Yeast Small Scale Transformation

1. Prepare carrier DNA by heating 200ul of Herring Testes DNA (20 mg/ml) to 90-100oC for 5-10 minutes. Immediately add 800ul of LiSORB to heated Herring Testes DNA and vortex to mix. Allow carrier to cool to room temperature.
2. Add 5ug plasmid DNA to 50ul Carrier DNA. (The remaining Carrier DNA can be stored at –20oC for later use).
3. Add 50ul of competent yeast to plasmid/carrier mix. Incubate at 30oC for 30 minutes.
4. Add 450ul 40% PEG3400 and mix gently by pipetting. Incubate cells at 30oC for 30 minutes.
5. Heat shock cells for 10 minutes at 42oC.
6. Pulse spin cells for 5-10 seconds.
7. Pipet off PEG.
8. Recover yeast cells by incubating them in 1ml selective DOB at 30oC for 1 hour while shaking.
9. Pellet yeast cells with a quick spin (<5 seconds). Pipet off 800ul of the supernatant. Resuspend yeast cell in the remaining supernatant.
10. Plate ~100ul of resuspended cells on selective DOBA plates. Incubate plates inverted at 30oC.