
Compressed (96% less
information, 0.56KB)

From: www.wikipedia.org

Image Compression: JPG

- Split image into color and gray-scale information (color is less important than boundaries)
→ reduce high frequency color information.
- Group pixel into 8x8 blocks and transform through discrete cosine transform...

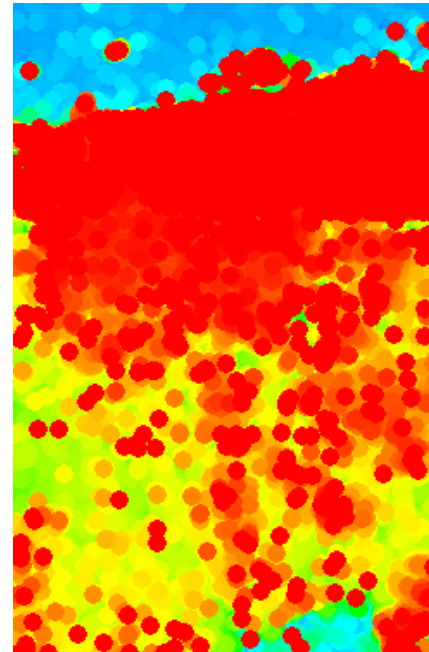
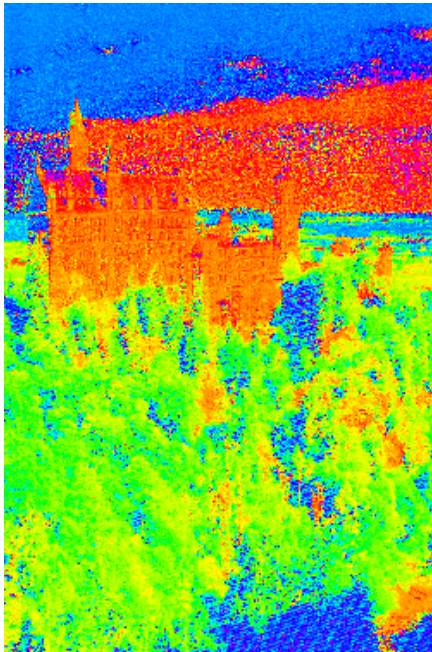


Image Compression: JPG

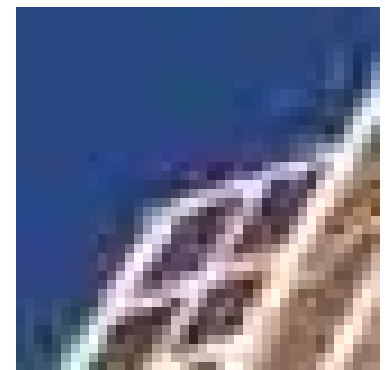
Pros:

- Small Files
- True Color
- Usable for most photos (real life) and presentations (powerpoint)



Cons:

- **Do not use for quantification !**
- „Unrelevant“ details get lost
- Every file-saving reduces the quality



Overview

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- Basic Image Processing (using ImageJ)

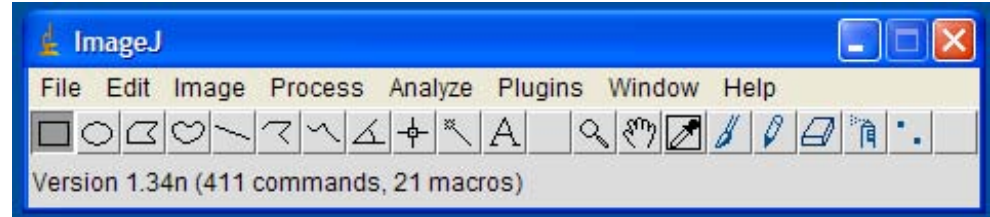
Image Viewers

- **ImageJ** (Java based, freeware, Win/MAC/Linux)
- **Irfanview** (www.irfanview.com/)
 - Freeware
 - Convert (e.g. tif → jpg)
 - Batch processing
- **ACDSee** (ACD Systems)
- **Microscope companies**
 - Zeiss Image Browser / Axiovision LE
 - Leica LCS Lite
 - Olympus Viewer

Image Representation

- **ImageJ**
- **Imaris** (Bitplane):
 - 2 floating licenses
 - installed on image processing workstations
- Photoshop, Paintshop, Illustrator, Corel Draw
(, Powerpoint)
- **Volocity** (Improvision):
 - 1 License, installed on 1 image processing workstation
- **Amira** (Mercury computer systems)
- Custom software of microscopes

Image Processing



- **ImageJ** (<http://rsb.info.nih.gov/ij/index.html>)
 - installed on all image processing workstations
 - Installation: www.embl.de/almf/html/EMBL_ImageJ.htm OR www.macbiophotonics.ca/imagej/installing_imagej.htm
 - Manual: <http://www.macbiophotonics.ca/imagej/>
 - Additional plugins: <http://rsb.info.nih.gov/ij/plugins/index.html>
- **Metamorph** (Universal Imaging),
 - installed on one image processing workstations
- Custom software of microscopes

Overview

Part I:

- File formats (data storage)
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Part II:

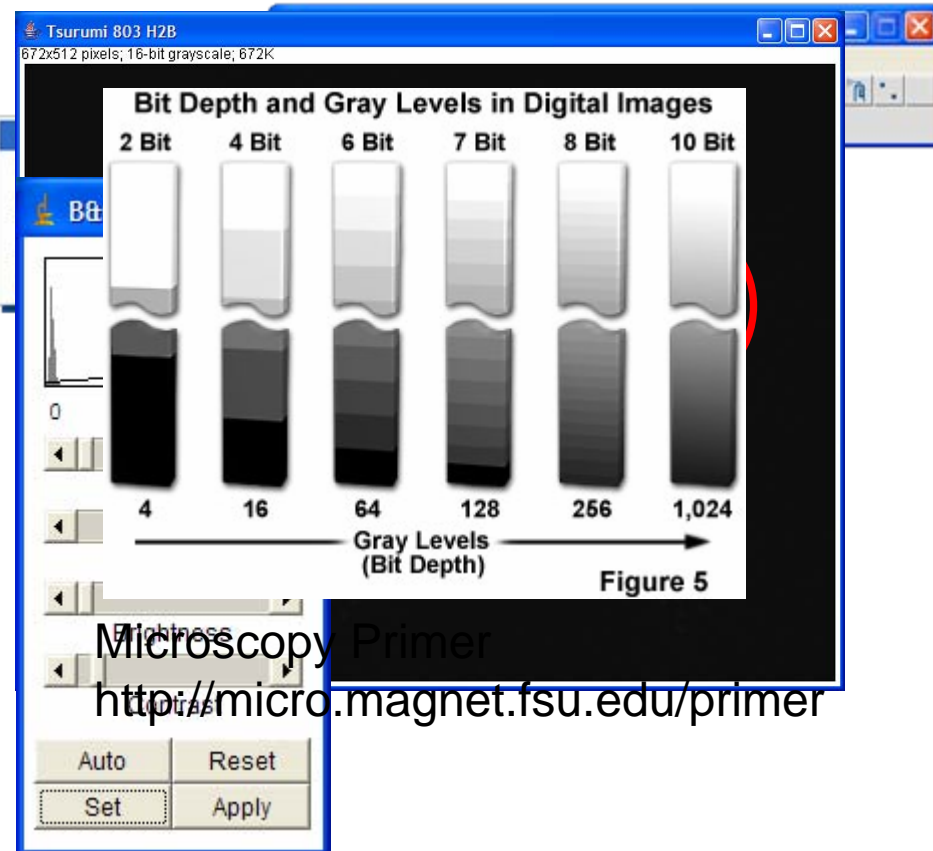
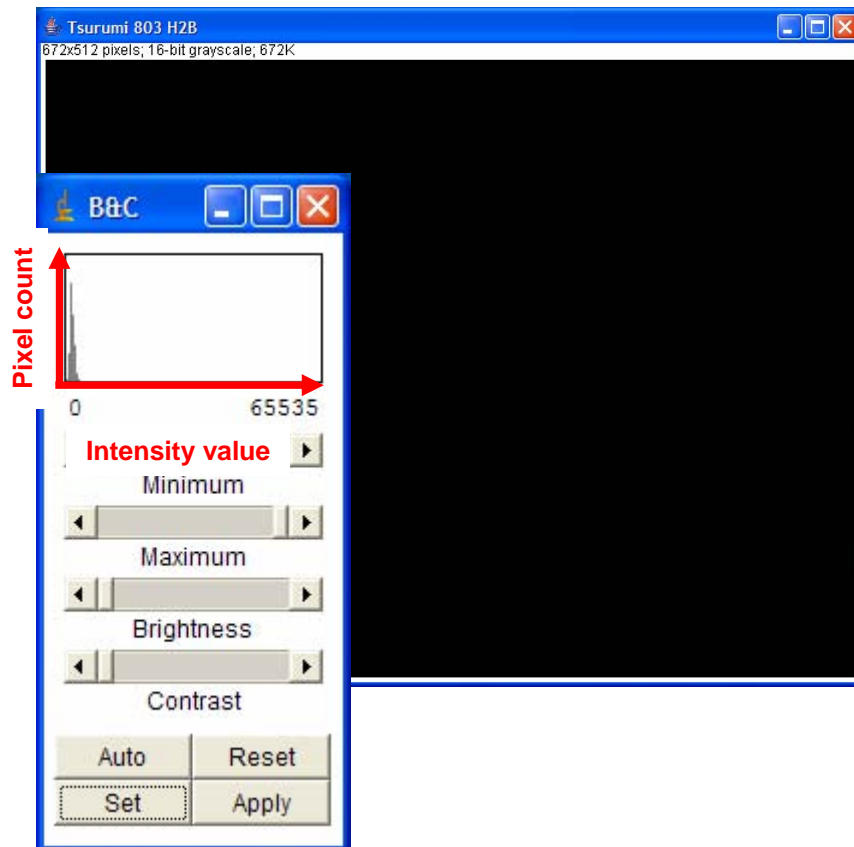
- **Basic Image Processing (using ImageJ)**

Image Processing Basics

- Visual Image Inspection
- Lookup tables (LUT) and LUT operations
- Histogram, brightness, contrast
- Filter
- Threshold
- Measurements
- Color functions

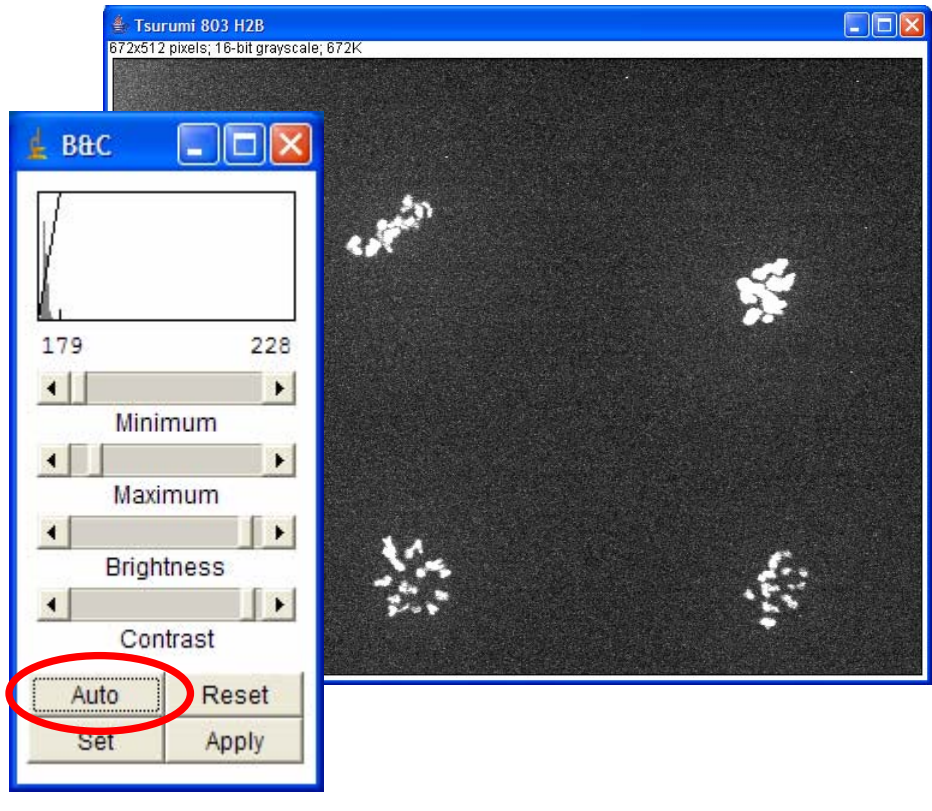
Visual Image Inspection

- Displaying images, histogram

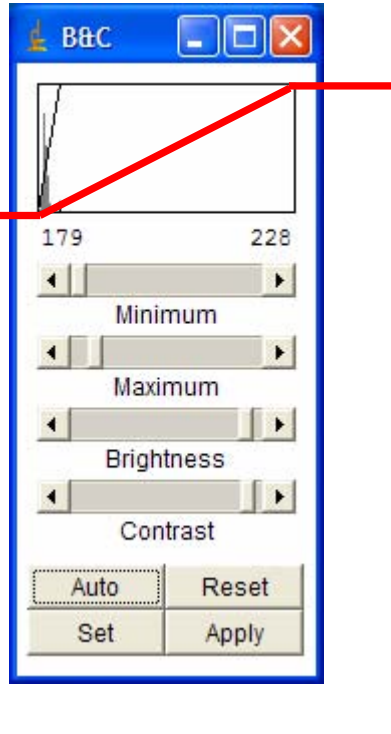


LUT operations

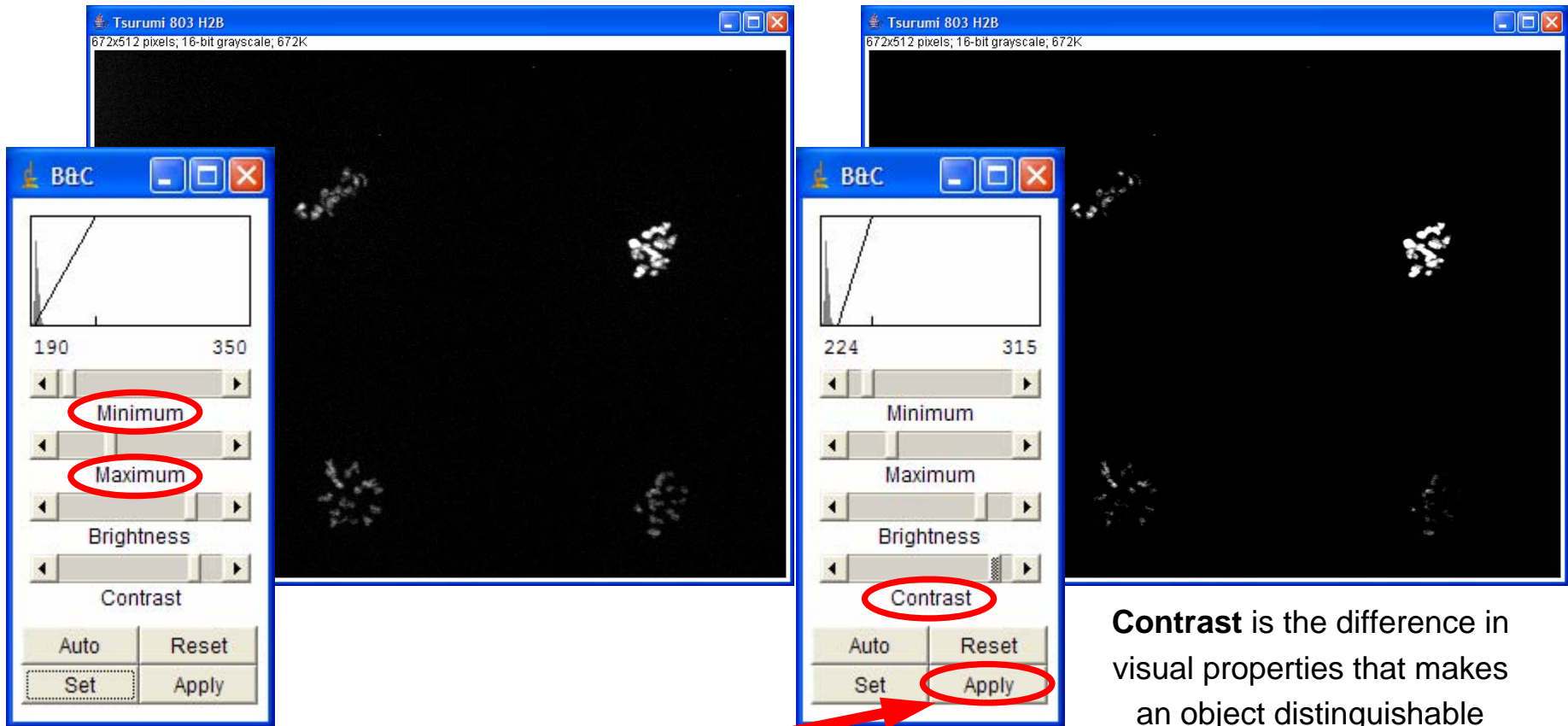
- Lookup table (LUT)
 - Displays can only show 256 gray values (8bit) per color
 - Data is unchanged, it's only "mapped" differently



Data Intensity	Displayed Intensity
0	0
...	...
179	0
180	5
181	10
...	...
226	
227	
228	255
229	255
65535	255



Brightness, Contrast

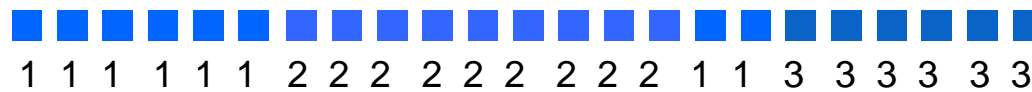


Caution: Apply modifies the data!

Contrast is the difference in visual properties that makes an object distinguishable from other objects and the background.

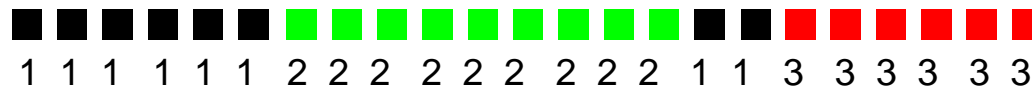
Color LUT

- The pixel contains a „pointer“ to an array, where the actual pixel values are stored



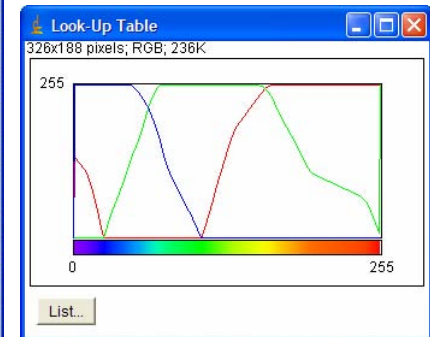
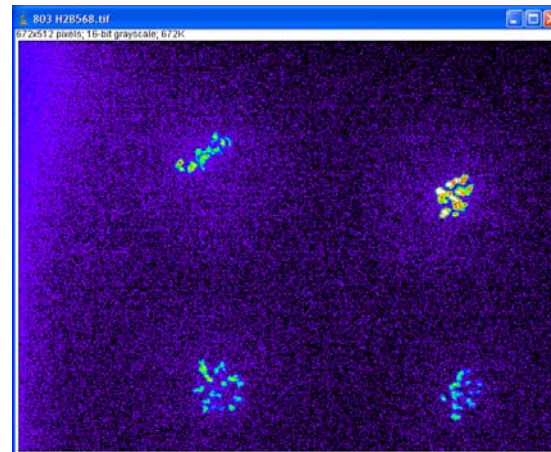
old LUT:

- 1: (0,102,255)
- 2: (51,102,255)
- 3: (10,100,200)



new LUT:

- 1: (0,0,0)
- 2: (0,255,0)
- 3: (255,0,0)

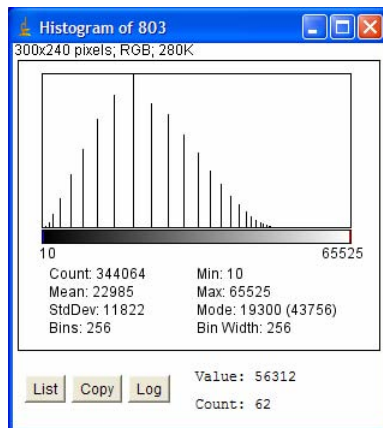
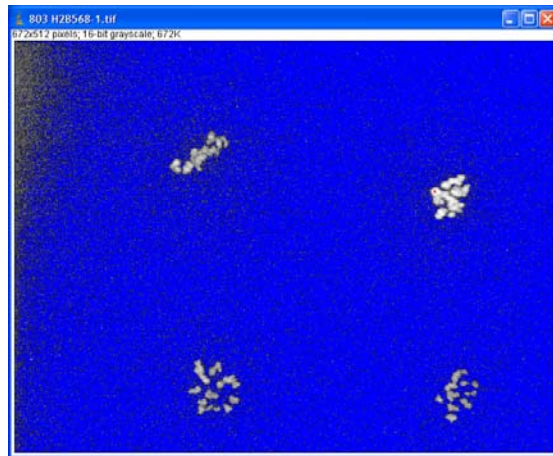


“Rainbow” LUT

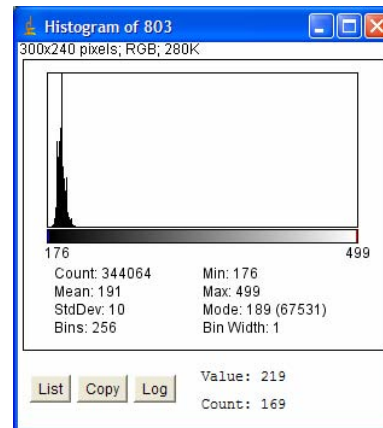
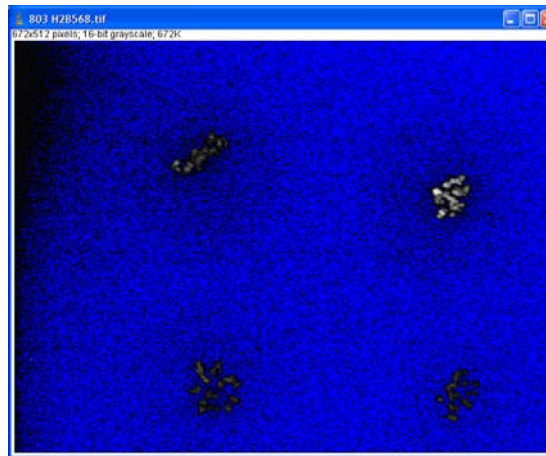
Non-linear Histogram Stretch

- Enhance contrast by (changing data):

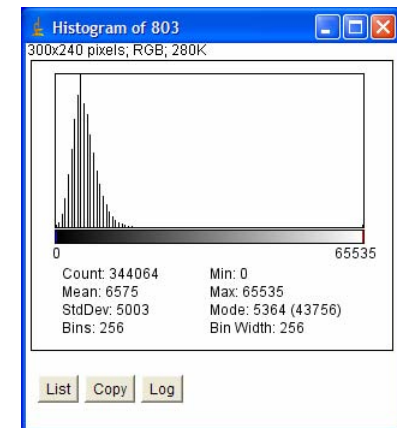
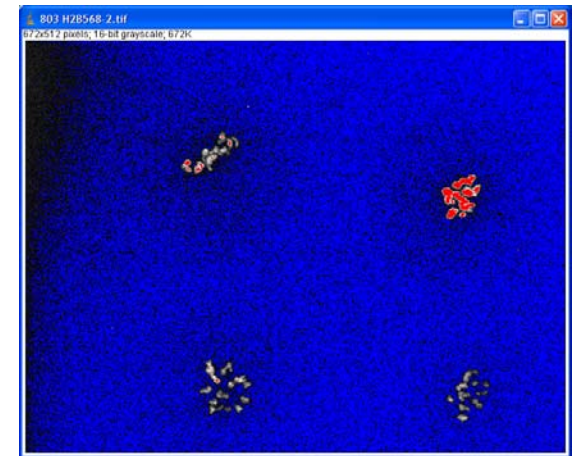
“**Equalization**” non-linear stretch
based on square root of the intensity



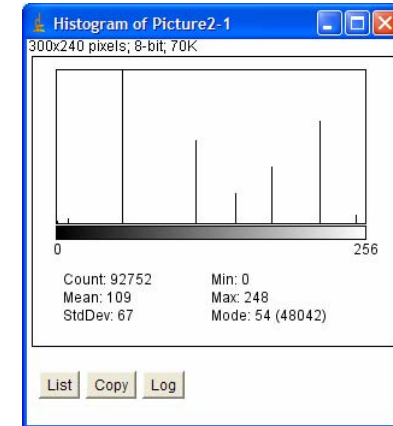
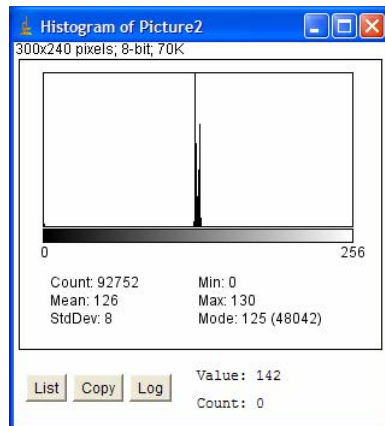
Raw data



Linear stretch
“**Normalization**”

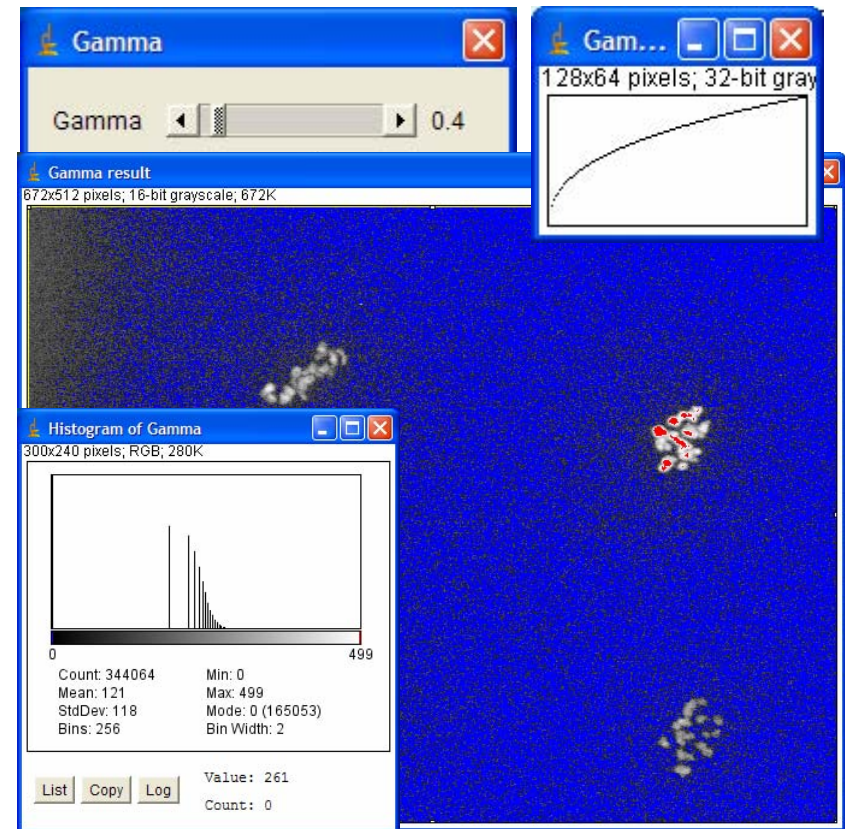
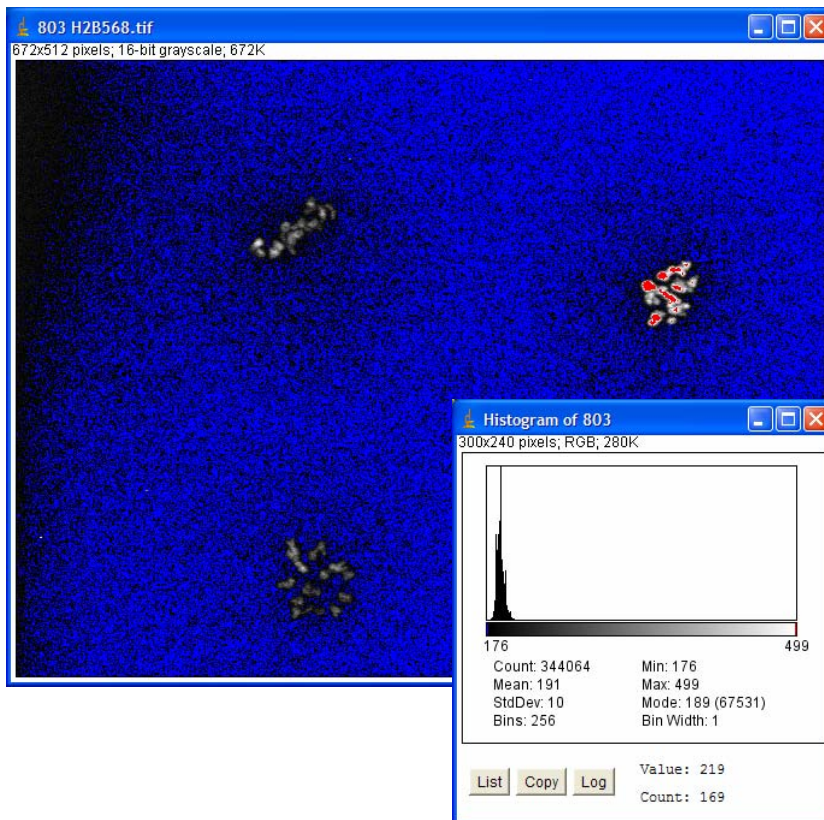


Equalization



Gamma

- Gamma is a non-linear histogram adjustment
- 8 bit images:
New intensity = $255 \times [(old\ intensity/255)^\gamma]$

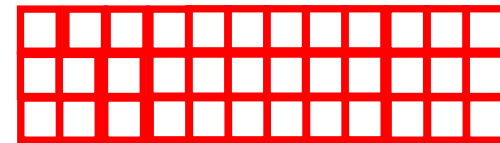
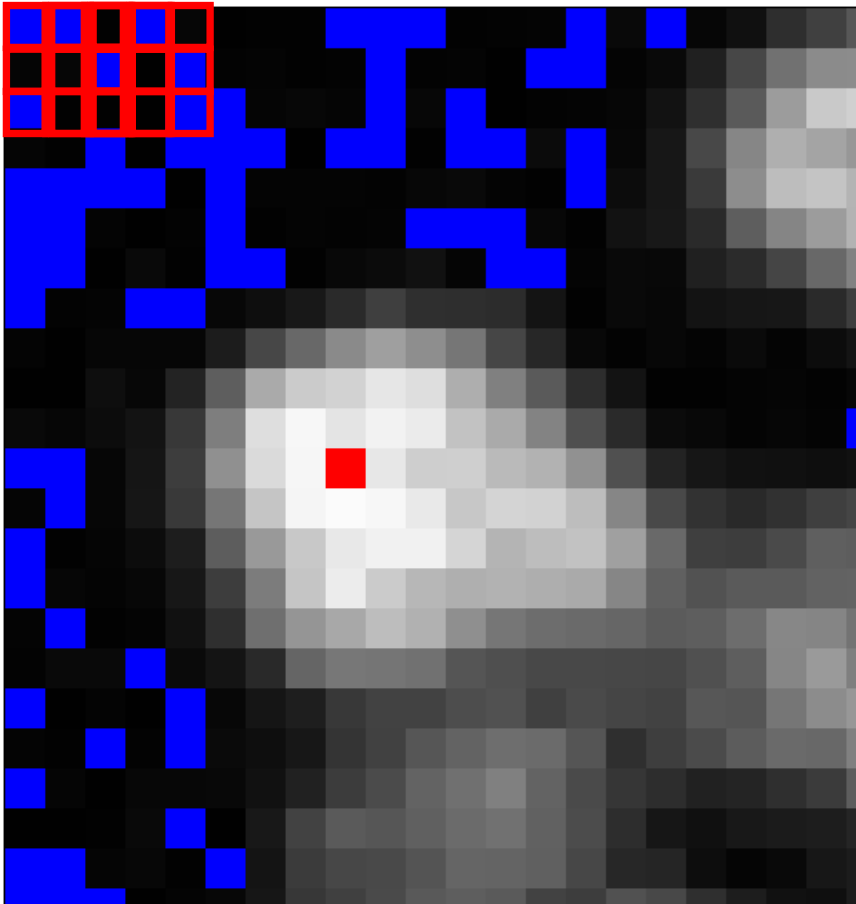
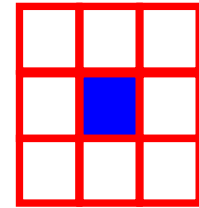


Filtering

- Image processing filters are mainly used to:
 - suppress the high frequencies in the image, *i.e.* smoothing the image, noise reduction
 - or suppress the low frequencies, *i.e.* enhancing or detecting edges in the image
- An image can be filtered either in the frequency or in the spatial domain.
 - Filtering in the frequency domain requires Fourier transform first and re-transformation after application of the filter.
 - Filtering in the spatial domain is done by convolving the image with the filterfunction.

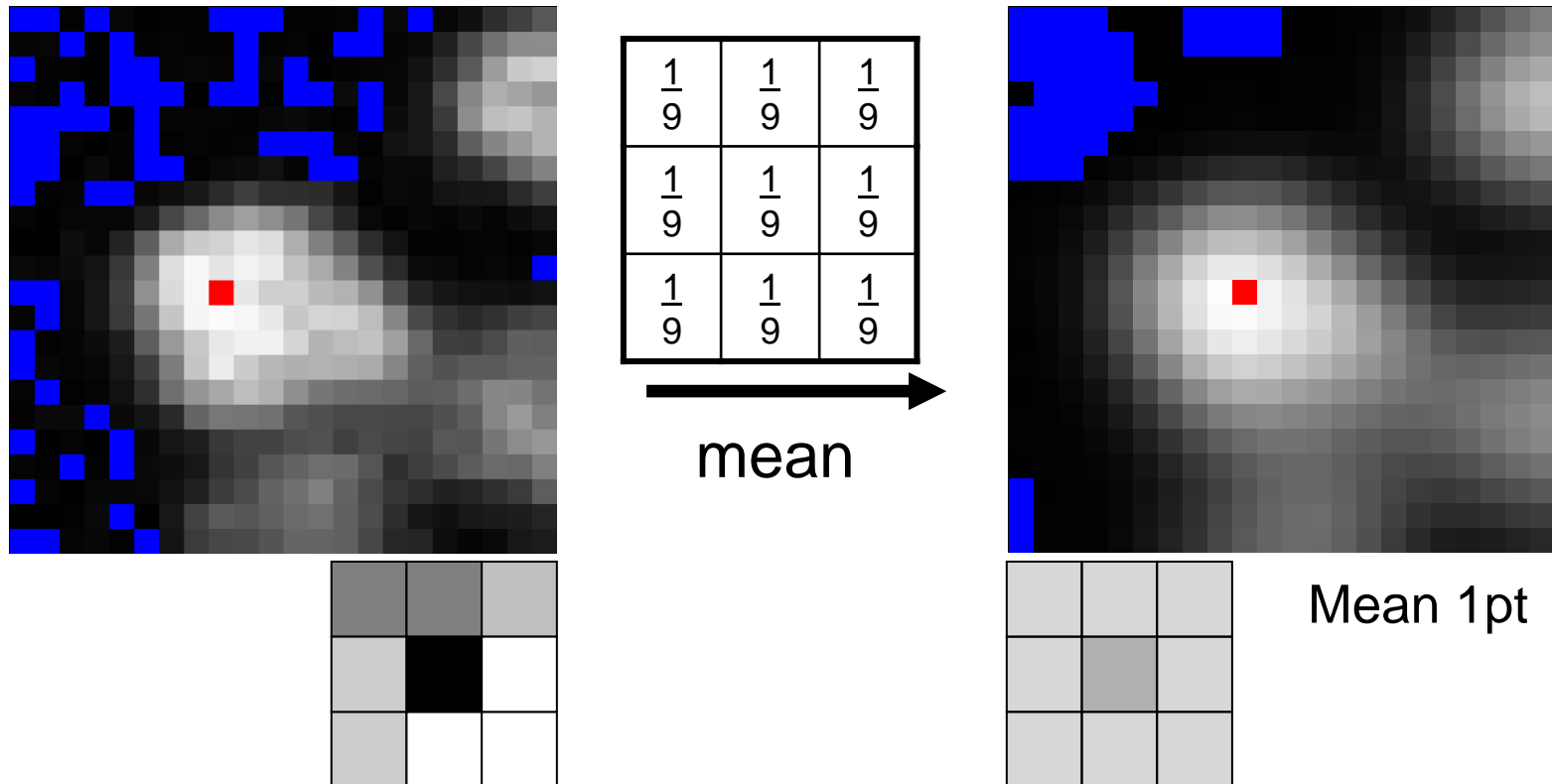
Filtering

- Shifting and multiplying a filter kernel



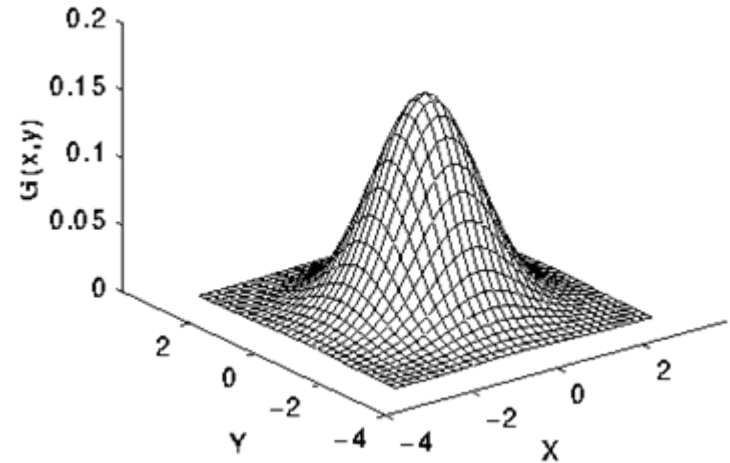
Filtered image

Noise Reduction: Mean



Noise Reduction: Gaussian

- Filtering with a gaussian bell-shaped kernel:



$$\frac{1}{16}$$

1	2	1
2	4	2
1	2	1

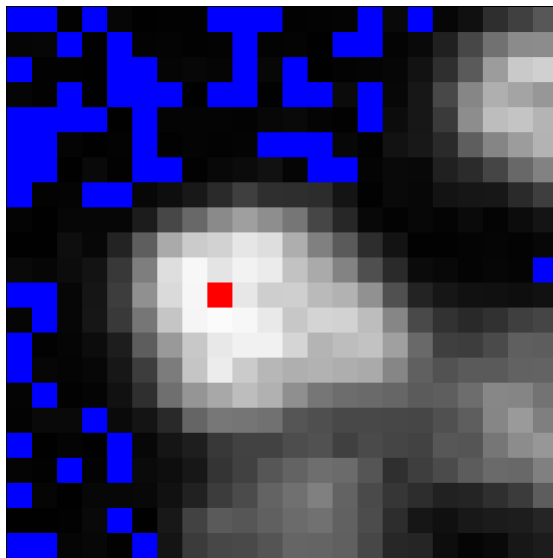
10	25	3
9	33	5
0	0	



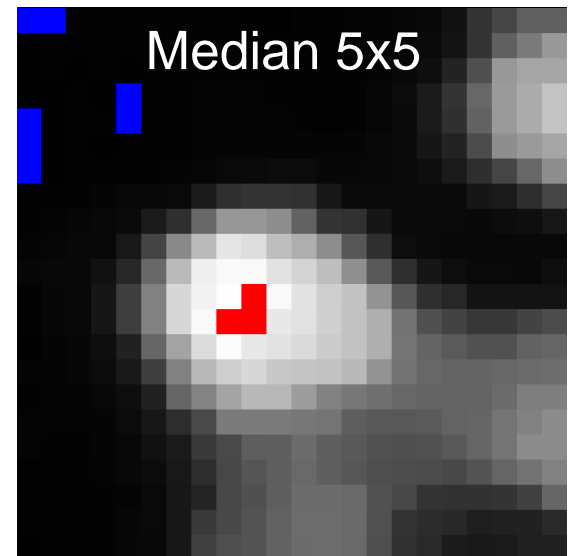
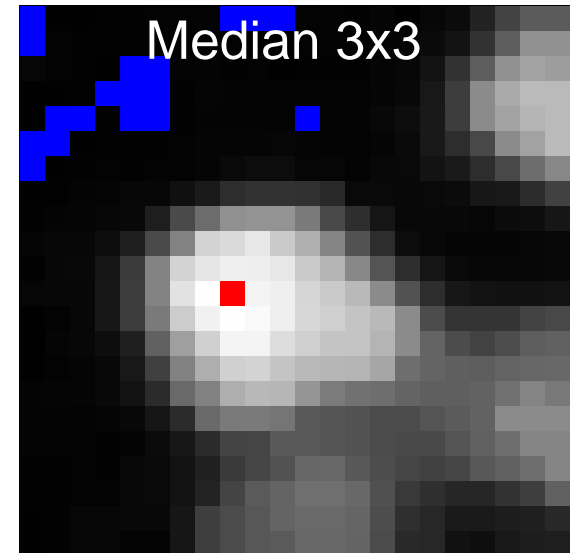
$$\frac{1}{16}$$

$$= 15$$

Noise Reduction: Median



→
median



123	125	126	130	140
122	124	126	127	135
118	120	150	125	134
119	115	119	123	133
111	116	110	120	130

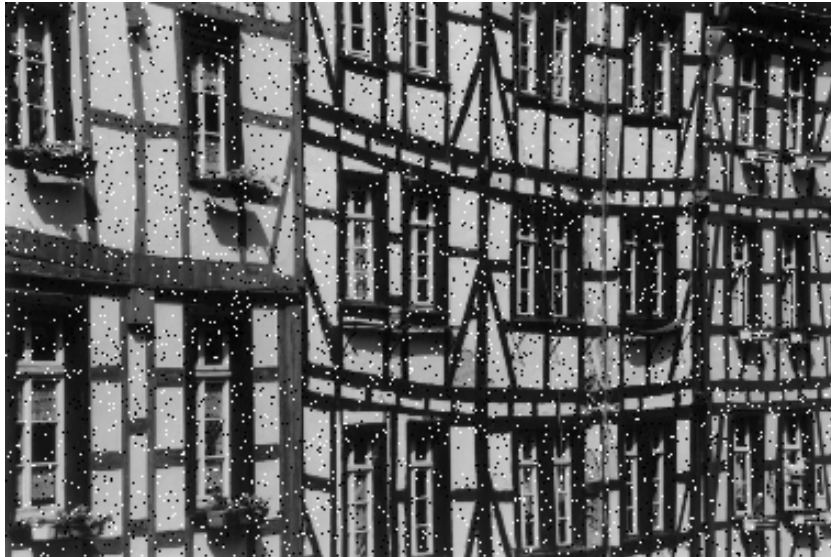
Neighbourhood values:

115, 119, 120, 123, 124,
125, 126, 127, 150

Median value: 124

Noise Reduction: Median, Mean

Median, 1pt



Mean, 1pt



Median-, Mean-, Max-, Min-Filter



Median, 5pt



Mean, 5pt



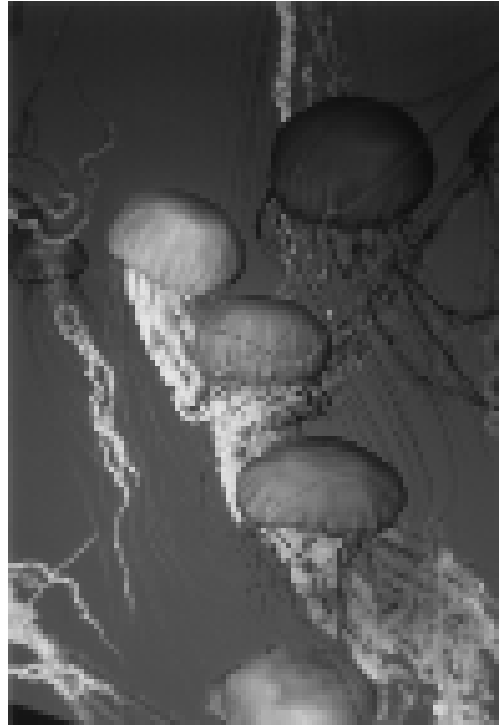
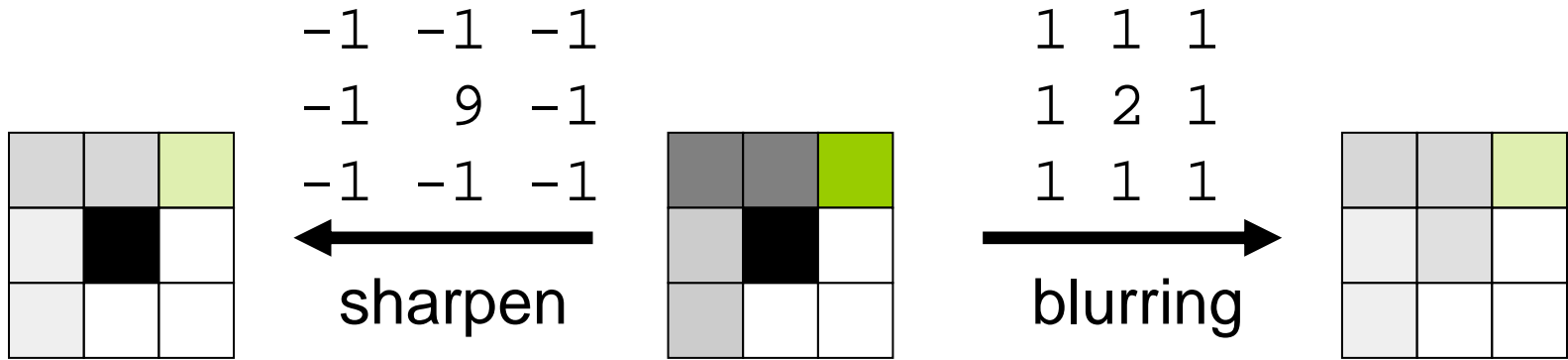
Min, 2pt



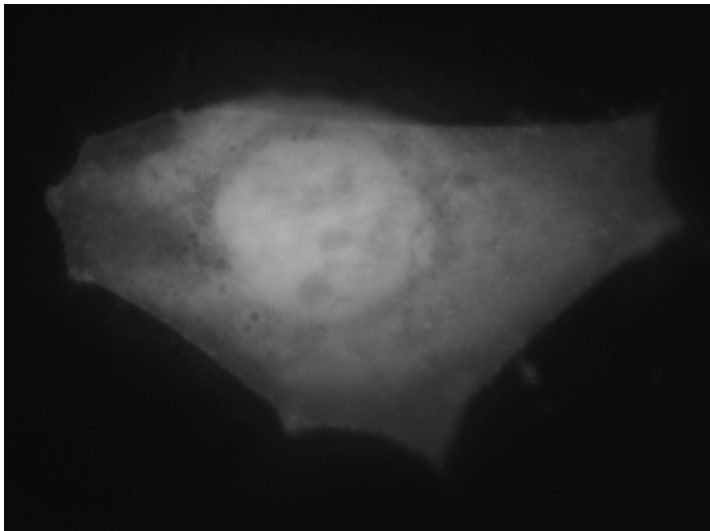
Max, 2pt



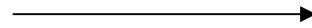
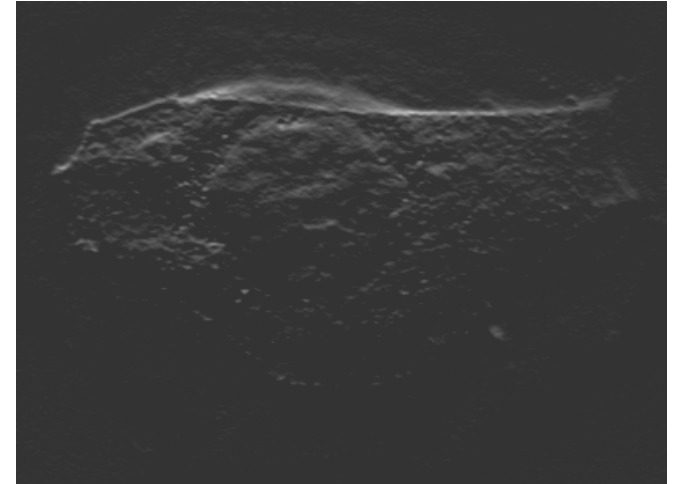
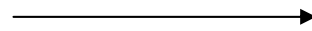
Sharpen / Blur



Example: Edge-Finding with derivatives



$$\begin{array}{ccc} -1 & -1 & -1 \\ 0 & 0 & 0 \\ 1 & 1 & 1 \end{array}$$

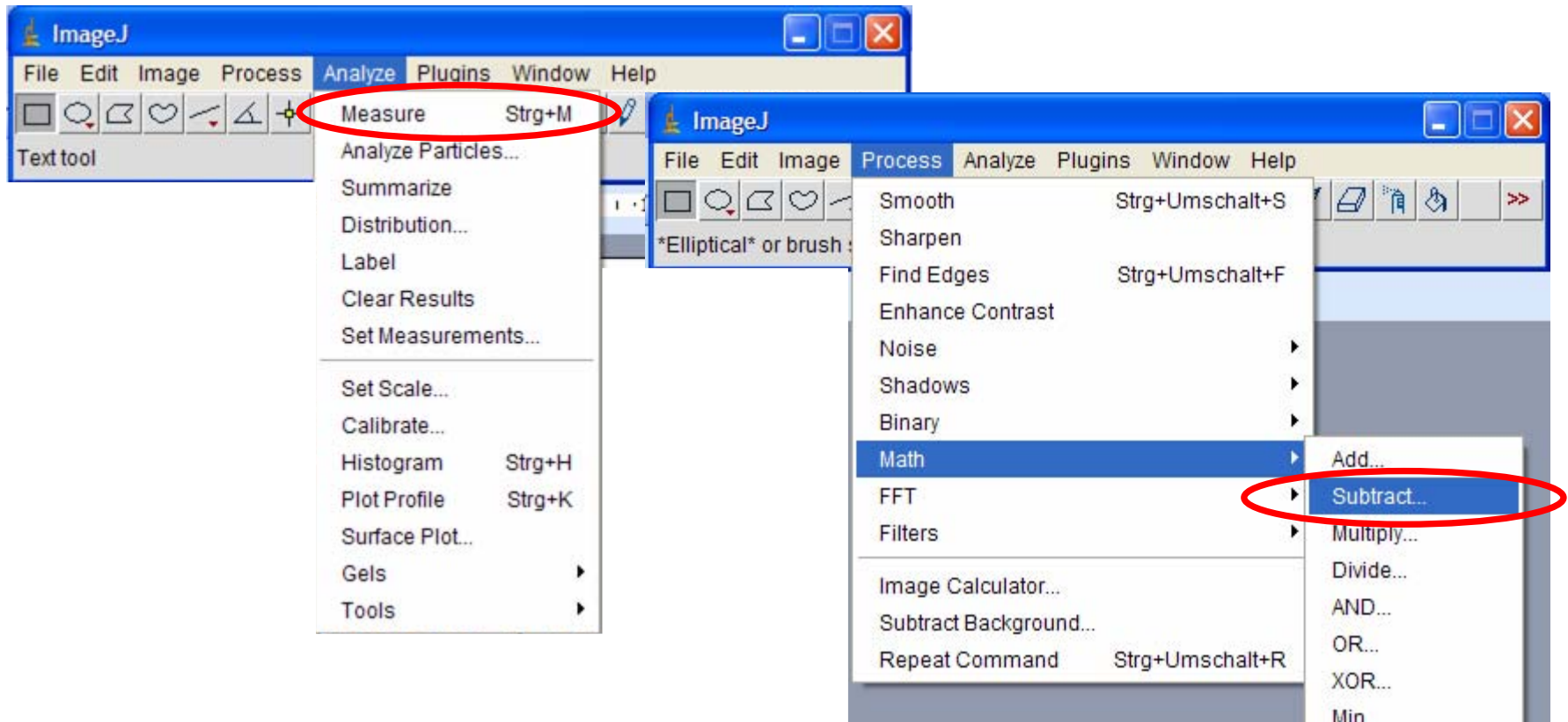


$$\begin{array}{ccc} -1 & -1 & -1 \\ 0 & 1 & 0 \\ 1 & 1 & 1 \end{array}$$



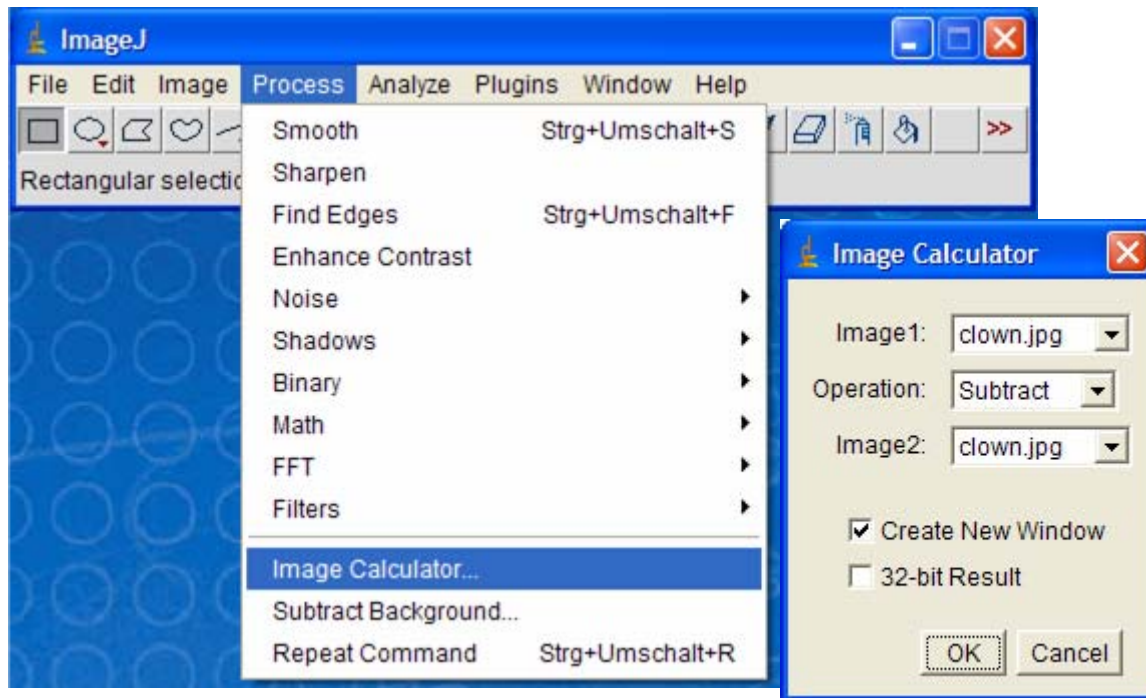
Background Subtraction

- Even background:
 - Measure the average intensity in the background and subtract this average from image



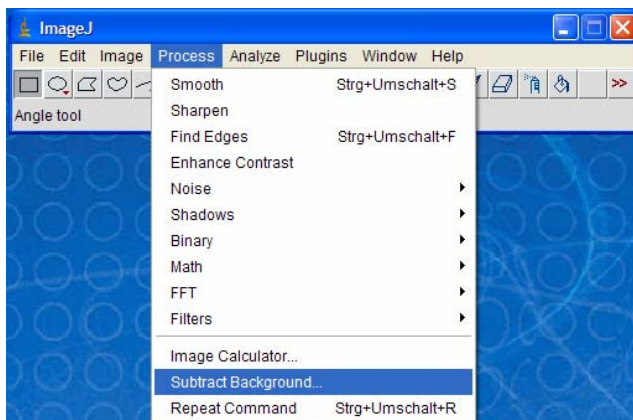
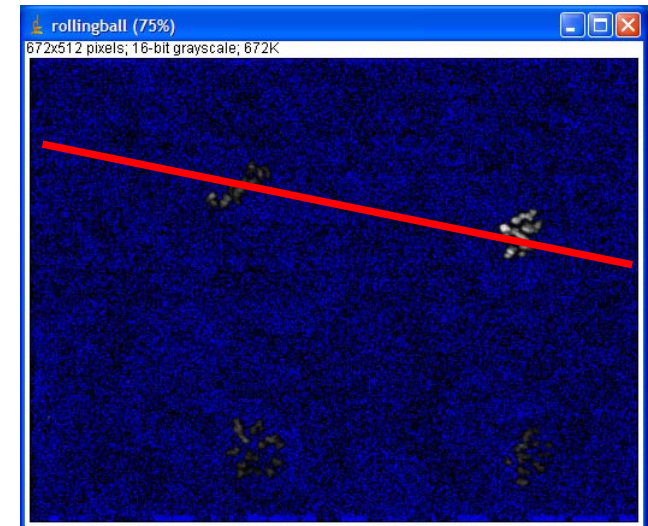
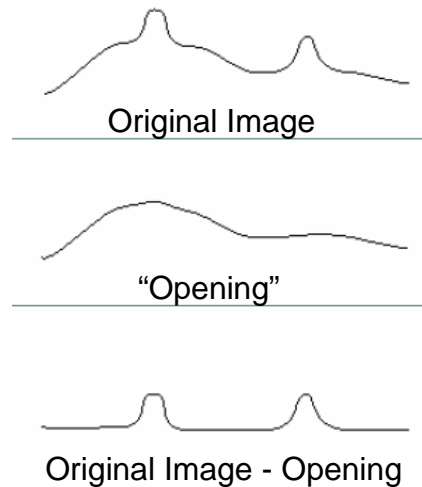
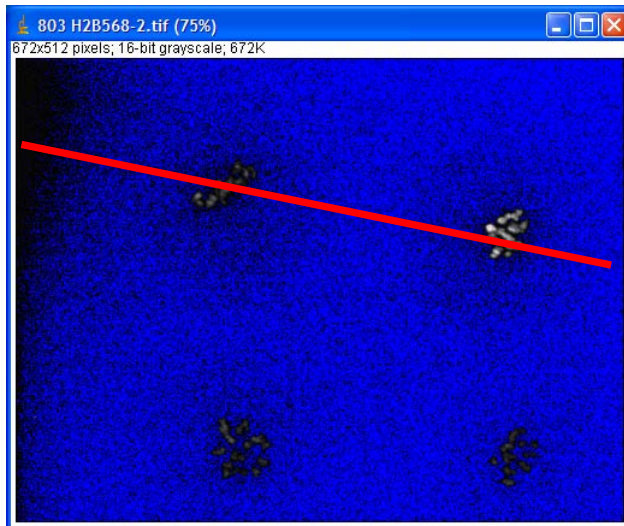
Background Subtraction

- Subtract “background image”
(same exposure time without illumination)

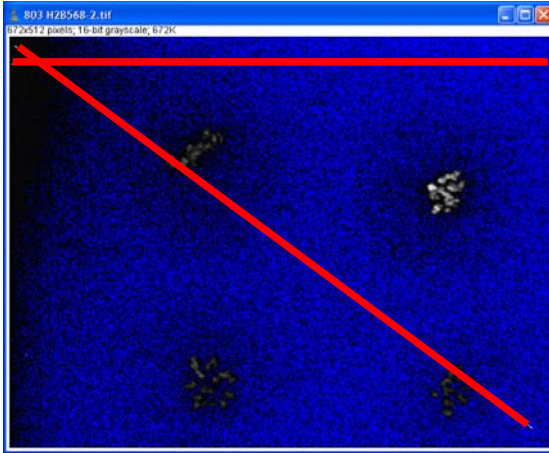


Background Subtraction

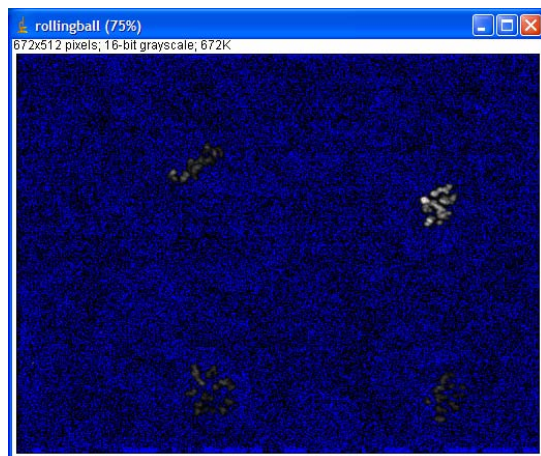
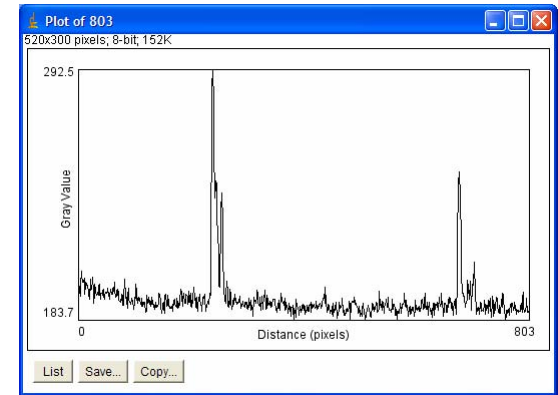
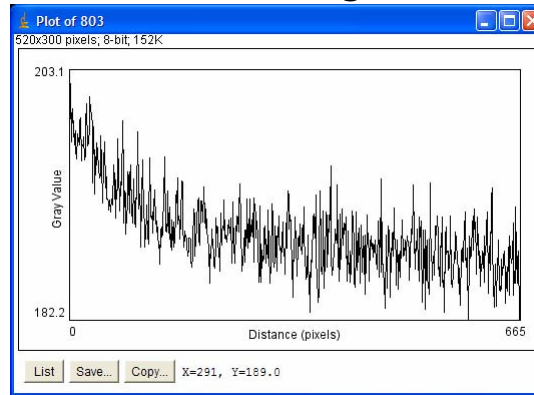
- Uneven background: Rolling ball filter
 - Use kernel larger than diameter of largest object



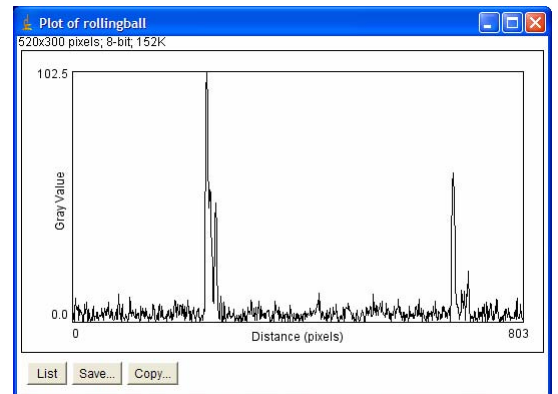
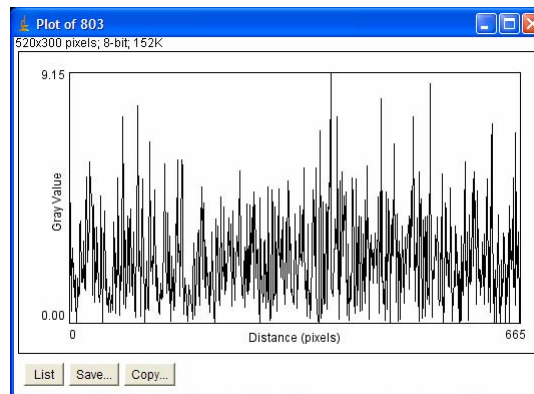
Line Profile



Without background subtraction

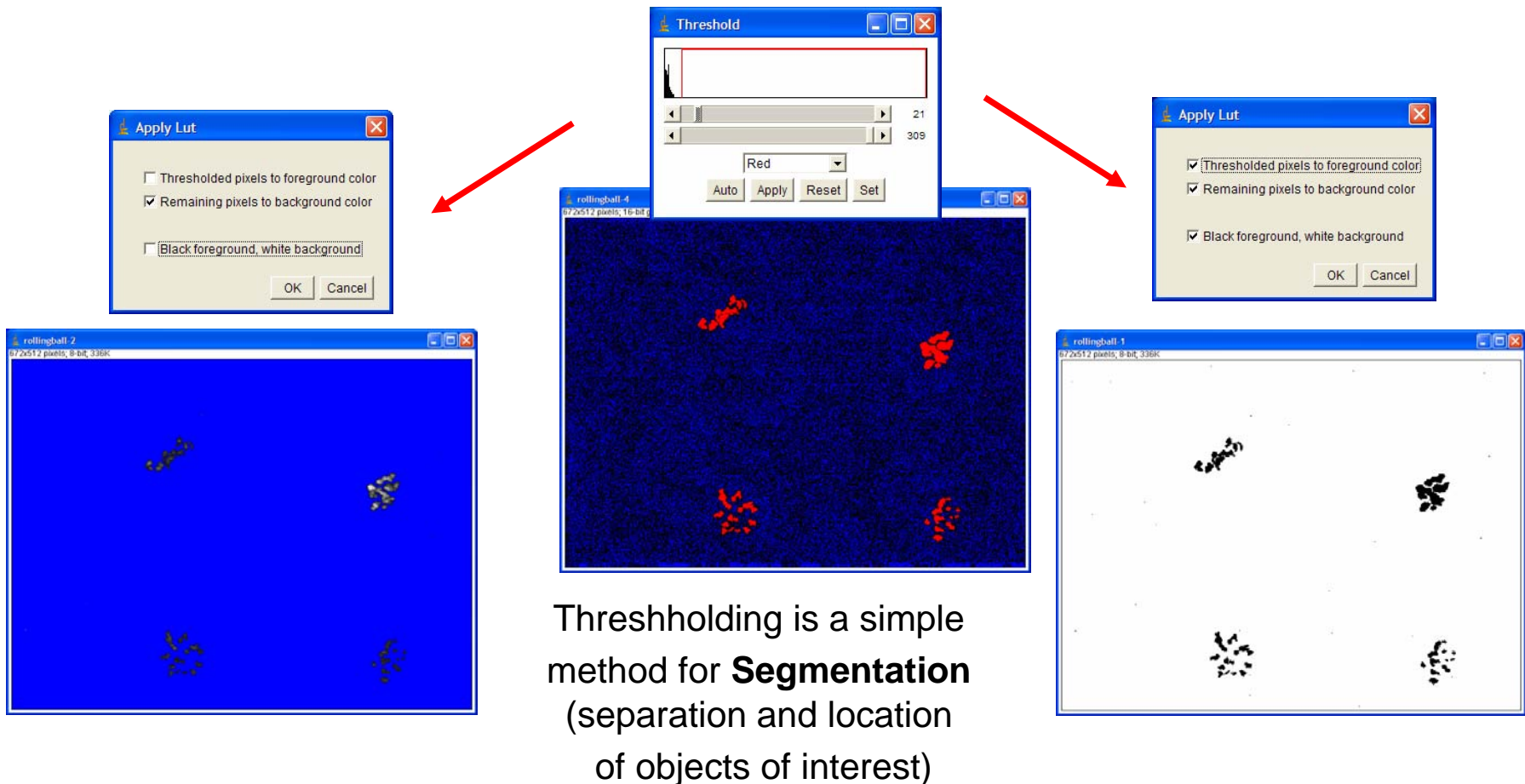


After rolling ball (50) background subtraction



Thresholding

Thresholding is used to change pixel values above or below a certain intensity value (threshold):



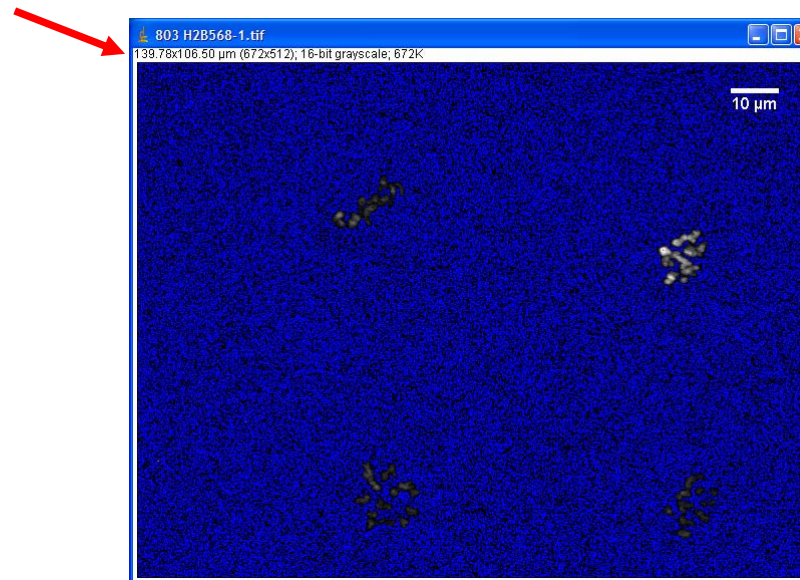
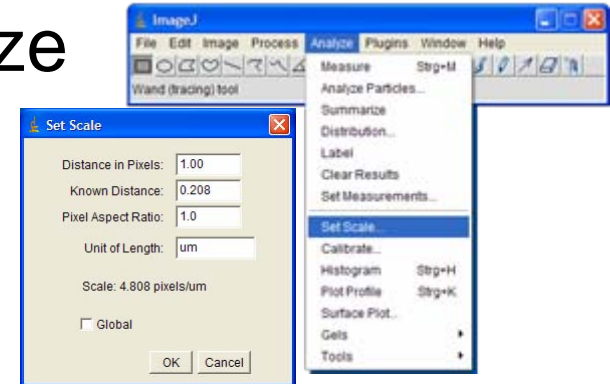
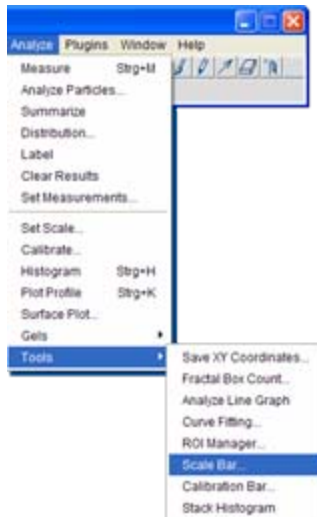
The diagram illustrates the thresholding process through a series of steps:

- Initial Image:** A window titled "rollingball_2" shows a blue background with several green, irregularly shaped objects.
- Thresholding Dialog:** A "Threshold" dialog box is shown, featuring a histogram, a color selection dropdown set to "Red", and buttons for "Auto", "Apply", "Reset", and "Set".
- Apply Lut Dialog:** An "Apply Lut" dialog box is shown with the following options:
 - Thresholded pixels to foreground color
 - Remaining pixels to background color
 - Black foreground, white backgroundButtons for "OK" and "Cancel" are at the bottom.
- Final Segmented Image:** A window titled "rollingball_1" shows the result: the green objects are now black on a white background.

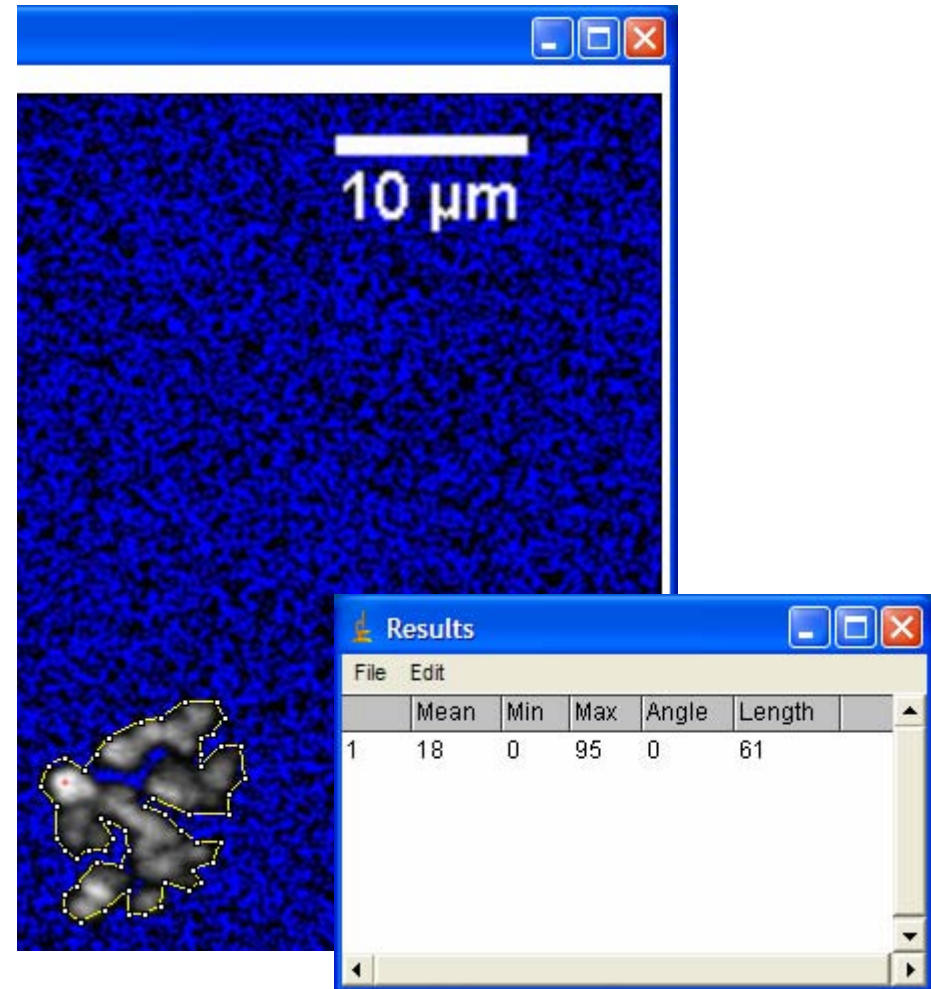
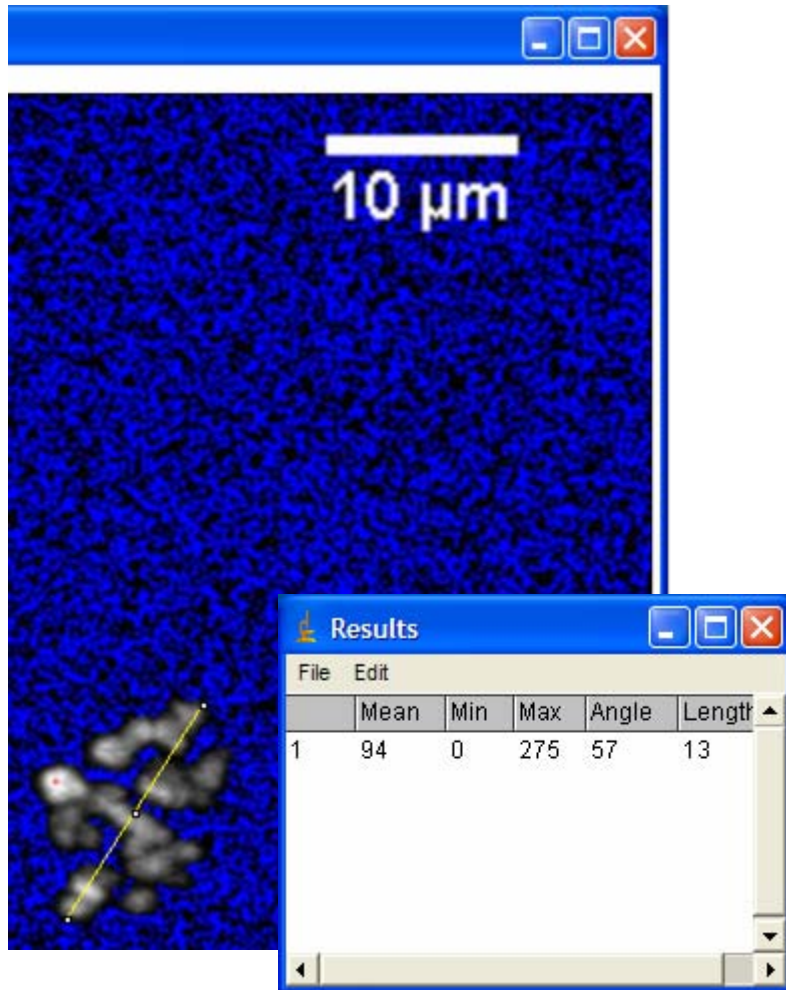
Thresholding is a simple method for **Segmentation** (separation and location of objects of interest)

Measuring Sizes

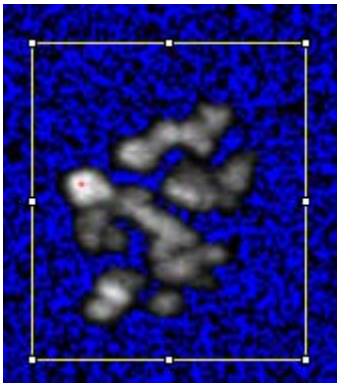
- Set Scale with pixel (voxel) size
- Include Scalebar



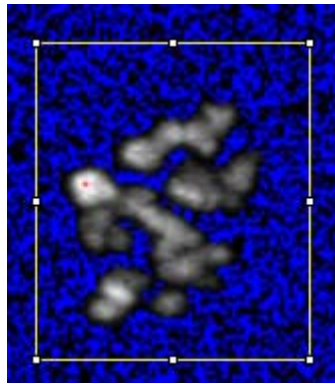
Measuring Length



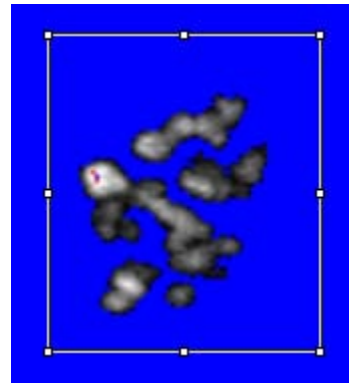
Area Measurement



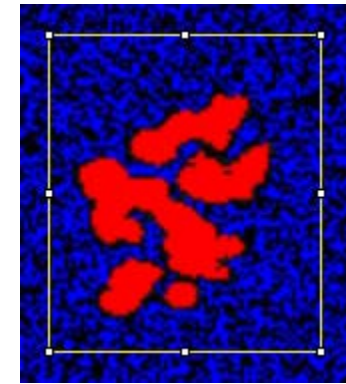
16bit image



32bit image

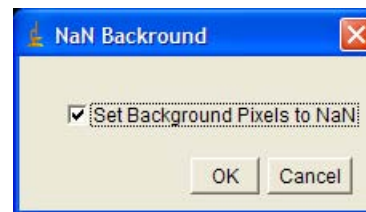


32bit image,
background thresholded
to "Not a Number"



16bit image,
same threshold
as in 32bit image
but not applied

	Area	Mean	Min	Max	Median
1	232	26	0	298	3
2	232	26	0	298	3
3	50	109	35	292	101
4	50	109	35	292	101



Analyze Particles

10 µm

Analyze Particles

Size (µm²): 0.50-Infinity

Circularity: 0.00-1.00

Show: Outlines

Display Results Exclude on Edges

Clear Results Include Holes

Summarize Record Starts

OK Cancel

Measure Strg+M

Analyze Particles...

Summarize

Distribution...

Label

Clear Results

Set Measurements...

Set Scale...

Calibrate...

Histogram Strg+H

Plot Profile Strg+K

Surface Plot...

Gels

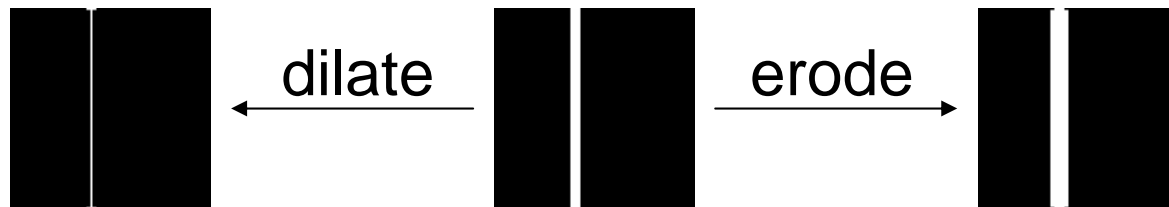
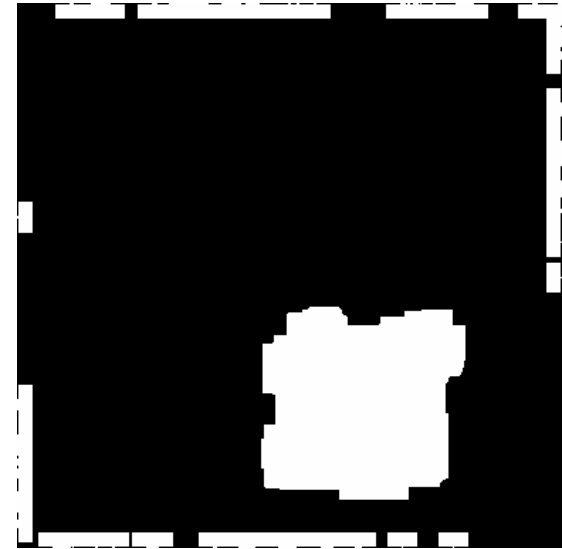
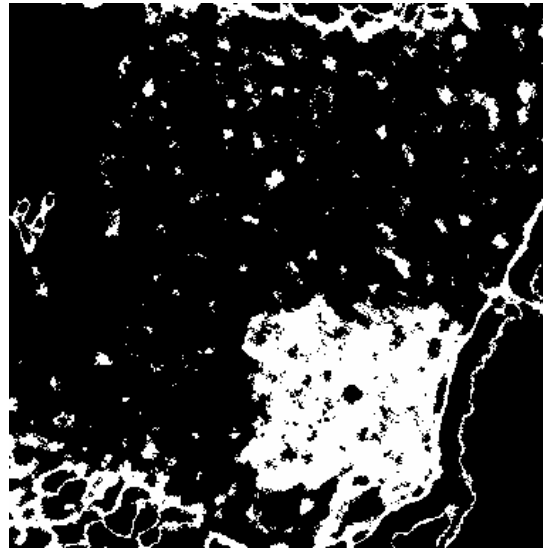
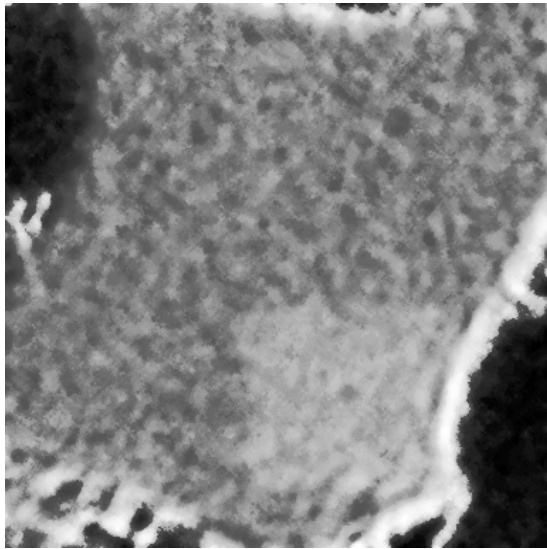
Tools

Segmented objects

Results

	Area	Mean	Min	Max	Median
1	11	121	35	253	122
2	35	110	22	309	100
3	6	120	35	290	107
4	2	84	35	163	84

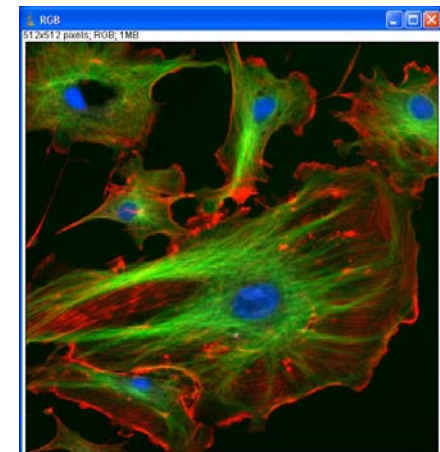
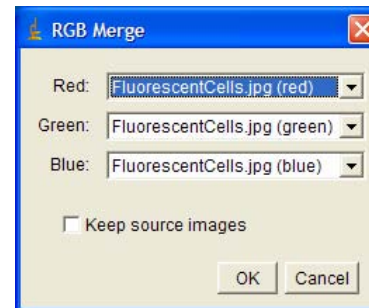
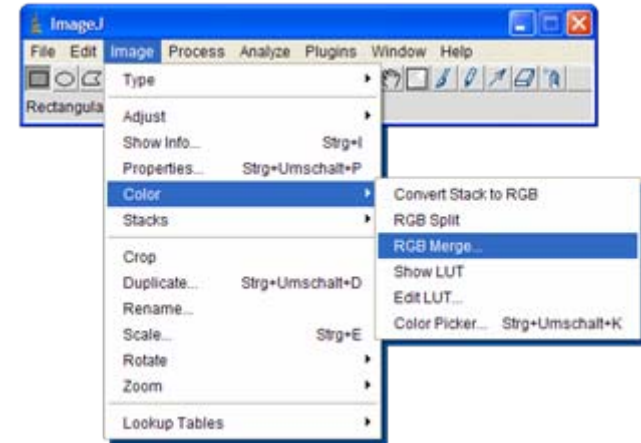
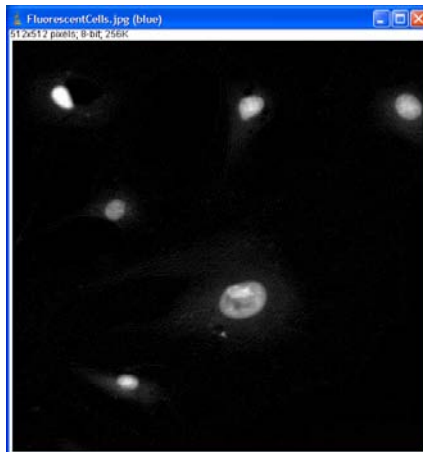
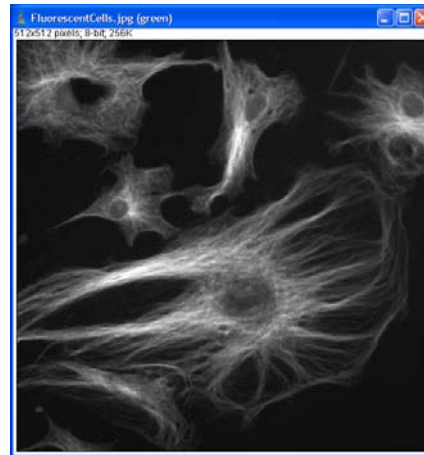
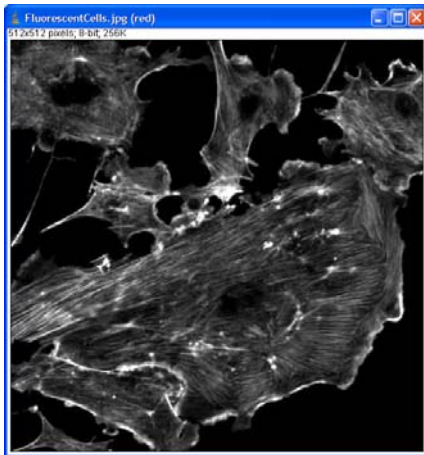
Threshold and Opening/Closing



Closing: Dilate/Erode
Opening: Erode/Dilate

Color Functions

- RGB Merge /RGB Split



Acknowledgements

- ALMF: Rainer, Timo, Arne, Yury
- Andreas Schleifenbaum (some slides)
- Chizuko Tsurumi (images)

Further Information

- Microscopy Primer
<http://micro.magnet.fsu.edu/primer/index.html>
- EAMNET Teaching Modules (Zeiss Basics!)
http://www.embl.de/eamnet/html/teaching_modules.html
- Digital Filters:
<http://homepages.inf.ed.ac.uk/rbf/HIPR2/filtops.htm>