Buffers For RNA Gel

1% Agarose Formaldehyde Gel : 100mls => 1g agarose

		150mls
10X MOPS	- 10mls	- 15mls *
HCHO 37%	- 5.4mls	- 8.1mls
DEPC H_2O	- 84.6mls	- 126.9mls
	100mls	150mls

* Microwave agarose in H_2O to dissolve. Cool to 60°C then add MOPS and HCHO.

Running Buffer: 1 liter

10X MOPS	- 100mls
HCHO 37%	- 53.6mls
DEPC H_2O	- 846.4mls
	1000mls

* Use ddH2O but first run tap a few secs before collecting H_2O.

Formaldehyde Gel-Loading Buffer : 50mls

10X MOPS	- 5mls	BPB	- 0.25%
HCHO 37%	- 8.9mls	XC	- 0.25%
Glycerol	- 25mls	EtBr	- 12mg/ml (100µg)
0.5M EDTA	- 100µl		
DEPC H_2O	- 11.1mls		
	50mls		

10X MOPS Buffer: - 1 liter

400mM	MOPS Acid	- 83.7g
100mM	$NaCH_3COO$	- 13.6g
20mM	EDTA	- 40mls (0.5M)
DEPC H_2O		- to 1000mls
		1000mls

* Filter Sterilize thru' 0.2µ nitrocellulose filters.

Sample preparation : 20 µl

RNA	– 4.5 µl	* add 2.0 µl loading dye just prior to loading.
10x MOPS	– 2.0 µl	
HcHo 39%	– 3.5 µl	
$HCHONH_2$	– 10.0 µl	
	20.0 µl	

Electrophoretic Conditions

- Soak Gel apparatus in dilute bleach or absolve detergent for a few hours
- Rinse thoroughly in RO H_2O
- Dry Gel rig and place Gel in tray into apparatus in Chemical hood.
- load Gel and run at 30 volts o/N. Not necessary to recirculate buffer.
- Prior to loading Gel. Heat Samples 65°C 5-10 mins then place on ice immediately.
- Centrifuge to get Sample to bottom of eppendorf.
- load Gel