Purification of DNA Oligos from Gel

1) Resuspend the oligos in water or TE buffer.
2) Pour Native-PAGE at appropriate concentration for size of oligos.
   ie. 20% Native PAGE for 15mer-20mer

   Make PAGE gel with 0.5X TBE buffer > ddH2O, acrylamide, TBE buffer.
   Does not need stacking gel.
3) Load samples with loading buffer without dye. Add one lane with dye at far right or left lane.
4) Run Gel in 0.5X TBE buffer at 150-400V COLD. Gel heats up. Make sure buffer is enough to keep gel cold. Use cooling chamber or cold room.
5) Run to approximately ¾ of the way to the bottom of the gel. Use mini gel.
6) Remove gel. Place on saran wrap. Place on TLC Fluorescence Plate. UV shadow with short wavelength light.
7) Mark dark DNA shadows with a pencil.
8) Cut out the band (in as small a piece as possible). Cut the gel slices into many small pieces. Place in 1.5ml eppendorf tube.
9) Add 500ul 0.3M NaAcetate (pH 4.8-5.2). Shake O/N at 37C in a vortexer.
10) Spin out gel slice (Save and redo if necessary). Phenol/CHCl3 extract. Precipitate with ethanol and acetate O/N at 80 C. Wash 2X with 70% ethanol to remove salt.
11) Resuspend in sterile H2O or TE buffer. (Use approx. 105ul)
12) Spec for concentration (use 5ul/500ul dilution 1:100)