Purification of DNA Oligos from Gel

1. Resuspend oligos in H_2O or TE buffer.

2. Pour Native-PAGE at appropriate concentration for size of oligos.
   e.g. 20% Native PAGE for 15mer-30mer

   Make PAGE gel with 0.5X TBE buffer→ ddH_2O, acrylamide, TBE buffer.
   *Does not need stacking gel.

3. Load samples with loading buffer without dye. Add one lane with dye at far right or left lane.

4. Run Gel in 0.5X TBE buffer at 150-400 V COLD. Gel heats up. Make sure buffer is enough to keep gel cold. Use cooling chamber or cold room.

5. Run to approximately 3/4 of the way to the bottom of gel. You can use minigel if available.

6. Remove gel. Place on Saran Wrap. Place on TLC Fluorescence Plate. UV shadow with short wavelength light.

7. Mark dark shadows. DNA shadows with UV light.

8. Cut out band (as small of a piece as possible). Cut gel slices into many small pieces. Place in 1.5 ml eppendorf tube.

9. Add 500µl 0.3 M NaAcetate (pH 4.8-5.2). Shake O/N at 37 °C in a vortexer.

10. Spin out gel slice (Save and redo if necessary). Phenol/$CHCl_3$ extract. Precipitate with ethanol and acetate. Wash 2X with 70% ethanol to remove salt.

11. Resuspend in sterile H_2O or TE buffer. (Use approx. 105µl)

12. Spec for concentration (use 5µl/500µl dilution 1:100)

13. Use primer in experiment.