

Indo-1 Analysis for Calcium Determination

- Ratiometric method for quantitation of internal cellular calcium levels
 - Requires ultraviolet illumination (argon ion laser)
 - Fluorescence recorded as the ratio of 405nm to 485nm
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REAGENT LIST:

- Indo-1 Staining Solution:
 - Indo-1,AM (Molecular Probes I-1223) 50ug
 - Anhydrous DMSO 50ul
 - Calcium ionophore:
 - 4-Bromo-Calcium ionophore A23187 (Sigma B-7272)
1mg/ml in DMSO
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PROTOCOL:

- 1- Spin down 1×10^6 cells per tube (1 tube per condition or variable being tested)
 - 2- Aspirate medium
 - 3- Resuspend in 0.2 ml cell culture medium + Indo-1 (2ug/ml indo-1)
 - 4- Incubate 15-30 minutes at 37 deg C.
 - 5- After incubation, complete volume to 1ml using cell culture media.
 - 6- Keep cells on ice until analysis.
 - 7- Briefly warm individual cell aliquots to 37 deg C a few minutes before analysis.
 - 8- Cells are run on the cytometer for 30 seconds to establish a baseline calcium level, then removed, stimulated, and replaced on the cytometer and followed for typically up to 10 minutes.
 - 9- Calcium ionophore positive control uses 5ul of A-23187 solution per 10^6 cells.
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TIPS:

- 1- Indo-1,AM staining solutions should only be mixed in small aliquots.
 - 2- Use calcium ionophore as a positive control, should give maximum dynamic range. If response is poor, check system and cell loading.
 - 3- Rinse flow system after ionophore control, using 70% ETOH followed by buffer rinse; carryover of ionophore can be a problem.
 - 4- Microscopic evaluation of Indo-1 stained samples using UV setup (i.e. DAPI,Hoechst) should show diffuse staining- excessive staining in cell compartments indicates overstaining (too long/too much)
 - 5- Incomplete loading can cause trouble in a number of cell types-
 - cells with high MDR activity may pump Indo out; try blocking this using verapamil or cyclosporin A.
 - use Pluronic (Molecular Probes) when loading the cells
 - esterase activity may not be sufficient in some cells
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