



**Penn Diabetes Research Center
Rodent Metabolic Phenotyping Core**
Director: Joseph Baur, Ph.D.
Technical Director: Jennifer Rojas, Ph.D.

Request For IKA C 3000 Bomb Calorimetry Analysis For Academic Investigators

Lab/PI:	Requestor:
Phone:	Request Date:
Email:	Account Number:

Select	Species
<input type="checkbox"/>	Mouse
<input type="checkbox"/>	Rat

Rodent fecal collection protocol:

1. Single-house rodents in clean housing cages for a pre-determined period of "time (t1)", allowing a minimum of ~0.8 to 1.0 gram of fecal material to accumulate. Standard collection times for mice are 48 to 72 hours and for rats, 24 to 48 hours. Alternatively, you can house the animals in metabolic cages with a wire floor for the measurement days (if available), to prevent coprophagia.
2. Determine body weight and food weight at "time zero (t0)" and at collection day "t1".
3. Collect fecal samples using forceps and weigh to determine "wet weight". Mass of "wet" feces placed into tube (and tare mass of tube) at "t1" (need to know how much is in the collection tube).

Sample Storage Samples should be stored at -80°C.

Information needed from core user (please fill-in information in grid box below).

- Animal ID
- Body weight and food weight at "t0" date
- Number of days when "t1" fecal samples were collected
- Body weight and food weight at "t1" date
- Mass of feces produced ("wet") at "t1"

Fecal and feed samples needed from core user (sent to core on dry ice):

- 0.8 to 1.0 gram fecal samples from "t1" timepoint in tubes.
- 1.0-2.0 gram feed sample (preferably a sample from the lot that was supplied to the animals at "t1" – will be considered "wet" until desiccated).

Select	Individual Assays	Sample amount	No Of Samples	Unit price	Minimum order	Total
<input type="checkbox"/>	Fecal sample	0.8-1.0 g		\$40/replicate	\$200	
<input type="checkbox"/>	Feed sample	1.0-2.0 g		\$40/replicate	\$200	

Total \$



Penn Medicine

Perelman School of Medicine
University of Pennsylvania Health System

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Sample Shipping and Delivery:

UPenn Investigators: Place samples in designated retainer bin in the drop-off freezer #2 in the 12-170 corridor.

External investigators: Please ship samples Monday through Wednesday so the core receives them no later than Thursday of any given week. Ship on dry ice to the following address: Attn: Jennifer Rojas, RM 172 TRC 12TH FL, 3400 CIVIC CENTER BLVD BLDG 421, ENDOCRINOLOGY/IDOM, PHILADELPHIA, PA 19104, USA.

Sample Labeling:

The sample box and each tube should be uniquely labeled (e.g. Date, Sample ID, Investigator's name/initials, and type of assay requested).

Please send order request and all inquiries to the Technical Director, Jennifer Rojas at Jennifer.Rojas@pennmedicine.upenn.edu.

Kindly acknowledge the Penn Diabetes Research Center grant P30-DK19525, and the services of the Rodent Metabolic Phenotyping Core in all publications and presentations.

Please consider the following guidelines when determining whether co-authorship is warranted for core personnel: <https://abrf.org/authorship-guidelines>.