# Alkaline Phosphatase usage

- 1. Alkaline Phosphatase
- 2. Shrimp Alkaline Phosphatase

#### Animal and Cell Culture Procedures

- 1. Cell freezing protocol
- 2. Small Molecule animal protocol
- D Freezing Cells
- 1) Ampoules and Vials Protocol
- 2) Freezing cells
- 3) Freezing media

#### BACmid Recombineering Protocols

- 1. A Highly Efficient Recombineering-Based Method for Generating Conditional Knockout Mutations
- 2. BAC DNA Mini-Preparation
- 3. Bac recombineering vectors
- 4. BAC Recombineering Protocol
- 5. BAC Recombineering protocol 1
- 6. Recombineering protocol 2
- 7. Recombineering protocol 3
- 8. Recombineering protocol 4
- 9. BAC d E3c
- 10. DNA Max-Prep Protocol
- 11. High-efficiency counterselection recombineering for site-directed mutagenesis in bacterial artificial
- 12. J. Virol.-2002-Zhou-6185-96
- 13. NEBlot Kit Instruction Manual Preparation of High Specific Activity DNA Probes
- 14. pEL04 map
- 15. pGalK map
- 16. pTamp map
- 17. pIGCN21 map
- 18. PL253 map
- 19. PL451 map
- 20. PL452 map
- 21. Preparing solutions for BAC DNA preparation
- 22. QIAGEN Large-Construct Kit Handbook
- 23. Recombineering a homologous recombination-based
- 24. Simple and highly efficient BAC Recombineering using
- 25. Strains

#### Bacteria Growth Transformation and Maintenance and Bacteriophase

- 1. 2 x YT Medium, M9 Minimal Medium
- 2. Antibiotic Solutions
- 3. Diluent
- 4. LB Medium, NZCYM Medium, NZYM Medium, NZM Medium
- 5. Media with Agar or Agarose
- 6. Solutions for Working with Bacteriophage
- 7. Storage Media, Stab Cultures, Bacterial Cultures Growing in Liquid Media and Agar Plates Terrific Broth, SOB Medium, SOC Medium
- 8. Terrific Broth, SOB Medium, SOC Medium

- T4 Polynucleotide Kinase Assays
- 1. Bacteriophage T4 Polynucleotide Kinase Exchange Reaction
- 2. Bacteriophage T4 Polynucleotide Kinase

#### Beta-Galactosidase and Chloramphenicol Acetyltransferase Assays

- 1. B-Gal Assay
- 2. Harvest and Assay for Chloramphenicol Acetyltransferase
- 🗁 Beta-Galactosidase Staining
- 1. Beta-Galactoside Staining of Cells
- 2. Histochemical Staining
- 3. Immunodetection of Specific Proteins
- C Chemiluminescent B-gal Detection
- 1. Galacto-Light

#### BOSC23 Transfection and Infection of NIH 3T3 Cells

1. Transfection of BOSC23 cells and infection of NIH 3T3 Cells

- D Tet Vector Transfection of BOSC23 cells and NIH3T3 cell Infection
- 1. Pear Transfection of BOSC23 and NIH3T3 Infection
- 2. Tet Vector Transfection of BOSC23 Cells and NIH3T3 Cell Infection

#### Brdu Staining FACS Protocols

- 1. Ca++ Analysis via FACS
- 2. BrdU Staining
- 3. Intracellular staining of Bcl-2 for FACS
- 4. Preparation of Cultured Cells for FACS Analysis
- 5. Propidium Iodide Staining
- 6. Simultaneous Surface and DNA Staining for FACS

#### Calcium Phosphate with BBS Transfection

- 1. Calcium Chloride-BBS Transfection Protocol
- 2. Calcium Phosphate Tranfection System

# Cell Culture Protocols

- 1. Cell Lysis for PCR Analysis
- 2. Collecting and Transfecting Cells
- 3. Epithelial Cell Lines
- 4. FICOLL
- 5. GPT selection
- 6. HVS Transformation of PBMCs
- 7. Lymphocyte Isloation by Ficoll Gradient Centrifugation
- 8. Maintaining 293GP Clones in Culture
- 9. Mammalian Cell Selection by Panning
- 10. Mycophenolic Acid Selection
- 11. Preparation of Cell Cultures and Vaccinia Virus Stocks
- 12. Preparation of Insect Cell Cultures and Baculoviral Stocks
- 13. Primary Lymphocyte Electroporation
- 14. Protein Expression in Insect Cells Using Baculoviral Vectors Overview
- 15. Spin infections
- 16. Transformation Efficiency Assay

- Competent Cells
- 1. CaCl2 Preparation of Competent E.coli
- 2. Preparing ultra-competent E.coli
- 🗁 E. Coli Transformation
- 1. Introduction of Plasmid DNA into cells

#### Chromatin Immunoprecipitation Assay

- 1. ChIP Protocol
- 2. Chromatin Immunoprecipitation from paraffin
- 3. Abcam\_CHIP Assay
- 4. ChIP Assay

#### Cloning and Expression Vectors

- 1. Episomal Mammalian Expression Vectors
- Cloning in Alkaline Phosphatase-Treated Vector
- 1. Cloning in Phosphatase-treated Cosmid Vectors

#### Colony Hybridization

1. Screening for clones by colony hybridization

#### CsCl Purification of Plasmid DNA

- 1. Large scale preparation of plasmid DNA
- 2. Reducing spin times for CsCl isolation for plasmid DNA

#### DAPI Staining of Nucleus

- 1. DAPI
- 2. DAPI European language

#### 🗁 Dialysis

1. Preparing dialysis tubing

# DNA Protocols

- DNA Fragment Purification from Agarose Gel
- 1. cDNA Amplification Kit
- 2. CHART RNA
- 3. DNA Ladder Protocol
- 4. DNA Max-Prep. Protocol
- 5. DNA Mini-Prep. Protocol V1
- 6. DNA Maxi-prep. V1 9.06
- 7. Garbella Cell gel Assay
- 8. Gel Shift Assay
- 9. Hirt supernatent DNA preparation
- 10. Mini-prep. for plasmid extraction
- 11. Mini prep protocol V2
- 12. Nuclei Paraffin DNA
- 13. Oligo DNA
- 14. Optimization of DNA-RNA Extraction from FFPE paper
- 15. Phenol-based methods for isolating DNA from Agarose Gels
- 16. Prep Crude DNA for PCR- Preparation of Phage Lamda DNA
- 17. Protocols for the in situ PCR-amplification and detection of mRNA and DNA sequences
- 18. Rapid Separation of Protein-Bound DNA
- 19. S35 DNA Sequencing
- 20. Salmon Sperm DNA
- 21. Simultaneous Purification of Genomic DNA and Total RNA (supplementary)
- 22. Trizol Reagent
- DNAse I Footprinting
- 1) DNase I protocol for footprinting ToxR membranes
- 2) Optimization and troubleshooting DNase I footprinting reactions
- 3) P32 Labeling of Large DNA Fragments
- 🗁 Phage Lambda DNA Preparation
- 1) Large Scale Preparation of Bacteriophage Lambda
- 2) Preparation of Phage Lambda DNA
- Durification by CNBr Coupling Protocol
- 1) Analysis of DNA-Protein Interactions Using Proteins Synthesized In Vitro from Cloned Genes
- 2) CNBr-Activated Sepharose 4 Fast Flow
- 3) CNBr-Activated Sepharose 4B
- 4) Detection, Purification, and Characterization of cDNA Clones Encoding DNA-Binding Proteins
- 5) DNA-Protein Interactions
- 6) DNase I Footprint Analysis of DNA-Protein Binding
- 7) Methylation Interference Assay for Analysis of DNA-Protein Interactions
- 8) Motility Shift DNA-binding Assay Using Gel Electrophoresis
- 9) Preparation of Nuclear and Cytoplasmic Extracts from Mammalian Cells
- 10) Purification by CNBr Coupling
- 11) Purification of DNA-Binding Proteins Using Biotin.Streptavidin Affinity Systems
- 12) Purification of Sequence-Specific DNA-Binding Proteins by Affinity Chromatography
- 13) Rapid Separation of Protein-bound DNA From Free DNA Using Nitrocellulose Filters
- 14) Types of Chromatography
- 15) UV-Crosslinking of Proteins to Nucleic Acids

### High Efficiency Plasmid Recovery

- 1. Efficient recovery of plasmid DNA with high nuclease activity
- 2. High-yield method for DNA isolation

#### ELISA Assays

- 1. DNA Quality for Transfection
- 2. Electrocomp. E. Coli Cells
- 3. Elisa w HRP, ABTS & acid stop
- 4. ELISA with HRP and TMB
- 5. Superfect Transfection
- 6. Transfection Reagents

#### EMSA Electro-mobility Shift Assays

- 1. Alternative to Polyacrylamide Gels Improves the Electrophoretic Mobility Shift Assay
- 2. Emsa protocol
- 3. Gel Shift Assay
- 4. PCR Fragment Separation Medium

#### Flow Cytometry

- 1. Apoptosis detection with permeable cells
- 2. Detection of apoptosis in unfixed samples with Hoechst 33342
- 3. DFCI Flow Cytometry Facility General Info
- 4. DNA staining with Chromomycin A3
- 5. Hoechst 33342 Staining of Viable Cells
- 6. Indo-1 Analysis for calcium determination
- 7. Propidium Iodide Staining for Cell Cycle Determination
- 8. Propidium Iodide Staining for Dead Cell Exclusion

#### **Fluorescence in Situ Hybridization**

- 1. BAC fluorescence ISH single color
- 2. FISH slides
- 3. FISH with TSA
- 4. Fluorescence In Situ Hybridization
- 5. In situ detection of RNA and DNA
- 6. In Situ DNA Hybridization Products
- 7. Introduction of Micromax Human cDNA Microarray System I
- 8. One color FISH Nick translation
- 9. Standard Protocol
- 10. Summary of TSA-Direct ISH
- 11. TSA and In Situ Hybridization
- 12. TSA Diagrams
- 13. TSA-Direct Kit Components
- 14. TSA-Direct Kits

# Genomic DNA Isolation System

- 1. Commentary
- 2. Construct Nested Deletions with BAL 31 Exonuclease and Exonuclease III
- 3. Construct Nested Deletions with BAL 31 Exonuclease
- 4. Contruction of Nested Deletions for DNA Sequencing
- 5. Genomic DNA Isolation System
- 6. Reagents and Solutions, Commentary

#### Immuno Assays

- 1. Western protocol
- 2. Double staining Immunfluorescence GBP-1 CD31
- 3. Double staining Ki-67-CD31
- 4. Immunofish
- 5. Preparation of Human Sera for EBV Latent Antigen Detection
- D Immunoaffinity Purification of Antibodies by GST Fusion Proteins
- 1) Immunoaffinity Purification of Antibodies against GST Fusion Proteins
- 2) Immunoaffinity purification
- Dimmunofluorescence
- 1) Immunofluorescence Protocol
- 2) Immunofluorescence staining
- 3) Immunofluorescent staining and flow cytometry procedures
- C Immunoperoxidase Staining
- 1) Immunoperoxidase staining procedure
- 2) Immunoperoxide staining cytospins
- Dimmunoprecipitation
- 1) Immunoblotting procedure
- 2) Immunoprecipitation protocol
- 3) Immunoprecipitation with Staphylococcus aureus cells
- 4) Immunoprecipitation
- 5) ImmunoPure and UltraLink Proteins L. A. G. AG
- 6) Monoclonal Antibodies to DNA Tumor Viruses
- 7) Preparation of Protein A Beads
- Difference Content in the second seco
- 1) Chemiluminescent Detection Methods
- 2) Immunodetection by ECL Amersham
- 3) Repeated Probing of Western Blots
- 4) RNA-RNA Hybridization
- 5) Western Stains Using Home-made ECL Chemiluminescent Reagents

# In Situ Protocols

- 🗁 In Situ Amplification and Hybridization
- 1. In Situ Amplication and Hybridization
- In Situ Hybridization of Bacteriophage Plaques
- 1. Hybridization to Nitrocellulose Filters with Bacteriophage Plaques Replicas
- 2. Immobilization of Bacteriophage Plaques
- 3. In Situ Hybridization of Bacteriophage Plaques

🗁 In Situ PCR

- 1. In Situ PCR Imperiale Protocol
- 2. IS-PCR DNAse Treatment

#### Instruction Protocols for sense basic Molecular Biology

- 1. A Guide to the Preparation and Use of Buffers
- 2. cDNA Amplification Kit
- 3. Core Footprinting System
- 4. Glogos Autorad Markers
- 5. Instructions for Using the Sonic Dismembrator
- 6. LacSwitch II Inducible Mammalian Expression System
- 7. Living Colors Fluorescent Protein Protocols
- 8. Matchmaker Two-Hybrid System
- 9. pGEM-T and pGEM-T Easy Vector Systems
- 10. Primer Extension System AMV Reverse Transcriptase
- 11. Push Column Beta Shield Device and Nuctrap Probe
- 12. Stratagene Random Primer Labeling Kit
- 13. Taq Extender PCR Additive
- 14. Yeast Protocols Handbook

#### In vitro Assays

- 1. In vitro Processivity Assay
- 2. Invitro Translation
- 3. invitro\_binding assay protocol
- 4. invitro\_kinase assay protocol
- 5. Ubiquitylation assay
- 6. HAT Assay
- 7. IP HAT Assay
- 8. Transcription Initiation by RNA Polymerase II
- Histone Deacetylase Assay
- 1) HDAC Protocol
- 2) Histone deacetylase assay kit
- 3) Histone H4 Peptide Radiolabeling
- 4) Anti-Histone H1 protocols
- TNT Protocols
- 1) TnT Coupled Reticulocyte Lysate Systems
- 2) TnT Quick Coupled Transcription Translation Systems

### C Kinase Assays

- 1. Histone Kinase Assay
- 2. In vitro Kinase Assay
- 3. Mercury In vitro Kinase Assay Kits
- 4. Protein Kinase Assay with Immunocomplex
- 5. SAPK Jun kinase assays

### 🗁 General Lab Guidelines

- 1. Chemical Hygiene Work Plan
- 2. CTSRMC Approval\_Robertson\_36814\_Apprv\_14NOV14
- 3. Exposure Control Plan
- 4. Lab Guidelines 100520
- 5. LMAX384 revised protocol 06.01.2015
- 6. Metric
- 7. RNAi-Ready pSiren-RetroQ-TetP Vector
- 8. Robertson Notebook Guidelines
- 9. Use of Tissue Culture Room

#### Lab Reagent Stock

- 1. A G sepharose Beads
- 2. Acrylamide Preperation
- 3. Dialysis tubing & CsCl
- 4. dNTP preparation
- 5. Electrophoresis and Transfer Equipement
- 6. Fluorescence mounting medium
- 7. Hybridoma growing instructions
- 8. Media Preperation
- 9. MINI PROTEAN BIORAD
- 10. Odysssey scanner
- 11. PCR + RT PCR Inventory
- 12. Protease Inhibitors
- 13. Puromycin, Hygromycin, G418
- 14. TPA, NaBu
- 15. TPA

# **Liposome Transfection**

- 1. Irvine Scientific Study and Survey
- 2. Lipofectamine
- 3. Optimizing Liposome Transfection Guidelines

#### 🗁 Luciferase

- 1. Luciferase assays-original protocol
- 2. Luciferase Assay Luminometer Protocol
- 3. Luciferase Assay System
- 4. Luciferase Reporter Vectors
- 5. Luciferin Protocol.pdf
- 6. tb281

# Matrigel Assay

- 1. Matrigel Invasion Chamber Guidelines
- 2. Matrigel Migration Assay
- 3. Matrigel Scratch Assay
- 4. Metastasis Experimental Strategies

### Membrane Hybridization Protocols

- 1. Analysis of Genomic DNA by Southern Hybridization
- 2. Buffers and Solutions
- 3. Gene Screen Plus Hybridization Transfer Membrane Manual Table of Contents
- 4. Hybridization to Nitrocellulose Filters Containing Bacteriophage Plaques or Bacterial Colonies
- 5. Isolation of DNA from Mammalian Cells
- 6. Random Priming BMB Kit
- 7. Sieving Agarose Gel Electrophoresis
- 8. Southern Blotting and Hybridization
- 9. Southern Transfer Membrane Hybridization

# Microarray Pathochip Protocols

- 1. 2.5% Gel Electrophoresis
- 2. Amplification
- 3. Amplification-Step C
- 4. DNA purification
- 5. DNA-RNA Extraction and Purification
- 6. Hybridization
- 7. Library Synthesis Reaction
- 8. Procedures for Pathochip Microarray Experiment for Tumor FFPE samples
- 9. Purification of Amplified DNA-QIAquick PCR Purification Kit
- 10. RNA purification
- 11. Sample Labeling. Step 1 Fluorescent Labeling of gDNA
- 12. Sample Labeling Procedure. Step 2 Clean-up Labeled samples
- 13. Vacuum Centrifuge Operation
- 14. Washing Microarray Scanning Steps
- 🗁 Data Analysis Using Partek Genomic Suite
- 1) Experimental design
- 2) Data Interpretation and Choosing Candidates Based on the Analyses
- 3) 4 types of Analysis done using Partek Genomic Suite
- 4) 1st step Normalization of the data

# Molecular Biology Protocols

- 1. Doublestaining Ki-67 CD31 mit EnVision
- 2. Factors that influence
- 3. FICOLL
- 4. Gelatin Zymography Protocol
- 5. Gst pulldown for proteomic study
- 6. Harvesting Cells for Luciferase Assay
- 7. High-risk behaviors among adult men and women in Botswana: Implications for HIV-AIDS prevention efforts
- 8. Knowledge HIV UB students
- 9. Lysis Buffer Preparation of LMP2 Membrane Extracts
- 10. Montaner CITI
- 11. Packaging Extract Protocol II
- 12. Passage of C-terminal Xbal Linker Mutant Virus from E1 C64 no. 22 LCL
- 13. Phase Lock Gel Manual
- 14. Preparation of Vectors Treated with Alkaline Phosphatase
- 15. Pulse chase Assay
- 16. Refractive Index Reference for CsCl
- 17. Ribonuclease Protection Assay
- 18. Shift Western Protocol
- 19. Staph Preparation
- 20. Stratagene Gigapack Gold Packaging Extract
- 21. Sucrose Gradient Centrifugation
- 22. Western Blot of Protein Expressed by Yeast
- P32 Labeling of GST Fusion Proteins
- 1) Generation of P32-GST Fusing Proteins
- 2) P32-GST Scale-up guidelines
- Difference of the second secon
- 1) Agar, Agarose and Concentrated Media
- 2) Antibiotics
- 3) Autoradiography
- 4) Buffers for Restriction Endonuclease Digestion
- 5) Cell Counts with a Hemacytometer
- 6) Chromatography Through Sephadex G-50
- 7) Common Electrophoretic Buffers
- 8) Commonly Used Gel-loading Buffers
- 9) Concentration of Nucleic Acids
- 10) Determination of Viability by Trypan Blue Exclusion
- 11) Drying Down 32P-labeled Nucleotides from Mixtures of Ethanol and Water
- 12) Enzymes
- 13) Glassware and Plasticware
- 14) Liquid Media
- 15) Mutation Detection by Cleavase Fragment Length Analysis
- 16) Measurement of Radioactivity in Nucleic Acids
- 17) Northern Blotting
- 18) Preparation of Buffers and Solutions
- 19) Preparation of Dialysis Tubing
- 20) Preparation of Multimers of Plasmids as Molecular Weight Markers
- 21) Preparation of Organic Reagents
- 22) Preparation of Stock Solutions
- 23) Proteolytic Enzymes

# 😽 Penn Medicine

- 24) Protocol for Sequencing by the Maxam-Gilbert Technique
- 25) Purification of Nucleic Acids
- 26) Quantification of DNA and RNA
- 27) Solutions for Working with Bacteriophage Lambda
- 28) Solutions BIOCHEMICAL TECHNIQUES
- Preparation of Competent Cells
- 1) Competent bacteria protocol
- 2) Competent Yeast Cells
- 3) Preparation of Competent Cells

#### Nuclear Protocols

- 1. Preparation of Nuclear and Cytoplasmic Extracts from Mammalian Cells
- 2. Nuclear Run-on Analysis of Transcription
- 3. Nuclear Extract Logeat
- 4. Nuclear Protein Extraction 1995
- 5. Nuclear Protein Extraction 1996
- 6. Nuclear Protein Extraction-small scale

# **Oligo Protocols**

- 1. Extraction and Purification of Oligo from Column
- 2. Oligonucleotide Purification Cartridge
- 3. Preparation of Adapter and Insertion into Vector
- 4. Purification of DNA Oligos from Gel
- 5. Reducing Fading of Immunofluorescene During Microscopy

#### PFLASH Phagemids

- 1. Multifunctional Cat and Luc Reporter Gene Vectors
- 2. pFLASH Phagemids Technical Manual

# Protein Protocols

- Drotein Expression Protocols
- 1. Amplification Using CHO Cell Expression Vectors
- 2. Characterization of Recombinant Vaccinia Viruses and Their Products
- 3. Generation of Recombinant Baculoviruses and Analysis of the Expressed Proteins
- 4. Generation of Recombinant Vaccinia Viruses
- 5. Overview of Protein Expression in Mammalian Cells
- 6. Overview of the Vaccinia Virus Expression System
- 7. Purification and Transfection of Baculoviral DNA for Generating Recombinant Viruses
- 8. Purification of untagged ubiquitin from bacterial cells
- 9. Transient Expression of Proteins Using COS Cells
- 10. Protein Interaction Assay by Blotting and Band-shifting Studies
- 11. Fractionation of nuclear or cytoplasm proteins
- 12. Immobilized Protein A
- 13. Pierce IgG (Protein A) Purification Kit
- 14. Protein Purification by GST Binding
- 15. Roche NiNTA protocol
- C GST-Protein Purification Protocols
- 1. Glutathione Sepharose 4B Instructions
- 1) GST purification
- 2) Preparation of Large Scale GST-proteins

#### ptTS-Neo Vector Information

- 1. Digest 1 of ptTS-Neo
- 2. Digest 2 of ptTS-Neo
- 3. ptTS-Neo Vector Information
- 4. Sequence Analysis of ptTS-Neo

#### Recipes for Solutions

1. Recipes

#### C Replicaplating Colonies onto Nitrocellulose Filters

- 1. Binding Liberated DNA Protocol
- 2. Replicating Colonies onto Nitrocellulose Filters

#### Representational Difference Analysis

- 1. A Modification of RDA
- 2. Representational Difference Analysis

# C RNA Related Protocols

- RNA Gel
- 🗁 RNA Gel Shift
- 1. E. Coli Ribonuclease III Methods
- 2. Gel Motility Shift Assay Protocol
- RNA Isolation
- 1. Controlling Ribonuclease Activity
- 2. DNA Multiprep
- 3. Electrophoresis of RNA Through Gels Containing Formaldehyde
- 4. Extraction, Purification, and Analysis of mRNA from Eukaryotic Cells
- 5. Gel Electrophoresis of RNA
- 6. Hybridization and Autoradiography
- 7. Isolation of mRNA from Mammalian Cells
- 8. Isolation of mRNA from Suspension of Cells to RNAZol B
- 9. Isolation of Total Cellular RNA
- 10. Isolation of Total RNA from Cultured Cells
- 11. Life Technology Trizol RNA and DNA Isolation
- 12. Nuclease-S1 Mapping of RNA
- 13. RNA Isolation by the RNAZol B Method
- 14. RNAZol B Isolation of RNA
- 15. Selection of Poly(A) RNA
- 16. Staining RNA After Transfer to Nitrocellulose Filters
- 17. Staining RNA Before and After Transfer to Nitrocellulose Filters
- 18. Straight A's mRNA Isolation System
- 19. Transfer of Denatured RNA to Nitrocellulose Filters
- 20. Transfer of Denatured RNA to Nylon Membranes
- 21. Ultraspec-II RNA Isolation System
- C RNA Preparation from B-Lymphocytes
- 1. RNA Preparation from B-Lymphocytes
- 🗁 RT PCR
- 1. RT PCR

# C Sequencing Protocols

- 1. Dideoxysequencing Using Sequenase
- 2. SEQUAGEL Sequencing System

# C Serum-Related Protocols

- 1. Serum Treatment for Tissue Culture
- 2. B-Cell Preparation
- 3. Isolation of Human T Cell Populations by Panning
- 4. Preparation of AET-Sheep Red Blood Cells
- 5. T-Cell Rosetting by AET-Sheep Red Blood Cells
- 6. Transfection of P3HR1
- 7. Virus Infection
- 8. Virus Preparation

# Southern Blotting

- 1. 50 mL Rehybridization
- 2. Gene Screen Plus
- 3. Nucleic Acid Transfers for BIOTRANS Nylon Membranes
- 4. PCR Analysis from Rich
- 5. Southern Blots by Lee
- 6. Stripping Probes from Blots

# SYBR Green Information

- 1. SYBR Safe DNA Gel Stain MSDS
- 2. SYBR Safe DNA Gel Stain Order Information
- 3. SYBR Safe DNA Gel Stain Product Sheet

# Taq Polymerase Preparation

- 1. Gene Choice Fact Sheet Taq DNA Polymerase
- 2. Grimm and Arbuthnot Rapid Purification of Recombinant Taq
- 3. Guidelines for Use of Robertson Lab Recombinant Taq Polymerase
- 4. Pluthero Purification of Taq
- 5. Pluthero Rapid Purification of Recombinant Taq
- 6. Preparation of Taq Polymerase
- 7. Purification of Taq Expressed in E. coli
- 8. Taq Amplification Using the pAKTaq Plasmid

# Transfection Protocols

- 1. Liposome-mediated Transfection
- 2. Transfection of DNA into Eukaryotic Cells

# Transient Transduction with Adenovirus Poly Lysine (AdpL)

- 1. Coupling of Adenovirus to Transferrin-Polylysine DNA Complexes Enhances Gene Delivery and Expression
- 2. Receptor-mediated Gene Delivery Using Defective or Inactivated Adenovirus Particles
- 3. Transient Transduction with AdpL
- 4. Virus Purification
- 5. Virus Purification on Dextrose Gradients

# C Vent DNA Polymerases

- 1. Deep Vent exo DNA Polymerase
- 2. Using Vent DNA Polymerases to Extend a Primer
- 3. Vent DNA Polymerase Technical Data Sheet

# Virus Preparation Protocols

- 1. DuPont P1 Packaging System
- 2. High Yield Method for Isolation of Lambda DNA
- 3. Lambda Packaging in PLK-A
- 4. Lentivirus Protocol
- 5. pQC-tTS-IN Retroviral Vector Information

# 🗁 Western Blotting

- 1. Detection of Proteins in Gels
- 2. Detection of Proteins on Blot Transfer Membranes
- 3. Section 4 Transfer Conditions
- 4. Section 5 Specific Protocols
- 5. Section 6 Blotting Membranes
- 6. Stripping and Reprobing Amersham
- 7. Stripping and Reprobing Membranes in Western Blotting
- 8. Western Blotting Lysis to Analysis
- 9. Western Blotting

# Yeast Protocols

- 1. Competent Yeast Cells
- 2. Direct Transfer of DNA from Yeast to E. coli by Electroporation
- 3. High Efficiency Yeast Transformation Using RNA as a Carrier
- 4. Improved Method for Transformation of Intact Yeast Cells
- 5. Izumi Yeast Transformation Transfer to E. coli
- 6. JAGGED1 and JAGGED2 Sequences
- 7. Protein Interaction Cloning.
- 8. Transformation of Yeast Cells
- 9. Yeast Small Scale Transformation