Yeast Small Scale Transformation

1. Prepare carrier DNA by heating 200ul of Herring Testes DNA (20 mg/ml) to 90-100°C for 5-10 minutes. Immediately add 800ul of LiSORB to heated Herring Testes DNA and vortex to mix. Allow carrier to cool to room temperature.

2. Add 5ug plasmid DNA to 50ul Carrier DNA. (The remaining Carrier DNA can be stored at –20°C for later use).

3. Add 50ul of competent yeast to plasmid/carrier mix. Incubate at 30°C for 30 minutes.

4. Add 450ul 40% PEG3400 and mix gently by pipetting. Incubate cells at 30°C for 30 minutes.

5. Heat shock cells for 10 minutes at 42°C.

6. Pulse spin cells for 5-10 seconds.

7. Pipet off PEG.

8. Recover yeast cells by incubating them in 1ml selective DOB at 30°C for 1 hour while shaking.


10. Plate ~100ul of resuspended cells on selective DOBA plates. Incubate plates inverted at 30°C.