^{32}P labeling of large DNA Fragments.

1. Purify DNA fragment from gel and Phenol/$CHCl_3$ extract a couple of times to remove residual agarose

2. Aliquot 200-500ng of DNA into screw cap tube and Add ddH_2O to a final volume of 40µl

DNA	- 10µl (will vary)	Δ 100°C 5-10 mins
ddH_2O	- 30µl	- quick spin and place on ice immediately.
Primers (hexa/nanomers)	- 20µl	
5X dCTP Buffer	- 20µl	add to tube on ice
3000 ci/mmol ^{32}P-dCTP	- 20µl	* use Stratagene Primer II Kit
Klenow	- 2µl	
	102µl	

- incubate tube at 37°C for 15-30 mins
- Add stop 2µl and mix gently
- Centrifuge to get labeled DNA to the bottom (will condense in cap)
- Nuctrap to clean up
- Count probe 1µl spotted on filter etc.
- Boil 5-10 mins before use.