

Vol Act: 1 unit /  $\mu$ l ; 1.000 ml  
D Nase (1) + (2) > 50 units  
Exo: > 50 units  
Nick: > 50 u ; R Nase: > 50 units  
S. A.: 3308 units / mg

Boehringer Mannheim GmbH

3/3/92

## Phosphatase, alkaline

from calf intestine

*Orthophosphoric-monoester phosphohydrolase*  
(alkaline optimum), EC 3.1.3.1

### Special quality for molecular biology

1 000 units, Cat. No. 713 023

1 000 units, Cat. No. 1097 075, high concentration

### Product description

#### Volume activities:

Cat. No. 713 023: ca.  $1 \times 10^3$  units/ml.

Cat. No. 1097 075: ca.  $20 \times 10^3$  units/ml.

Alkaline phosphatase is assayed according to (1). One unit of alkaline phosphatase is the enzyme activity which hydrolyzes 1  $\mu$ mol of 4-nitrophenyl phosphate in 1 min at 37 °C under assay conditions. Note: According to (1) 5 units alkaline phosphatase (37 °C; diethanolamine buffer) correspond to 1 unit alkaline phosphatase (25 °C; glycine/NaOH buffer).

See data label for lot-specific values.

**Storage buffer:** Triethanolamine, 30 mmol/l; NaCl, 3 mol/l; MgCl<sub>2</sub>, 1 mmol/l; ZnCl<sub>2</sub>, 0.1 mmol/l; pH ca. 7.6.

**Activity determination:** The activity determination is performed according to (1) at 37 °C in diethanolamine buffer, 1 mol/l; 4-nitrophenyl phosphate, 10 mmol/l; MgCl<sub>2</sub>, 0.5 mmol/l; pH 9.8.

**Specific activity:** ca. 2000 units/mg according to (1) and (2). See data label for lot-specific values.

**Stability:** stable at +4 °C.

#### Supplied buffer:

Dephosphorylation buffer, 10-times concentrated: Tris-HCl, 0.5 mol/l; EDTA, 1 mmol/l; pH 8.5 (20 °C).

**Incubation procedure (3, 4):** The reaction assay is adjusted with 1/10 volume 10  $\times$  dephosphorylation buffer.

- 50 pmol 5' terminal phosphorylated DNA fragments (3'-recessed, 5'-recessed or blunt-ended) are incubated with 1 unit alkaline phosphatase at 37 °C for 60 min or
- 50 pmol 5' terminal phosphorylated RNA fragments are incubated with 1 unit alkaline phosphatase at 50 °C for 60 min.

**Inactivation of alkaline phosphatase (4, 5):** Add 1/10 volume of EGTA, 200 mmol/l, to the reaction assay and heat to 65 °C for 10 min. To achieve complete inactivation of alkaline phosphatase, an extraction with phenol/chloroform/isoamylalcohol (50:48:2) should be performed.

### Quality control

See data label for lot-specific values.

#### Absence of deoxyribonucleases:

1) 1  $\mu$ g  $\lambda$ DNA is incubated with alkaline phosphatase for 1 h at 37 °C in 25  $\mu$ l dephosphorylation buffer. The number of enzyme units which shows no degradation of  $\lambda$ DNA after agarose gel electrophoresis is stated under "DNase(1)".

2) 1  $\mu$ g *Eco RI/Hind III* fragments of  $\lambda$ DNA is incubated with alkaline phosphatase for 1 h at 37 °C in 25  $\mu$ l dephosphorylation buffer. The number of enzyme units which shows no change in the banding pattern after agarose gel electrophoresis is stated under "DNase(2)".

**Absence of nicking activities:** 1  $\mu$ g supercoiled pBR322 DNA is incubated with alkaline phosphatase for 1 h at 37 °C in 25  $\mu$ l dephosphorylation buffer. The number of enzyme units which shows no relaxing of the supercoiled structure of pBR322 DNA after agarose gel electrophoresis is stated under "Nick".

**Absence of exonucleases:** A 40-mer oligonucleotide is incubated with increasing amounts of alkaline phosphatase for 4 h at 37 °C and checked for exonuclease contamination by <sup>32</sup>P-labeling with polynucleotide kinase\* and subsequent separation on a 8% polyacrylamide sequencing gel. The number of enzyme units which shows no degradation of the oligonucleotide is stated under „Exo“.

**Absence of ribonucleases:** 5  $\mu$ g MS2 RNA are incubated with alkaline phosphatase for 1 h at 50 °C in 50  $\mu$ l dephosphorylation buffer. The number of enzyme units which shows no change of MS2 RNA after agarose gel electrophoresis is stated under "RNase".

#### References

- Moessner, E. et al. (1980) Z. Physiol. Chem. **361**, 543.
- Bradford, M. (1976) Anal. Biochem. **72**, 248.
- Maxam, A. M. & Gilbert, W. (1980) Methods Enzymol. **65**, 499.
- Chaconas, G. & v. d. Sande, J. H. (1980) Methods Enzymol. **65**, 75.
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\* available from Boehringer Mannheim GmbH