# **IS-PCR** Accompanying Protocol: DNAse Treatment

- 1. Execute standard deparaffinization, rehydration and PrK treatment (steps 1-9 above).
- 2. Incubate the tissue sections with Dnase in a humid chamber at 37°C O/N (~19hrs). I like to use the Boekel hybridization oven that accommodates the large humid chamber. Use 30-40λ of reaction mix to cover each prostate section. Use less for smaller tissues.

### Reaction Mix:

1X Reaction Buffer (supplied as 10X)

 $0.6U/\lambda$  DNAse ( $1U/\lambda$  stock) In the future buy a more concentrated stock! Sterile diH<sub>2</sub>O to volume.

- 3. Incubate in a large volume of **Rinse Solution** @RT for 10' (40 mM Tris-Cl pH 7.4, 2 mM CaCl<sub>2</sub> and 6mM MgCl<sub>2</sub>).
- 4. Rinse in multiple washes of sterile water (3) and let air dry for PCR.

## **IS-PCR** Recipes

### TBS (Tris-Buffered Saline)

100 mM Tris-Cl pH 7.4 150 mM NaCl Autoclave sterilize. Store at RT.

#### Proteinase K Stock

Reconstitute lyophilized proteinase K to final concentration of 20 mg/mL, in a sterile

10 mM Tris pH 7.5, 20 mM CaCl, 50% glycerol solution, then make 500λ aliquots. This will enhance enzyme stability and allow repeated withdrawls from aliquots without diminishing the enzyme's activity. DO NOT RECONSTITUTE IN WATER!

Store at -20 °C.

#### <u>BSA V</u>

There are two stocks, one for PCR and another for the blocking step.

Use a nuclease-free stock for PCR (ex. Gibco BRL)

For the blocking step, reconstitute BSA powder in TBS to a stock concentration of 200 mg/mL. Sterile filter and make 10 mL aliquots. Store both at  $-20 \text{ }^{\circ}\text{C}$ .

#### CDB (Conjugate Dilution Buffer)

Make fresh--per mL add together:

100λ 1M Tris pH 7.3 150λ 1M MgCl<sub>2</sub>

50λ 200 mg/mL BSA V.

700λ diH<sub>2</sub>O

### APSB (Alkaline Phosphatase Substrate Buffer)

100 mM Tris-Cl pH 9.5 A little trick: autoclave MgCl<sub>2</sub> separately from the Tris and salt to avoid precipitation.

150 mM NaCl Store at 4°C to preserve alkaline pH.

50 mM MgCl<sub>2</sub>